# ROLE OF MAGNESIUM TO ALLEVIATE DAMAGING EFFECT OF SALINITY ON DEVELOPMENT AND YIELD PERFORMANCE OF RICE PLANT (*Oryza sativa* L.)

A THESIS

BY

# FAISAL MHAMUD



## DEPARTMENT OF AGRICULTURAL BOTANY

# SHER-E-BANGLA AGRICULTURAL UNIVERSITY June, 2018

# ROLE OF MAGNESIUM TO ALLEVIATE DAMAGING EFFECT OF SALINITY ON DEVELOPMENT AND YIELD PERFORMANCE OF RICE PLANT (*Oryza sativa* L.)

### BY

## FAISAL MHAMUD

Registration No.: 11-04403

A Thesis

Submitted to the Department of Agricultural Botany Sher-e-Bangla Agricultural University, Dhaka, in partial fulfillment of the requirements for the degree of

## MASTER OF SCIENCE IN AGRICULTURAL BOTANY

**SEMESTER: JANUARY-JUNE/2018** 

**Approved by:** 

Dr. Kamrun Nahar Associate Professor Department of Agricultural Botany Sher-e-Bangla Agricultural University Supervisor Dr.Md. Sazzadur Rahman

Senior Scientific Officer Plant Physiology Division Bangladesh Rice Research Institute Gazipur-1701 Co-Supervisor

**Dr. Kamrun Nahar** Associate professor Chairman Department of Agricultural Botany Sher-e-Bangla Agricultural University



Sher-e-Bangla Agricultural University

Sher-e-Bangla Nagar, Dhaka-1207

PABX: +88029144270-9

Fax: +88029112649

Ref:

Date: .....

# CERTIFICATE

This is to certify that the thesis entitled, "ROLE OF MAGNESIUM TO ALLEVIATE DAMAGING EFFECT OF SALINITY ON DEVELOPMENT AND YIELD PERFORMANCE OF RICE PLANT (Oryza sativa L.)" submitted to the Department of Agricultural Botany, Faculty of Agriculture, Sher-e-Bangla Agricultural University, Dhaka in partial fulfillment of the requirements for the degree of MASTER OF SCIENCE IN AGRICULTURAL BOTANY, embodies the results of a piece of bonafide research work carried out by FAISAL MHAMUD Registration No. 11-04403 under my supervision and guidance. No part of the thesis has been submitted for any other degree or diploma.

I further certify that such help or source of information, as has been availed of during the course of this investigation has duly been acknowledged.

Dated : June, 2018 Place: Dhaka, Bangladesh Dr. Kamrun Nahar Associate Professor Department of Agricultural Botany Sher-e-Bangla Agricultural University, Dhaka

Supervisor

## ACKNOWLEDGEMENTS

All praises are due to Almighty Allah, the Supreme Ruler of the universe who enabled the Author to complete the present piece of work.

I would like to express my heartiest respect, deep sense of gratitude, sincere and profound appreciation to my supervisor, **Associate Prof. Dr. Kamrun Nahar**, Chairman, Department of Agricultural Botany, Sher-e-Bangla Agricultural University, Dhaka for her sincere guidance, scholastic supervision, constructive criticism and constant inspiration throughout the course and in preparation of the manuscript of the thesis.

I would like to express my heartiest respect and profound appreciation to my Cosupervisor, **Dr. Md. Sazzadur Rahman**, Senior Scientific Officer, Plant Physiology Division, BRRI, Gazipur for his constructive suggestions to discuss the result and preparation of thesis.

I express my sincere respect to all the teachers of Department of Agricultural Botany, Shere-Bangla Agricultural University, Dhaka for providing the facilities to conduct the experiment and for their valuable advice and sympathetic consideration in connection with the study.

I would like to thank all of my roommates and friends especially and senior brother to help me in my research work.

Mere diction is not enough to express my profound gratitude and deepest appreciation to my father, mother, brothers, sisters, and friends for their ever ending prayer, encouragement, sacrifice and dedicated efforts to educate me to this level.

June, 2018 SAU, Dhaka The Author

# ROLE OF MAGNESIUM TO ALLEVIATE DAMAGING EFFECT OF SALINITY ON DEVELOPMENT AND YIELD PERFORMANCE OF RICE PLANT (*Oryza sativa* L.)

#### BY

## FAISAL MHAMUD

### ABSTRACT

A pot experiment was conducted at the net house of the Department of Agricultural Botany, Sher-e-Bangla Agricultural University, Dhaka-1207, during Boro rice cropping season of the year of 2017-2018 to investigate the role of magnesium to alleviate damaging effect of salt stress on development and yield performance of rice plant (Oryza sativa L.). The experiment was conducted using two salinity levels with magnesium viz.  $S_0$  = Control,  $S_1$  = 4 dSm<sup>-1</sup> salt,  $S_1Mg_1$  = 4 dSm<sup>-1</sup> salt + 80 ppm magnesium,  $S_1Mg_2 = 4 \text{ dSm}^{-1} \text{ salt} + 160 \text{ ppm magnesium}$ ,  $S_2 = 6 \text{ dSm}^{-1} \text{ salt}$ ,  $S_2Mg_1 = 1000 \text{ salt}$  $6 \text{ dSm}^{-1} \text{ salt} + 80 \text{ ppm magnesium}, \text{ S}_2\text{Mg}_2 = 6 \text{ dSm}^{-1} \text{ salt} + 160 \text{ ppm magnesium}.$  The experiment was set in Randomized Complete Block Design (RCBD) having one factor with three replications. The results on the effect of morphological and yield contributing characters indicated that plant height; numbers of tillers, leaf length, leaf breath, and SPAD value, effective tiller, number of non effective tiller, panicle length and filled grain, unfilled grain, thousand seed weight grain yield were influenced by salinity and magnesium. Among the 3 salinity level (0,4 & 6dSm<sup>-1</sup>), the damaging effect was found more in higher stress (6dSm<sup>-1</sup> salt).But magnesium supplementation along with salt greatly reduced the damaging effect of salt. Out of 2 level of magnesium, 160 ppm magnesium. Significantly reduced the damaging effect of salt. Therefore, for cultivation of BRRI dhan67 under the saline condition magnesium application could be a better practice for getting higher yield.

# CONTENTS

Chapter	Title	Page
	ACKNOWLEDGEMENT	i
	ABSTRACT	ii
	LIST OF CONTENTS	iii-iv
	LIST OF TABLES	V
	LIST OF FIGURES	vi
	LIST OF APPENDICES	vii
	LIST OF ACRONYMS	viii
1	INTRODUCTION	1
2	<b>REVIEW OF LITERATURE</b>	5
3	MATERIALS AND METHODSS	21
3.1	Experiment location	21
3.2	Soil condition	21
3.3	Climatic condition	21
3.4	Selection of cultivars	22
3.5	Experimental design	22
3.6	Treatments	22
3.7	Collection and preparation of soil	22
3.8	Sterilization of seed	23
3.9	Sowing of seeds in seed bed	23
3.10	Raising of seedlings	23
3.11	Seedling transplant in the pots	23
3.12	Collection of data	24
3.12.1	Plant height	24
3.12.2	Total tiller hill <sup>-1</sup>	24
3.12.3	Length and breadth of the leaf	24
3.12.4	SPAD value	24
3.12.5	Number of effective tillers	25
3.12.6	Number of non-effective tillers	25
3.12.7	Total dry matter	25
3.12.8	Panicle length	25
3.12.9	Number of filled grain	25
3.12.10	Number of unfilled grain	25
3.12.11	1000-seed weight	26
3.12.12	Grain yield hill <sup>-1</sup>	26
3.13	Statistical analysis	26

Chapter	Title	Page
4	<b>RESULTS AND DISCUSSION</b>	27
4.1	Plant height	27
4.2	Number of total tiller hill <sup>-1</sup>	29
4.3	Leaf length	31
4.4	Leaf breath	31
4.5	SPAD Value of leaf	31
4.6	Number of effective tillers	33
4.7	Number of non-effective tillers	35
4.8	Panicle length	35
4.9	Number of filled grain panicle <sup>-1</sup>	36
4.10	Number of unfilled grain panicle <sup>-1</sup>	36
4.11	Thousand seed weight	36
4.12	Grain yield hill <sup>-1</sup>	37
5	SUMMARY AND CONCLUSION	39
	REFERENCES	41
	APPENDICES	51

# LIST OF TABLES

Number	Title	Page	
01	Effect of salinity with magnesium on leaf length, leaf		
	breath, SPAD value and total dry mater weight on rice	32	
02	Effect of salinity with magnesium on number of effective		
	tiller and non-effective tiller hill <sup>-1</sup> on rice	34	
03	Effect of salinity with magnesium yield and yield		
	contributing character of on rice	38	

# LIST OF FIGURES

Number	Title	Page
01	Effect of different levels of salinity and magnesium on the plant height of rice at different days after transplanting	28
02	Effect of different level of salinity with magnesium on the number of tiller of rice at different Days after transplanting	30

# LIST OF APPENDICES

Appendices	Title	Page
Ι	Map showing the experimental sites under study	51

# LIST OF ABBREVIATIONS

AEZ	=	Agro-Ecological Zone
BARI	=	Bangladesh Agricultural Research Institute
BBS	=	Bangladesh Bureau of Statistics
LAI	=	Leaf area index
ppm	=	Parts per million
et al.	=	And others
Ν	=	Nitrogen
TSP	=	Triple Super Phosphate
MP	=	Muriate of Potash
RCBD	=	Randomized complete block design
DAS	=	Days after sowing
ha <sup>-1</sup>	=	Per hectare
g	=	gram (s)
Kg	=	Kilogram
μg	=	Micro gram
SAU	=	Sher-e-Bangla Agricultural University
SRDI	=	Soil Resources and Development Institute
HI	=	Harvest Index
No.	=	Number
Wt.	=	Weight
LSD	=	Least Significant Difference
${}^{0}C$	=	Degree Celsius
mm	=	Millimeter
Max	=	Maximum
Min	=	Minimum
%	=	Percent
cv.	=	Cultivar
NPK	=	Nitrogen, Phosphorus and Potassium
CV%	=	Percentage of coefficient of variance
Hr	=	Hour
Т	=	Ton
viz.	=	Videlicet (namely)

#### **CHAPTER I**

#### **INTRODUCTION**

Rice(*Oryza sativa* L.) is the staple food for more than half of the world's population. In humid and sub-humid Asia,rice is the single most important source of employment and income for rural people and it provides 50%-80% of the calories consumed for more than three billion Asians (Hossain and Fischer, 1995;Khush, 2005). However, rice is very sensitive to salinity stress and is currently listed as the most salt sensitive cereal crop with a threshold of 3 dSm<sup>-1</sup> for most cultivated varieties (USDA, 2016), whereas, generally, a soil is only considered saline (salt-affected) if it has an ECe (electrical conductivity of saturation extract)about above (Rengasamy, 2006).Even at ECe as low as 3.5 dSm<sup>-</sup>

<sup>1</sup>, rice loses about 10% of its yield, and 50% yield loss was recorded for rice at ECe 7. 2 dSm<sup>-1</sup>. Asia accounts for 90% of the world's production and consumption of rice but "sea level rise is already increasing rice growing areas of the world" (Dayton, 2014). Salinity is a general term used to describe the presence of elevated levels of different salts such as sodium chloride, magnesium and calcium sulphates and bicarbonates in soil and water (Hoang *et al.* 2015). With more than 830 million hectares (ha) of saltaffected land globally (Mannus, *et al.* 2005) and approximately two million hectare of land uncultivable due to excessive salinity added each year (Umali, 1993), salinity is a growing worldwide problem. Irrigation and extensive clearing of vegetation, which bring the groundwater with soluble salts to, or close to, the soil surface, are the two major human activities that accelerate salinity. Of the 230 million hectare of the world's irrigated land, 45 million hectare (20%) has been salt-affected (Shrivastava and Kumar, 2015). When growing on salt-affected soils, crops must compete with salts in the soils for water as well as to cope with ion toxification, nutritional disorders and poor soil physical conditions to survive, therefore, their productivity was reduced (Shrivastava and Kumar, 2015, Munns and Tester, 2005). By 2050, the world's population is predicted to reach 9.6 billion people and food production needs to increase approximately 70% by 2050 or 44 million metric tons annually to provide sufficient food for this population (FAO, 2009, 2012; United Nations Population Fund, 2016). This is a challenge because there is very little potential for future expansion of arable lands, whereas environmental stresses affecting crop production are increasing (Tester and Langridge, 2010; Eckardt, 2009, Cominelli, *et al.* 2013). Salinity and water deficit were listed as the two most critical factors that limit global crop production (Munns, 2011). To help sustain the increasing population, crops with enhanced salinity tolerance must be developed to increase productivity on salt-affected lands.

Salinity is one of the most brutal environmental stresses that adversely influenced the capability of plant to uptake water, and this quickly causes reduction in growth rate, along with a suite of metabolic changes (Munns, 2002, Vaidyanathan *et al.* 2003). It seriously limits agricultural productivity with significant crop loss through worldwide (Munns and Tester 2008). Now more than 800 million hectares of land throughout the world are salt affected which is 6% of the world's total land area (FAO 2008) and this area is increasing day by day because of global warming with consequent rise in sea level and increase in tidal surges, particularly in coastal areas in the globe (Wassmann

*et al.* 2004). In Bangladesh, about 1.06 million hectares out of 2.85 million hectares of coastal and off-shores land is affected by different degrees of salinity (Murshed, 2008).

Magnesium (Mg) is one of the essential mineral nutrients for the growth and development of plants. Apart from being a central atom of the chlorophyll molecule, Mg also acts as activator or regulator of many key enzymes in plant physiological processes (Marschner et al. 1995, Shaul, 2002). Both Mg deficiency and over supply have detrimental effects on plant photosynthesis (Shabala and Y. Hariadi, 2005) consequently resulting in abnormal or restricted growth of plants (Shaul, 2002). The cation competitive effects frequently lead to Mg deficiency in the field (Mengel and Kirkby, 1987).Mg deficiency in plants occurs worldwide influencing productivity and quality in agriculture (Hermans, 2005). Magnesium (Mg) is major plant macro-nutrient that plays important roles in stomatal movement, osmoregulation, enzyme activity, cell expansion, neutralization of non-diffusible negatively charged ions, and membrane polarization. Application of magnesium can complete with sodium ion and reduce the salt stress. Because salt stress mainly occurs due to salt toxicity in plant. Metabolic toxicity of Na is largely due to its ability to compete with Mg for binding site essential for cellular function effects of salt stress.

Therefore, it is crucial to enhance rice tolerance to salinity stress to enable this staple crop to provide enough food for rice consuming communities. Although some success has been reported for enhanced salinity stress tolerance in rice, the achievements so far are quite modest.

Therefore, the present work has been designed and planes with the following objectives:

- To study the growth, development, yield contributing characters and yield of rice plant under salt stress
- To observe the roles of Mg in alleviating adverse effect of salt stress on vegetative and yield performance of rice plant

#### **CHAPTER II**

#### **REVIEW OF LITERATURE**

Sivasankaramoorthy (2014) studied under pot experiments 2008. Results indicated that significant increases were recorded in percentage of germination, seedling fresh and dry weights, seedling length, water content, catalase activity and photosynthetic pigments (chlorophyll a, b and total chlorophylls as well as carotenoids) under the low level concentration (20 mM) of NaCl or CaCl<sub>2</sub> and their combination (1:1). On other hand increasing salt concentration in nutrient solution caused significant decrease in all of these parameters. The great reduction occurred under high salinity level of NaCl (50 mM). Meanwhile, peroxidase activity increased significantly with increasing salinity levels from 20 mM to 50 mM of both applied salinity types. Besides, peroxidase activity under NaCl salinity showed a marked increase followed by NaCl + CaCl<sub>2</sub> (1:1) and CaCl<sub>2</sub> at 50 mM.

The successful establishment of crop mainly depends on good quality seed which germinate rapidly, uniformly and able to withstand under environmental adversity after sowing. Laboratory experiment was conducted by Yousof (2013) at Seed Technology Unit, Mansoura, Egypt to evaluate the effect of rice seed priming (c.v. Giza 177) with CaCl2 at osmotic potentials (- 0.75, -1.00, -1.25, -1.50 MPa and distilled water) for 6, 12, 24, 36 and 48 hours on seed germination, seed and seedling vigor under normal and salinity stress. The obtained results of this study indicated that salinity levels (9 dSm<sup>-1</sup>) delayed germination, seed and seedling vigor characters compared with normal salinity (0.3 dSm<sup>-1</sup>). Rice seed priming with CaCl<sub>2</sub> at (-1.00

MPa) followed by (-1.25 MPa) gave the highest values of germination percentage, speed of germination index, germination rate, germination co-efficient, germination energy %,seedlings length, seedlings dry weight, water uptake % and decreasing mean germination time as well as time to 50 % germination compared with control (distilled water) under salinity stress. Whereas, priming seed with distilled water (control) resulted in the lowest germination characters under salinity stress. Priming duration 24 h showed its superiority in improving germination characters comparing with other priming durations. It could be concluded that, priming rice seed (c.v. Giza 177) with osmotic potential (-1.00 MPa) for 24 h can reduce the injurious effects of salinity stress.

Wu and Wang (2012) conducted to investigate the effects of  $Ca^{2+}$  on cation accumulation and K<sup>+</sup>/Na<sup>+</sup> selectivity, in this study, two-week-old rice (*Oryza sativa* L.) plants were exposed to 25 or 125 mmolL<sup>-1</sup> NaCl with or without 10 mmolL<sup>-1</sup>  $CaCl_2$ . At low salinity (25 mmolL<sup>-1</sup> NaCl),  $Ca^{2+}$  significantly decreased Na<sup>+</sup> accumulation in roots, increased K<sup>+</sup> accumulation in shoots, and maintained higher K<sup>+</sup>/Na<sup>+</sup> ratios in both roots and shoots of rice plants. At high salinity (125 mmolL<sup>-1</sup> NaCl), however,  $Ca^{2+}$  did not have any effects on Na<sup>+</sup>, K<sup>+</sup> accumulation and K<sup>+</sup>/Na<sup>+</sup> ratios in plants. Further analysis showed that, at low salinity, the addition of  $Ca^{2+}$ significantly enhanced the selective absorption and transport capacity for K<sup>+</sup> over Na<sup>+</sup> in rice. Although Na<sup>+</sup> efflux and Na<sup>+</sup> influx was remarkably reduced by  $Ca^{2+}$ under both low and high salt stresses, their ratio was lowered only under low salt stress. In summary, these results suggest that  $Ca^{2+}$  could regulate K<sup>+</sup>/Na<sup>+</sup> homeostasis in rice at low salinity by enhancing the selectivity for  $K^+$  over  $Na^+$ , reducing the  $Na^+$  influx and efflux, and lowering the futile cycling of  $Na^+$ .

Suriyan Cha-um et al. (2012) evaluate its role in regulating physiological and biochemical process in response to salt-induced stress. Two rice genotypes, Pokkali salt tolerant and IR29 salt susceptible, grown on liquid Murashige and Skoog medium (MS) supplied by 1.98 mM CaCl<sub>2</sub> (control) were compared to 2 (3.96 mM), 4 (7.92 mM) and 8 (15.84 mM) folds exogenous CaCl<sub>2</sub> pretreatment subsequently exposed to 200 mM NaCl salt stress. Thus, the present investigation evaluated the potential of exogenous calcium chloride (CaCl<sub>2</sub>) supply in improving the growth performance and photosynthetic ability in salt stressed rice. In IR29 salt susceptible rice, leaf area of salt-stressed seedling was significantly recovered by exogenous application of 7.92 mM CaCl<sub>2</sub>, which was greater by 1.38-folds over that in 1.98 mM CaCl<sub>2</sub> application. Exogenous CaCl<sub>2</sub> (7.92 mM) enhanced proline accumulation in both Pokkali (3.26 mmol g-1FW) and IR29 (4.37 mol g-1FW) genotypes, and reduced relative electrolyte leakage thereby indicating its positive role in membrane stability. Treatment of 7.92 mM CaCl<sub>2</sub> significantly enhanced the photosynthetic abilities, including maximum quantum yield of PSII (Fv/Fm), photon yield of PSII (PSII), photochemical quenching (qP) and net photosynthetic rate (Pn), in two genotypes of salt-stressed rice seedlings, especially in salt susceptible IR29 genotypes. The study concludes that an exogenous application of 7.92mM CaCl<sub>2</sub> significantly enhanced the photosynthetic abilities and overall growth performances in the photoautotrophic growth of salt-stressed rice seedlings. Exogenous calcium in the culture media may absorb by root tissues, transfer to whole plant and function as salt defense mechanisms including calcium signaling in the abscisic acid (ABA) regulation system and calcium sensing in stomatal closure when plant subjected to salt stress.

Kazemi and Eskandari (2011) carried out in 2009, in order to investigate the effects of salt stress ( $S_1$ = 0.0,  $S_2$ = 2.0,  $S_3$ = 4.0,  $S_4$ = 6.0 and  $S_5$ = 8.0 ds.m<sup>-2</sup> Nacl) on germination and seedling properties of three rice cultivars (Anbar, LD and Hamar). Experiment was arranged as split-plot based on randomized complete block design in three replications, with the salinity levels in main plots and rice cultivars in subplots. The results indicate that all traits were significantly (P≤0.05) affected by salt stress, where germination, plumule and radicle length and weight were decreased with increasing in salt concentration. The extent of these reductions was related with the variations rice cultivar under different salt stress condition. By increasing NaCl concentration, seed germination delayed and decreased in all cultivars. Regarding the relationship between speed of germination and seed vigor, salt stress which can be suggested for cultivation under salinity condition.

Ashraf *et al.* (2010) conducted a hydroponics experiment to evaluate the role of potassium (K) and silicon (Si) in mitigating the deleterious effects of NaCl on rice cultivars differing in salt tolerance. Two salt-sensitive (CPF 243 and SPF 213) and two salt-tolerant (HSF 240 and CP 77-400) rice cultivars were grown for six weeks in <sup>1</sup>/<sub>2</sub> strength Johnson's nutrient solution. The nutrient solution was salinized by two

NaCl levels (0 and 100 mmol  $L^{-1}$  NaCl) and supplied with two levels of K (0 and 3 mmol  $L^{-1}$ ) and Si (0 and 2 mmol  $L^{-1}$ ). Applied NaCl enhanced Na<sup>+</sup> concentration in plant tissues and significantly ( $P \le 0.05$ ) reduced shoot and root dry matter in four rice cultivars. However, the magnitude of reduction was much greater in salt-sensitive cultivars than salt-tolerant cultivars. The salts interfered with the absorption of K<sup>+</sup> and Ca<sup>2+</sup> and significantly ( $P \le 0.05$ ) decreased their uptake in rice cultivars. Addition of K and Si either alone or in combination significantly ( $P \le 0.05$ ) inhibited the uptake and transport of Na<sup>+</sup> from roots to shoots and improved dry matter yields under NaCl conditions. Potassium uptake, K<sup>+</sup>/Na<sup>+</sup> ratios, and Ca<sup>2+</sup> and Si uptake were also significantly ( $P \le 0.05$ ) increased by the addition of K and/or Si to the root medium. In this study, K and Si-enhanced salt tolerance in rice cultivars was ascribed to decreased Na<sup>+</sup> concentration and increased K<sup>+</sup> with a resultant improvement in K<sup>+</sup>/Na<sup>+</sup> ratio, which is a good indicator to assess plant tolerance to salt stress.

Zafar *et al.* (2004) investigated the response of rice cultivars Basmati-370 (saltsensitive) and IR6 (salt-tolerant) to 2 salinity levels (4.0 (control) and 10 dSm<sup>-1</sup>) in a pot experiment in a wire-house. They took four harvests at an interval of 10 days each after imposition of salinity treatment, and growth and chemical analyses of plant samples were carried out. Plant biomass showed an inverse relationship with increasing salinity levels. A general trend of decrease in dry weight of plant with salinity was noted in both cultivars. The mean values for dry weight were higher in Basmati-370 in the control condition. Analysis of variance showed a significant increase in Na<sup>+</sup> and Cl<sup>-</sup> uptake with increasing salinity. Varietal means were highly significant and the maximum increase in Na<sup>+</sup> uptake (18.69%) was recorded in Basmati-370. Harvest means showed that Na<sup>+</sup> uptake increased with the passage of time. However, at maturity there was a decline in Na<sup>+</sup> content in both cultivars. Cl<sup>-</sup> increased with increasing salinity levels. Cultivar x treatment interaction revealed an increase in Na<sup>+</sup> and Cl<sup>-</sup> uptake over the control in both cultivars. However, it was less in IR-6. The cultivars differed significantly for K<sup>+</sup>, Ca<sup>2+</sup>, P and N uptake. K<sup>+</sup> and Ca<sup>2+</sup> uptake increased with the passage of time. Basmati-370 and IR-6 showed 45.20 and 15.55% decrease in Ca<sup>2+</sup> over the control. P and N uptake increased with increasing salinity levels. An increase of 23.21% P uptake was recorded in Basmati-370 compared to IR-6. However, IR-6 accumulated higher (22.16%) N compared to Basmati-370 under the control and saline conditions.

Salt stress can be ascribed to two salts calcium salts and sodium salts, although most of the salt stresses in nature are due to Na salts, particularly NaCl. Salinity effects can be classified as osmotic, toxic or nutritional. Salt stress causing toxicity could be termed primary salt injury and that causing osmotic stress and nutritional stress (including deficiency of other nutrients) is secondary salt-induced stress (Manneh, 2004). Choi *et al.* (2003) observed that the plant height decreased in the 0.5% saline water in the soil. Khan *et al.* (1997) conducting a pot experiment with three rice 2005cultivars reported that plant height was seriously decreased by salinity. Similar opinion was also postulated by Saleque *et al.* (2005). During vegetative period, the most common salinity effect was stunting of plant growth, whereas leaf withering was less apparent (Alam *et al.*, 2001). The mutant variety maintained its superiority in various

characteristics such as plant height, higher number of fertile panicles per plant and high plant yield (Baloch *et al.*, 2003).

Baba and Fujiyama (2003) investigated short-term (72 h) responses of the water and nutritional status to Na-salinization in rice (Oryza sativa L. cv. Koshihikari) and tomato (Lycopersicon esculentum Mill cv. Saturn) using pot experiments. The shortterm effect of supplemental K and Ca to the nutrient solution on the water status and absorption and transport of ions in the plants was also investigated. In both species, Na salinity resulted in the deterioration of the water status of tops and in nutritional imbalance. However, in rice, it was possible to prevent the deterioration of the nutrient status by enhancing the transport of cations, especially K, while tomato could maintain an adequate water status by inhibiting the water loss associated with transpiration. On the other hand, the water status in rice and the nutritional status in tomato markedly deteriorated by high Na level in the solution. Supplemental K and Ca could not ameliorate the water status in both species, and even worsened the status in rice. In rice, a close relationship was observed between the osmotic potential (OP) of the solution, water uptake and water content. The water status of rice, therefore, seemed to depend on OP of the solution. Supplemental K and Ca, on the other hand, were effective in the amelioration of the nutritional status. In tomato, supplemental Ca could improve the nutritional balance by suppressing the transport of Na and enhancing that of the other cations in avoidably the deterioration of the water status. Thus, the differences in the responses of the water and nutritional status of rice and tomato to high Na salinization and to supplemental K and Ca were evident in a shortterm study and supported a similar tendency observed in a long-term study.

Many scientists have suggested that selection is more convenient and practicable if the plant species possesses distinctive indicators of salt tolerance at the whole plant, tissue or cellular level (Ashraf, 2002; Epstein and Rains, 1987; Jacoby, 1999; Munns, 2002). Physiological criteria are able to supply more objective information than agronomic parameters or visual assessment while screening for component traits of complex characters (Yeo, 1994). There are no well-defined plant indicators for salinity tolerance that could practically be used by plant breeders for improvement of salinity tolerance in a number of important agricultural crops. This is partly due to the fact that the mechanism of salt tolerance is so complex that variation occurs not only among the species but, in many cases, also among cultivars within a single species (Ashraf, 1994a; 2002). During the course of plant growth, the form and functions of various organs undergo significant change and the ability of the plant to react to salinity stress depend on those genes that are expressed at the stage of development during which the stress is imposed (Epstein and Rains, 1987). The mechanism of salinity tolerance becomes even more complicated when the response of a plant also varies with the concentration of saline medium and the environmental conditions in which the plant is grown.

Din *et al.* (2001) used artificially salinized soils to see the effect of foliar and soil application of K on rice. Results indicated that the number of tillers plant<sup>-1</sup>, paddy and straw yield and grains to straw ratio significantly decreased with the increase in salinity. All K application methods increased the above parameters significantly at all salinity levels over distilled water spray. Increasing levels of salinity decreased K concentration in shoots and straw, which was increased significantly by foliar and soil

application. Both methods of K application remained at par with each other. Sodium concentration increased with increase in salinity in both shoots and straw and decreased by foliar and soil application of K. Foliar application of K proved better than soil application in this respect. The K/Na ratio decreased significantly by the increase of salinity, while this ratio increased significantly by the foliar and soil application of K.

Abdullah et al. (2001) performed an experiment on the effect of salinity stress (50 mM) on floral characteristics, yield components, and biochemical and physiological attributes of the sensitive rice variety IR-28. The results showed significant decrease in panicle weight, panicle length, primary branches per panicle, filled and unfilled grain, total grains and grain weight per panicle, 1000-grain weight and total grain weight per hill. They further observed significant reduction in both chlorophyll a and chlorophyll b content in different parts of the rice leaves at saline condition. In another experiment, Abdullah et al. (2002) studied the effect of salinity on photosynthate translocation in panicle branches and developing spikelets, carbohydrate content of different vegetative parts and suggested that reduction in grain number and grain weight in salinized panicles was not merely due to reduction in pollen viability and higher accumulation of Na<sup>+</sup> and less K<sup>+</sup> in different floral parts but also due to higher accumulation of photosynthates (sugar) in primary and secondary panicle branches, panicle main stalk and panicle stem coupled with reduced activity of starch synthetase in developing grains.

Muhling and Lauchli (2001) observed that Na<sup>+</sup> accumulation in leaves, particularly in the leaf apoplast, could be responsible for Na<sup>+</sup> toxicity in maize leaves. Lower Na<sup>+</sup>

concentrations were found in leaves of a more salt tolerant maize cultivar (Pioneer 3769) compared to a salt-sensitive maize cultivar (Pioneer 3751). They also found that the Na<sup>+</sup> concentration in the leaf apoplast of Pioneer 3751 significantly increased with higher Na<sup>+</sup> supply. They concluded that the Na<sup>+</sup> concentration in the leaf apoplast was not high enough to be responsible for the decline in leaf growth. The K<sup>+</sup> and Ca<sup>2+</sup> concentrations in whole leaves decreased with salt treatment, while K<sup>+</sup> increased and Ca<sup>2+</sup> remained constant in leaf apoplasts under salt stress.

Aslam et al. (2001) conducted in solution and soil cultures and in naturally saltaffected field. In the case of solution culture, Ca at the rate of 5, 10, 20, 40, 80 and 160  $\mu$ g Ca mL<sup>-1</sup> was applied in the presence (80 mol m<sup>-3</sup>) and absence (0 mol m-3) of NaCl salinity; whereas, in case of soil culture, Ca at the rate of 0, 50, 100 and 200 kg ha-1was applied to artificially prepared saline (ECe 9 dSm<sup>-1</sup>, SAR 5.46, pHs7.8), saline sodic (ECe9 dSm<sup>-1</sup>, SAR 28.2, pHs 8.2) soils and in naturally salt affected field (ECe 6 dSm<sup>-1</sup>, SAR 16.1, pHs 8.2). Three cultivars of differential salinity tolerance used to investigate the ameliorative nutritional aspects of Ca were: KS-282 (salt tolerant), BG 402-4 (mixed behavior) and IR-28 (salt sensitive). Application of Caimproved all growth characteristics (tillering capacity, shoot and root lengths, shoot and root weights) because of external Ca supply @ 20-40 µg Ca mL<sup>-1</sup> in solution culture in the presence of NaCl salinity. Shoot Na<sup>+</sup> and Cl<sup>-</sup> decreased; whereas, K<sup>+</sup> concentration and  $K^+$ : Na<sup>+</sup> ratio improved because of Ca supply to saline medium. Paddy and straw yields, plant height and panicle length were significantly higher in saline as compared to saline sodic soil. Application of 200 kg Ca ha-1proved statistically superior to control in respect of panicle length, number of tillers, paddy and straw yield under both saline and saline sodic soils as well as in naturally salt affected field. The ameliorative effect of Ca was due to reduced shoot Na+ and Cl<sup>-</sup> concentration and better ratio of  $K^+$  to Na<sup>+</sup> in shoot. Seed setting was improved in all the three cultivars because of external Ca supply to saline and saline sodic soils.

Zeng and Shannon (2000)studied on salinity effects on seedling growth and yield components of rice. They used cultivar M-202 rice and irrigated with nutrient solutions of control and treatments amended with NaCl and CaCl<sub>2</sub> (2:1 molar concentration) at 1.9, 3.4, 4.5, 6.1, 7.9, and 11.5 dSm<sup>-1</sup> electrical conductivity. They found seedling growth was significantly reduced by salinity at the lowest salinity treatment, 1.9 dSm<sup>-1</sup>. At 1.9 and 3.4 dSm<sup>-1</sup>, significant reduction of seedling growth occurred at longer cumulative thermal time than at higher salt levels. Seedling survival was significantly reduced when salinity was 3.40 dSm<sup>-1</sup> and higher. Tiller number per plant and spikelet number per panicle contributed the most variation in grain weight per plant under salinity. Reductions in seedling survival, tiller number per plant, and spikelet number per panicle were the major causes of yield loss in M-202 under salinity.

Most of Bangladesh's coastal region lies on the southwest coastal region of the country. Approximately 30% of the crops land of Bangladesh is located in this region (Mondal et al., 2001) and continuous to support crops productivity and GDP growth. But in the recent past, the contribution of crops to GDP has decreased because of salinity. In total, 52.8% of the cultivable land in the coastal region of Bangladesh was affected by salinity in 1990 (Karim et al., 1990) and the salt affected area has increased by 14600 ha per year (SRDI,

15

2001). SRDI had made a comparative study of the salt affected area between 1973 to 2009 and showed that about 0.223 million ha (26.7%) of new land has been affected by varying degrees of salinity during the last four decades and that has badly hampered the agro-biodiversity (SRDI, 2010). Farmers mostly cultivate low yielding, traditional rice varieties. Most of the land kept fallow in the summer or pre-monsoon hot season (March-early June) and autumn or post-monsoon season (October- February) because of soil salinity, lack of god quality irrigation water and late draining condition. In the recent past, with the changing degree of salinity of southwest coastal region of Bangladesh, crop production becomes very risky and crop yields, cropping intensity, production levels of crop and people's quality of livelihood are much lower than that in the other parts of the country. Cropping intensity in saline area of Bangladesh is relatively low, mostly 170% ranging from 62% in Chittagong coastal region to 114% in Patuakhali coastal region (FAO, 2007).

In most of the cases, the negative effects of salinity have been attributed to increase in Na<sup>+</sup> and Cl<sup>-</sup> ions in different plants hence these ions produce the critical conditions for plant survival by intercepting different plant mechanisms. Although both  $Na^+$  and  $Cl^-$  are the major ions produce many physiological disorders in plant, Cl<sup>-</sup> is the most dangerous (Tavakkoli et al., 2010). Salinity at higher levels causes both hyper ionic and hyper osmotic stress and can lead to plant demise. The outcome of these effects may cause membrane damage, nutrient imbalance, altered levels of growth regulators, enzymatic inhibition and metabolic dysfunction, including photosynthesis

16

which ultimately leading to plant death (Mahajan and Tuteja, 2005; Hasanuzzaman *et al.*, 2012).

The available literature revealed the effects of salinity on the seed germination of various crops like Oryza sativa (Xu et al., 2011), Triticum aestivum (Akbarimoghaddam et al., 2011), Zea mays, Brassica spp. and Helianthus annuus It is well established that salt stress has negative correlation with seed germination and vigor. Higher level of salt stress inhibits the germination of seeds while lower level of salinity induces a state of dormancy.

Franco *et al.* (1999) studied the effect of supplemental  $CaCl_2$  on growth and osmoregulation in NaCl stressed cowpea seedlings. They found that salinity inhibited the length of root and shoot of cowpea but the inhibitory effect could be ameliorated by the addition of  $Ca^{2+}$ . The concentration of organic osmoregulators (proline, soluble carbohydrates, soluble amino-nitrogen, and soluble proteins) increased in root tips of seedlings grown in salt-stressed condition with supplemental Ca. They indicated that  $Ca^{2+}$  could have a protective effect in root tips, which is of fundamental importance for the maintenance of root elongation in NaCl stressed cowpea seedlings.

Salinity affected rice during pollination, decreased seed setting and grain yield (Maloo, 1993). Finck (1977) suggested that deficiency of K and Ca elements might play a significant role in plant growth depression in many saline soils. Girdhar (1988)

observed that salinity delayed germination, but did not affect the final germination up to the EC of 8 dSm<sup>-1</sup> by evaluating the performance of rice under saline water irrigation. In normal conditions, the Na<sup>+</sup> concentration in the cytoplasm of plant cells was low in comparison to the K<sup>+</sup> content, frequently  $10^{-2}$  versus  $10^{-1}$  and even in conditions of toxicity, most of the cellular Na<sup>+</sup> content was confined into the vacuole (Apse *et al.*, 1999).

Bohra and Doerffling (1993) grew a salt-tolerant (Pokkali) and a salt-sensitive (IR28) variety of rice (*Oryza sativa* L.) in a phytotron to investigate the effect of K (0, 25, 50 and 75 mg K kg<sup>-1</sup> soil) application on their salt tolerance. Potassium application significantly increased potential photosynthetic activity (Rfd value), percentage of filled spikelets, yield and K concentration in straw. At the same time, it also significantly reduced Na and Mg concentrations and consequently improved the K/Na, K/Mg and K/Ca ratios. IR28 responded better to K application than Pokkali. Split application of K failed to exert any beneficial effect over basal application.

Aslam *et al.* (1993) observed significant reduction in shoot and root fresh weights by different types of salinity such as NaCl alone, NaCl + CaCl<sub>2</sub>, Na<sub>2</sub>CO<sub>3</sub> alone and a salts mixture. On the plant growth, NaCl alone was found to be the most toxic, Na<sub>2</sub>CO<sub>3</sub> alone was the least harmful, and NaCl + CaCl<sub>2</sub> and the salts mixture were intermediate. They found similar results in both solution culture experiment and the experiments conducted in salinized soils. They considered the better root growth under high salinity condition as the capacity of the tolerant genotypes to combat the adverse effect of salinity. Aslam *et al.* (2003) investigated the effect of supplemental

Ca on rice growth and yield in solution and soil cultures, and in naturally salt affected field. In solution culture, Ca was applied at 5, 10, 20, 40, 80 and 160 µg/mL with 80 mM NaCl and without NaCl and in soil culture 0, 50, 100 and 200 kg Ca ha<sup>-1</sup> was applied to artificially prepared salinity (EC 9 dSm<sup>-1</sup>). Three cultivars, differing in salt tolerance, were used, namely K8-282 (salt tolerant), BG 402-4 (moderately tolerant) and IR-28 (salt sensitive). Application of Ca at 20-40 µg/ml improved tillering capacity, shoot and root length, shoot and root weights in solution culture in the presence of NaCl. Shoot  $Na^+$  and  $Cl^-$  decreased, whereas  $K^+$  concentration and  $K^+/Na^+$ ratio increased because of Ca supply to saline medium. Grain and straw yields, plant height and panicle length were significantly higher in saline compared to saline sodic soil. Application of 200 kg Ca ha<sup>-1</sup> proved statistically superior to the control in respect of panicle length, numbers of tillers, grain and straw yields under both saline and saline sodic soil as well as in naturally salt-affected field. Seed setting was improved in all cultivars because of external Ca supply to saline and saline sodic soils. Aslam *et al.* (2003) stated that an increase in potassium and  $K^+/Na^+$  ratio was an indication of salt tolerance due to the application of additional Ca in both salt tolerant and susceptible rice cultivars under saline environment. These authors maintained that salt affected soils showed an improvement in the paddy yield of both salt tolerant and salt sensitive rice cultivars due to Ca application as gypsum at the rate of 25% of gypsum requirement of soil.

Gypsum (CaSO<sub>4</sub>, 2H<sub>2</sub>O) is widely used for ameliorating saline/sodic soils due to its tendency of replacing its Ca<sup>2+</sup> with exchangeable Na<sup>+</sup> on the soil complex. In addition, gypsum application to saline/sodic soils improve yield of paddy and forage grasses in

arid and semi arid regions due to the effects of  $Ca^{2+}$  on plant composition such as decrease in the concentration of Na and improve plant-tissue concentrations of P, K, Zn, Cu, Mg and K:Na ratio (Rengel, 1992). The addition of supplemental Ca to the root environment was a means of enhancing plant tolerance to salt stress (Epstein, 1998). This might favour the increase of  $Na^+$  inside the cells, change enzyme activity resulting in cell metabolical alterations; disturbance in K<sup>+</sup> uptake and partitioning in the cells, and throughout the plant that might even affect stomatal opening, thereby, impairing the ability of the plant to grow. This author assumed that the addition of Ca<sup>2+</sup> to the root environment of salt stressed plants would maintain or enhance the selective absorption of  $K^+$  at high  $Na^+$  concentrations and prevent the deleterious effects of the excess of Na<sup>+</sup>. Another role attributed to supplemental Ca<sup>2+</sup> addition was its help in osmotic adjustment and growth via the enhancement of compatible organic solutes accumulation (Girija et al., 2002). Under salt stress conditions there was a decrease in the Ca/Na ratio in the root environment which affected membrane properties, due to displacement of membrane-associated Ca<sup>2+</sup> by Na<sup>+</sup>, leading to a disruption of membrane integrity and selectivity (Cramer et al., 1985; Kinraide, 1998).

#### CHAPTER III

#### **MATERIALS AND METHODS**

This chapter presents a brief description about experiment period, site description, climate condition, crop or planting materials, treatments, experimental design and layout, crop growing procedure, fertilizer application, intercultural operation, data collection and statistical analysis.

#### **3.1 Experiment location**

The experiment was conducted at the experimental shed of the Department of Agricultural Botany, Sher-e-Bangla Agricultural University, Dhaka during the period from December, 2017-April, 2018

### **3.2 Soil condition**

The soil of experimental area situated to the Modhupur Tract (UNDP, 1988) under the AEZ no. 28 and Tejgoan soil series (FAO, 1988). The soil was sandy loam in texture with pH 5.47 - 5.63. The physical and chemical characteristics of the soil have been presented in Appendix I.

#### **3.3 Climatic condition**

The experimental area was under the sub-tropical climate that is characterized by less rainfall associated with moderately low temperature during Boro season, (October-March) and high temperature, high humidity and heavy rainfall with occasional gusty winds during Aus season (April-September).

#### 3.4 Selection of cultivars

BRRI dhan67 is a salt tolerant variety which was, used as the test crop in this experiment. It can tolerant salinity 12-14 dS/m (upto 3 weeks) during seedlings stage and 8 dSm<sup>-1</sup> during maturity stage.

### 3.5 Experimental design

The experiment was set in Randomized Complete Block Design (RCBD) having one factor with three replications. The treatment combination of the experiment was assigned at random into 21 pots of each at 3 replications.

### 3.6 Treatments

The experiment consisted of one factors viz, Salinity level and Magnesium Concentration

- 1.  $S_0 = Control$
- 2.  $S_1 = 4 \text{ dSm}^{-1} \text{ salt}$
- 3.  $S_1Mg_1 = 4 dSm^{-1} salt + 80 ppm Mg$
- 4.  $S_1Mg_2 = 4 dSm^{-1} salt + 160 ppm Mg$
- 5.  $S_2 = 6 dSm^{-1} salt$
- 6.  $S_2Mg_1 = 6 dSm^{-1} salt + 80 ppm Mg$
- 7.  $S_2Mg_2 = 6 dSm^{-1} salt + 160 ppm Mg$

### 3.7 Collection and preparation of soil

The soils of the experiments were collected from Sher-e-Bangla Agricultural University (SAU) farm. The soil was non-calcarious Red Brown Terrace soil with loamy texture belonging to the AEZ Madhupur Tract. The collected soil was pulverized and inert materials, visible insect pest and plant propagules were removed. The soil was dried in the sun, crushed carefully and thoroughly mixed.

#### 3.8 Sterilization of seed

Prior to germination seeds were surface sterilized with 1% sodium hypochlorite solution. The glass vials containing distilled water for seed rinsing was sterilized for 20 minutes.

#### 3.9 Sowing of seeds in seed bed

The sterilized seed were soaked with water for 24 hours, washed thoroughly in clean water, and incubated for sprouting, which were sown in the wet seed bed. Required amount of fertilizers were applied one day before sowing seeds in the seed bed.

### 3.10 Raising of seedlings

The seedlings were grown in pots and the soil was used as growth medium. Chemical fertilizers namely Urea, Triple Supper Phosphate (TSP) and Muriate of Potash (MoP) were used for N, P and K at the rate of 120, 100 and 75 kgha<sup>-1</sup> respectively before final preparation of the seed bed. The fertilizers were applied one day before sowing seeds in the seed bed. Sterilized seeds were imbibed in distilled water for 24 hours and then washed thoroughly in fresh water, and the seeds were incubated for sprouting. After sprouting, they were placed in the pots.

#### **3.11** Seedling transplant in the pots:

The chemical fertilizers *i.e.*, Urea, Triple Supper Phosphate (TSP) ,Murate of potash (MoP) and Gypsum were added for N, P, K and S in all the pot soils at the rate of 100 kg N, 60 kg  $P_2O_5$ , and 20 kg S ha<sup>-1</sup>, respectively. Magnesium Sulphate (MgSO<sub>4</sub>) used as per treatment. The whole amount of TSP, MOP, Gypsum and  $1/3^{rd}$  of urea were applied before the final preparation of the pots. Thereafter the pots containing soil were moistened with water. Five weeks old seedlings of selected rice cultivars were

transplanted in the respective pots. There were two hills in each pot. Two weeks after transplanting the salt solutions were applied in each pot according to the treatments. To avoid osmotic shock, salt solutions were added in three equal installments on alternate days until the expected conductivity was reached. The electric conductivity (EC) of each pot was measured everyday with a EC meter and necessary adjustments were made by adding water. The remaining 2/3<sup>rd</sup> urea were top dressed at two equal divisions after 25 and 50 days of transplanting.

### 3.12 Collection of data

#### 3.12.1 Plant height

The plant height (cm) was measured at different day after transplanting from the surface level of the growth media to the tip of the longest leaf at 20,35,50 and 65 days after transplanting by taking the average value of ten random samples. But the final height was measured from soil surface to the tip of the spikelet of longest panicle.

### **3.12.2** Total tiller hill<sup>-1</sup>

Total tiller number hill<sup>-1</sup> was counted at different days after transplanting.

### 3.12.3 Length and breadth of leaf

Length and breadth of leaf of each sample plant was recorded and sum total of them was divided by the total number of leaves of the sample plant.

#### 3.12.4 SPAD value

Three leaves were randomly selected from each plant. The top and bottom of each leaves were measured with at LEAF as at LEAF value. Then it was averaged and total

chlorophyll content was measured by the conversion of at LEAF value into SPAD units.

## 3.12.5 Number of effective tillers

Effective tiller number hill<sup>-1</sup> was counted at harvesting. There were two hills in each pot. The effective tiller number hill<sup>-1</sup> was counted from the pot.

## 3.12.6 Number of non effective tillers

Number of sterile tillers was also counted by subtracting the number of effective tillers from the total tiller number hill<sup>-1</sup>.

## 3.12.7 Total dry matter

The total dry matter was recorded by drying the straw at  $80 \pm 2^{\circ}$ C for 48 hours and calculated from summation of leaves, stem, roots and panicle weight as observed in an electronic balance.

## 3.12.8 Panicle length

Average panicle length (cm) was calculated by taking the lengths of all the panicles hill<sup>-1</sup>.

## 3.12.9 Number of filled grain

Average number of filled grain panicle<sup>-1</sup> was calculated by counting the number of

filled grain of 5 panicles hill<sup>-1</sup>.

## 3.12.10 Number of unfilled grain

Number of unfilled grain panicle<sup>-1</sup> was also counted.

## 3.12.11 1000 grain weight

1000-seed were counted, which were taken from the seed sample of each plot separately, then weighed in an electrical balance and data were recorded.

### 3.12.12 Grain yield

The grain yield of each hill which had effective tiller was recorded.

## 3.13 Statistical analysis

The collected data were analyzed statistically following CRD design by MSTAT-C computer package program developed by Russel (1986). The treatment means were compared by Least Significant Test (LSD) and regression analysis were performed as and where necessary.

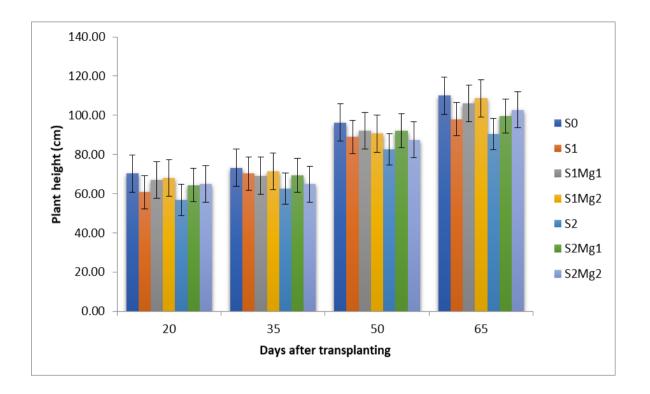
#### **CHAPTER IV**

#### **RESULTS AND DISCUSSION**

Result obtained from the present study have been presented and discussed in this chapter. The data have been presented in different tables and figures. The results have been presented and discussed, and possible interpretations are given under the following headings.

### 4.1 Plant height

The height of the plant was significantly influenced by salinity and magnesium at 20, 35, 50 and 65 DAT (Days after Transplanting) (fig. 1). The highest plant height (70.27, 73.13, 96.27, 110 cm at 20, 35, 50 and 65 DAT, respectively) was observed at control (S<sub>0</sub>) treatment, which was statistically similar with S<sub>1</sub>Mg<sub>2</sub> (67.97, 71.43, 90.70 and 108.70 cm at 20, 35, 50 and 65 DAT, respectively) and the lowest plant height (56.80, 62.50, 62.50, 82.73 and 90.33 cm at 20, 35, 50 and 65 DAT, respectively) was observed at S<sub>2</sub> (6 dSm<sup>-1</sup> salt) treatment. Choi *et al.* (2003) observed that the plant height decreased in the 0.5% saline water in the soil. Khan *et al.* (1997) conducting a pot experiment with three rice cultivars reported that plant height was seriously decreased by salinity. Similar opinion was also postulated by Saleque *et al.* (2005).



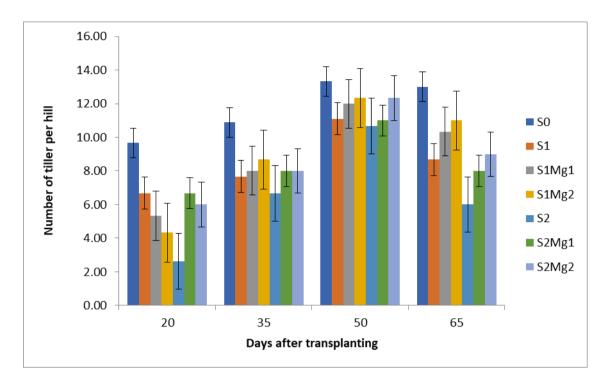
$$\begin{split} S_0 &= \text{Control}, \ S_1 &= 4 \ dSm^{-1} \ \text{salt}, \ S_1 M g_1 &= 4 \ dSm^{-1} \ \text{salt} + 80 \ \text{ppm} \ M g_{\,,} \ S_1 M g_2 &= 4 \ dSm^{-1} \ \text{salt} + 160 \ \text{ppm} \ M g, \ S_2 &= 6 \ dSm^{-1} \ \text{salt}, \ S_2 M g_1 &= 6 \ dSm^{-1} \ \text{salt} + 80 \ \text{ppm} \ M g, \ S_2 M g_2 &= 6 \ dSm^{-1} \ \text{salt} + 160 \ \text{ppm} \ M g \end{split}$$

Fig.1. Effect of different levels of salinity and magnesium on the plant height of

rice at different days after transplanting

# 4. 2 Number of total tiller hill<sup>-1</sup>

Total number of tiller hill<sup>-1</sup> was statistically influenced by different salinity level with magnesium at 20, 35, 50 and 65 DAT (Figure2). The maximum number of tiller hill<sup>-1</sup> (9.67, 10.89, 13.33 and 13.00 at 20, 35, 50 and 65 DAT, respectively) was produced from control treatment ( $S_0$ ) and the minimum number of tiller hill<sup>-1</sup> (2.63, 6.67, 10.67, 6.00 at 20, 35, 50 and 65 DAT, respectively) was produced form  $S_2$  treatment. Zeng and Shannon (2000) stated that tiller number hill<sup>-1</sup> and spikelet number panicle<sup>-1</sup> contributed the most variation in grain yield hill<sup>-1</sup> under salinity. Choi *et al.* (2003) observed that tiller number of rice decreased in 0.5% saline water in the soil with low salinity level. Zeng *et al.* (2001) observed that reduction in tiller number per plant was significant only when plants were salinized for 20 days duration before panicle initiation (PI) of rice. The tiller number decreased significantly at 15.62 dSm<sup>-1</sup> salinity level in BR11 rice (Gain *et al.*, 2004). Grattan *et al.* (2002) reported that salinity threshold for rice yield was the EC of 3.0 dSm<sup>-1</sup> and tiller densities reduced by 40% as compared to control (4dSm<sup>-1</sup>).



Mg,  $S_0$ = Control,  $S_1$ = 4 dSm<sup>-1</sup> salt,  $S_1Mg_1$ = 4 dSm<sup>-1</sup> salt + 80 ppm Mg ,  $S_1Mg_2$  = 4 dSm<sup>-1</sup> salt + 160 ppm  $S_2$ = 6 dSm<sup>-1</sup> salt,  $S_2Mg_1$  = 6 dSm<sup>-1</sup> salt + 80 ppm Mg,  $S_2Mg_2$  = 6 dSm<sup>-1</sup> salt + 160 ppm Mg

**Fig. 2**. Effect of different salinity with magnesium on the number of tiller of rice at different days after transplanting

### 4.3 Leaf length

Length of leaf showed statistically significant differences due to the different levels of salinity with magnesium. The longest leaf (48.73 cm) was found at control treatment, which was statistically similar with  $S_1Mg_2$ ,  $S_2Mg_1$ ,  $S_1Mg_1$ ,  $S_2Mg_2$  and  $S_1$  the lowest leaf length (37.17 cm) was recorded at  $S_2$  treatment (Table 1).

### 4.4 Leaf breath

Breath of leaf showed statistically significant differences due to the different levels of salinity with magnesium. The maximum leaf breath (1.48 cm) was obtained at control treatment( $S_0$ ), which was followed by  $S_1Mg_1$ ,  $S_2 Mg_1$  and  $S_1Mg_2$  and the minimum leaf breath (1.13 cm) was recorded at  $S_2$  treatment (Table 1).

### 4.5 SPAD Value of leaf

Analysis of variance indicated that the effect of salinity with magnesium on rice on SPAD value of leaf was varied significantly (Table. 1). The maximum SPAD value (42.13) was found in  $S_2$  treatment. The minimum SPAD value (30.53) was found from  $S_0$  (Control) treatment, which was statistically similar with  $S_1Mg_2$ . Salachna and Zawadzińska, (2014) found that medium- and high-molecular-weight chitosan resulted in higher relative chlorophyll content (SPAD).

Treatment	Leaf length (cm)	Leaf Breath (cm)	SPAD value
S <sub>0</sub>	48.73 a	1.48 a	30.53 d
$S_1$	39.33 bc	1.18 ab	38.00 bc
$S_1Mg_1$	42.60 abc	1.30 ab	31.83 d
$S_1Mg_2$	46.00 a	1.23 ab	36.70 c
<b>S</b> <sub>2</sub>	37.17 c	1.13 b	42.13 a
$S_2Mg_1$	44.00 ab	1.21 ab	40.37 ab
$S_2Mg_2$	46.83 a	1.18 ab	41.07 ab
LSD (0.05)	5.77	0.29	3.34
CV (%)	5.04	7.46	10.03

Table 1. Effect of salinity with magnesium on leaf length, leaf breath and SPAD value on rice

 $\overline{S_0= \text{ Control}, S_1=4 \text{ dSm}^{-1} \text{ salt}, S_1Mg_1=4 \text{ dSm}^{-1} \text{ salt} + 80 \text{ ppm Mg}, S_1Mg_2=4 \text{ dSm}^{-1} \text{ salt} + 160 \text{ ppm Mg}, S_2=6 \text{ dSm}^{-1} \text{ salt}, S_2Mg_1=6 \text{ dSm}^{-1} \text{ salt} + 80 \text{ ppm Mg}, S_2Mg_2=6 \text{ dSm}^{-1} \text{ salt} + 160 \text{ ppm Mg}$ 

In a column figures having similar letter do not differ significantly whereas figures with dissimilar letter differ significantly as per LSD.

#### 4.6 Number of effective tillers

The number of effective tillers hill<sup>-1</sup> of rice was significantly influenced by different levels of salinity with Magnesium (Table2). The highest number of effective tillers hill<sup>-1</sup> (11.00) was recorded at control treatment and the lowest (3.00) was found at  $S_2$ treatment. Bohra and Doerffling (1993) observed that plant height, number of tillers and shoot dry weight reduced under salinity stress in both salt tolerant and salt sensitive rice cultivars. They maintained that salinity stress wasted more energy in salt sensitive rice cultivars than that in salt tolerant ones. Khatun et al. (1995) found that salinity delayed flowering, reduced the number of productive tillers, the number of fertile florets per panicle. Salt tolerance indexes in terms of seed yield, seed weight per panicle, spikelet number per panicle, and tiller number per plant were reduced with increasing salinity (Zeng et al., 2002). Our results also indicate that the percent effective tiller hill<sup>-1</sup> was badly affected at higher salinity levels. However, Magnesium supplementation has mitigated to some extent by increasing effective tiller per hill. In both salinity treatment (4dSm<sup>-1</sup> and 6dSm<sup>-1</sup>) addition of 160 ppm magnesium has increased 1.3 effective tillers per hill (Table 2)

	Number of effect	ive tiller	Number of non-effective		
Treatment	hill <sup>_1</sup>		tiller hill <sup>-1</sup>		
S <sub>0</sub>	11.00	a	0.87	b	
$S_1$	7.67	b	1.00	b	
$S_1Mg_1$	9.67	ab	1.00	b	
$S_1Mg_2$	9.00	ab	1.00	b	
S <sub>2</sub>	3.00	c	2.67	a	
$S_2Mg_1$	7.00	b	1.33	b	
$S_2Mg_2$	8.00	ab	1.00	b	
LSD (0.05)	2.82		0.76		
CV (%)	12.04		10.18		

Table 2. Effect of salinity with magnesium on Number of effective tiller andNon-effective tiller hill<sup>-1</sup> on rice

 $\begin{array}{l} S_0 = \mbox{ Control, } S_1 = 4 \ dSm^{-1} \ salt, \ S_1 M g_1 = 4 \ dSm^{-1} \ salt + 80 \ ppm \ Mg \ S_1 M g_2 = 4 \ dSm^{-1} \ salt + 160 \ ppm \ Mg, \ S_2 = 6 \ dSm^{-1} \ salt, \ S_2 M g_1 = 6 \ dSm^{-1} \ salt + 80 \ ppm \ Mg, \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ salt \ Sm^{-1} \ salt + 160 \ salt \ Sm^{-1} \ salt \ salt \ Sm^{-1} \ salt \ Sm^{-1} \ salt \ salt \ Sm^{-1} \ salt \ salt$ 

In a column figures having similar letter do not differ significantly whereas figures with dissimilar letter differ significantly as per LSD.

#### 4.7 Number of non-effective tillers

The mean effect of different salinity levels with magnesium influenced the number of non-effective tillers hill<sup>-1</sup>(Table 2). The highest number of non-effective tillers hill<sup>-1</sup>(2.67) was observed in S<sub>2</sub> treatment and it was least in control treatment(0.87), The result that magnesium supplementation enhances panicle growth through minimizing toxicity effect of salinity. which was statistically similar with S<sub>1</sub>, S<sub>1</sub>Mg<sub>1</sub>, S<sub>1</sub>Mg<sub>2</sub>, S<sub>2</sub>Mg<sub>1</sub> and S<sub>2</sub>Mg<sub>2</sub>. Alam *et al.*, (2001) stated that the salinity at reproductive stage of rice depressed grain yield much more than that at the vegetative growth stage and at critical salinity levels it might give normal straw yield of rice but produced little or no grain. They also observed that when the plants were continuously exposed to saline media, salinity affected the panicle initiation, spikelet formation, fertilization of florets and germination of pollen grains and hence caused an increase in number of sterile florets. The mutant variety maintained its superiority in various characteristics such as plant height, higher number of fertile panicles per plant (Baloch *et al.*, 2003).

#### 4.8 Panicle length

Length of panicle showed statistically significant differences due to the different levels of salinity with magnesium (Table 3). The longest panicle (24.30 cm) was found at control treatment and the lowest panicle length (21.97 cm) was recorded at  $S_2$  treatment. Khatun *et al.* (1995) and Alam *et al.* (2001) reported that salinity severely reduces the panicle length, seed setting percentage and panicle weight. Similar to panicle bearing tillering in Magnesium supplemented salinity treatment, panicle length also recovered by Mg- supplementation, But the effective is more prominent in higher salinity level (6 dSm<sup>-1</sup> salt) (Table 3).

# **4.9** Number of filled grain panicle<sup>-1</sup>

The number of filled grain panicle<sup>-1</sup> was significantly influenced by different salinity levels with magnesium (Table 3). The highest number of filled grain per panicle (165.90) was recorded at control treatment, which was followed by  $S_1Mg_1$ ,  $S_1Mg_2$ , and  $S_2Mg_1$  and the lowest (106.90) was found at  $S_2$  treatment. Primary branches per panicle was greatly affected in higher salinity stress (Table 3).However the mitigation effect of Magnesium was found better for low salinity level (4dSm<sup>-1</sup> salt)

# 4.10 Number of unfilled grain panicle<sup>-1</sup>

The number of unfilled grain panicle<sup>-1</sup> was significantly influenced by different salinity levels with magnesium (Table 3). The highest number of unfilled grain per panicle (14.67) was recorded from  $S_2$  treatment, the lowest (4.00) was found from control treatment.

#### 4.11 Thousand seed weight

A highly significant variation in thousand seed weight of rice cultivars was observed due to the different salinity levels with magnesium (Table 3). The highest thousand seed weight (20.99g) was recorded at control treatment and it was lowest (16.23g) at  $S_2$  treatment.Similar affect was observed for thousand grain weight i.e more reduction of 1000- grain weight was observed for higher salinity level but mitigation of reduction in 1000-grain weight was least in 4dSm<sup>-1</sup> salinity with 1mM of MgSO<sub>4</sub>

## 4.12 Grain yield hill<sup>-1</sup>

A highly significant variation in grain yield hill<sup>-1</sup> of rice cultivars (BRRI dhan67) was observed due to the different salinity levels with magnesium (Table 3). The highest grain yield (26.57 g) was recorded at control treatment, which was followed by  $S_1Mg_2$ and  $S_1Mg_1$  and it was lowest (15.67 g hill<sup>-1</sup>) at  $S_2$  treatment. In comparison to control (without salt), the reduction of grain yield was high in  $S_2$  (6dSm<sup>-1</sup>) salinity (47.28%) followed by  $S_1$  (4dSm<sup>-1</sup>) salinity (39.45%) (Table 3).But in both of the salt treatment, Magnesium supplementation greatly mitigated. The damaging effect of salinity thereby reducing grain yield reduction. However, the mitigation effect was more in less salinity stress (4dSm<sup>-1</sup>), where yield reduction was 11.4% and 4.61% for 80 and 160 ppm magnesium respectively. Grain yield is the function of number of panicles hill<sup>-1</sup>, number of filled grain panicle<sup>-1</sup> and 1000-grain weight. All the yield contributing characters contributed for the yield reduction hill<sup>-1</sup> under saline conditions; contribution of the seriously affected number of unfilled grains panicle<sup>-1</sup> was the highest (Grattan et al., 2002). Baloch et al. (2003) observed that the mutant variety of rice "Shua-92" maintained its superiority to other varieties in various characteristics such as plant height, higher number of fertile panicles per plant, more fertile grains per panicle, heavy grain size and high plant yield at 7.11- 8.0 dSm<sup>-1</sup> level of salinity.

Treatment	Length of panicle	Number of filled grain panicle <sup>-1</sup>	Number of unfilled grain panicle <sup>-1</sup>	1000 seed weight (g)	Yield per plant (g)
S <sub>0</sub>	24.30 a	165.90 a	4.00 b	20.99 a	26.57 a
$S_1$	22.50 ab	130.20 c	12.33 ab	17.60 c	16.34 d
$S_1Mg_1$	23.73 ab	141.40 b	5.33 ab	18.34 bc	23.40 b
$S_1Mg_2$	23.93 ab	149.60 b	5.47 ab	19.61 b	25.01 ab
$S_2$	21.97 b	106.90 d	14.67 a	16.23 d	15.67 e
$S_2Mg_1$	22.97 ab	140.80 b	11.13 ab	18.02 c	18.19 c
$S_2Mg_2$	23.93 ab	125.70 c	6.33 ab	17.16 cd	18.42 c
LSD (0.05)	1.82	9.91	8.79	1.31	2.18
CV (%)	8.17	8.34	8.33	4.02	12.89

Table 3. Effect of salinity with magnesium yield and yield contributing character of on rice

 $\begin{array}{l} \hline S_0 = \mbox{ Control, } S_1 = 4 \ dSm^{-1} \ salt, \ S_1 M g_1 = 4 \ dSm^{-1} \ salt + 80 \ ppm \ Mg \ S_1 M g_2 = 4 \ dSm^{-1} \ salt + 160 \ ppm \ Mg, \ S_2 = 6 \ dSm^{-1} \ salt, \ S_2 M g_1 = 6 \ dSm^{-1} \ salt + 80 \ ppm \ Mg, \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_2 = 6 \ dSm^{-1} \ salt + 160 \ ppm \ Mg \ S_2 M g_3 \ salt + 160 \ ppm \ Mg \ S_2 M g_4 \ salt + 160 \ ppm \ Mg \ S_3 M g_4 \ salt + 160 \ ppm \ Mg \ S_4 M g_4 \ salt \ salt \ S_4 M g_4 \ salt \ S_4 M$ 

In a column figures having similar letter do not differ significantly whereas figures with dissimilar letter differ significantly as per LSD.

#### **CHAPTER V**

# SUMMARY AND CONCLUSION

A pot experiment was conducted at the net house of the Department of Agricultural Botany, Sher-e-Bangla Agricultural University, Dhaka-1207, during *Boro* rice growing season (December to April) of the year of 2017- 18 to observe the role of magnesium to alleviate damaging effect of salt stress on development and yield performance of rice plant (*Oryza sativa* L.). The experiment was conducted using seven salinity levels with magnesium viz.  $S_0 = \text{Control}$ ,  $S_1 = 4d\text{Sm}^{-1}$  salt,  $S_1\text{Mg}_1 = 4d\text{Sm}^{-1}$  salt + 80 ppm Mg,  $S_1\text{Mg}_2 = 4d\text{Sm}^{-1}$  salt + 160 ppm Mg,  $S_2 = 6d\text{Sm}^{-1}$  salt,  $S_2\text{Mg}_1 = 6d\text{Sm}^{-1}$  salt + 80 ppm Mg,  $S_2\text{Mg}_2 = 6d\text{Sm}^{-1}$  salt + 160 ppm Mg. The experiment was set in Randomized Complete Block Design (RCBD) having one factor with three replications.

The results on the effect of morphological characters indicated that plant height; numbers of tillers, leaf length, leaf breath, and SPAD value, effective tiller, number of non effective tiller, panicle length, filled grain, unfilled grain and thousand seed weight grain yield were significantly influenced by salinity and magnesium. All the measured morphological parameters were found highest in non saline control treatment .But the parameters gradually decline in 4 and 6 dSm<sup>-1</sup> salt treatment. However, Magnesium supplementation greatly mitigated the damaging effect of salt for the growth, development and yield of BRRI dhan67. But the mitigation effect was more prominent at low salinity stress (4dSm<sup>-1</sup>) compared to higher stress (6dSm<sup>-1</sup>).

Moreover application of 160 ppm, showed better recovery compared to the 4dSm<sup>-1</sup> salt stress.

Considering above facts, it could be concluded that, application of magnesium under salt stress condition could be a good option for obtaining better yield of salt tolerant varieties like BRRI dhan67. Further, research needed to get a concrete recommendation considering more varieties and a number of salt stress and magnesium level.

### REFERENCES

- Abdullah, Z., Khan, M.A., and Flowers, T.J. (2001). Causes of sterility in seed set of rice under salinity stress. *J. Agron. Crop Sci.*. **187**(1): 25-32.
- Abdullah, Z., Khan, M.A., Flowers, T.J., Ahmad, R. and Malik, K.A. (2002). Causes of sterility in rice under salinity stress. *Pros. Saline Agric*. 177-187.
- Alam, S.M., Ansari, R., Mujtaba, S.M. and Shereen, A. (2001). Salinization of millions of hectares of land continues to reduce crop productivity severely worldwide. *In*: Saline Lands and Rice: Industry & Economy. *Pakistan Economist.* 17: 60-71.
- Apse, I.K. (1988). Effect of saline irrigation water in the growth, yield and chemical comp osition of the rice crop grown in a saline soil. J. Indian Soc. Soil Sci. 36: 324-329.
- Apse, M.P., Aharon, G.S., Snedden, W.A. and Blumwald, E. (1999). Salt tolerance conferred by overexpression of a vacuolar Na<sup>+</sup>/H<sup>+</sup> antiport in *Arabidopsis. Sci.* 285: 1256-1258.
- Ashraf, M. (1994a). Breeding for salinity tolerance in plants. *Crit. Rev. Plant Sci.* 13: 17-42.
- Ashraf, M. (2002). Salt tolerance of cotton: some new advances. *Crit. Rev. Plant Sci.*21: 1-30.

- Ashraf, M., Rahmatullah, A.R., Bhatti, A.S., Afzal, M., Sarwar, A., Maqsood, M.A. and Kanwal, S. (2010). Amelioration of salt stress in rice (*Oryza sativa* L.) by supplying potassium and silicon in hydroponics. *Pedosphere*. **20**(2): 153–162
- Aslam, M., Mahmood, I.H., Qureshi, R.H., Nawaz, S., Akhtar, J. and Ahmad, Z. (2001). Nutritional Role of Calcium in Improving Rice Growth and Yield under Adverse Conditions. *Int. J. Agri. Biol.*, 3(3): 292-297.
- Aslam, M., Muhammad, N., Qureshi, R.H., Ahmad, Z., Nawaz, S. and Akhtar, J. (2003). Calcium and salt-tolerance of rice. *Common. Soil Sci. Plant Anal.* 34 (19&20): 3013-3031.
- Baba, T and Fujiyama, H. (2003). Differences in short-term responses of rice and tomato to sodium salinization and supplemental potassium and Calcium. *Soil Sci. Plant Nutr.* 49: 669-675.
- Baloch, A.W., Soomro, A.M., Javed, M.A., Bughio, H.R., Alam, S.M., Bughio, M.S.,
  Mohammed, T. and Mastoi, N.N. (2003). Induction of salt tolerance in rice
  through mutation breeding. *Asian J. Plant Sci.* 2: 273-276.
- Bohra, J.S. and Doerffling, K. (1993). Potassium nutrition of rice (*Oryza sativa* L.) varieties under NaCl salinity. *Plant and soil*. **152**(2): 299-303.
- Choi, W.Y., Lee, K.S., Ko, J.C., Choi, S.Y. and Choi, D.H. (2003). Critical saline concentration of soil and water for rice cultivation on a reclaimed saline soil. *Korean J. Crop Sci.* 48(3): 238-242.

- Colmer, T.D., Teresa, W.M.F., Richard, M.H. and Andre, L. (1994).Interactions of Ca2+ and NaCl stress on the ion relations and intercellular pH of sorghum bicolor root tips: An in vivo 31P. NMR study. *J. Expt. Bot.*, **45**: 1037–44.
- Cominelli, E., Conti, L., Tonelli, C., Galbiati, M. (2013) Challenges and perspectives to improve crop drought and salinity tolerance. New Biotechnol, **30**, 355–361.
- Davitt, D., Jarrell, W.M. and Stevens, K.L. (1981). Sodium-Potassium ratio in soil solution and plant response under saline conditions. *Soil Sci. Soc.Am. J.*, 45: 80–86.
- Dayton, L. (2014). Agribiotechnology: Blue-sky rice. Nature, 514, S52–S54.
- Din, C., Mehdi, S.M., Sarfraz, M., Hassan, G. and Sadiq, M. (2001). Comparative Efficiency of Foliar and Soil Application of K on Salt Tolerance in Rice. *Pakistan J. of Biological Sci.* 4(7): 815 – 817.
- Eckardt, N.A. (2009). The future of science: Food and water for life. Plant Cell . 21, 368–372.
- Epstein, E. and Rains, D.W. (1987). Advances in salt tolerance. *In:* Gableman, H.W. and Loughman, B.C. (eds.), Genetic Aspects of Plant Mineral Nutrition. Martinus Nijhoff, Boston. pp. 113-125.
- Finck, A. (1977). Soil salinity and plant nutritional status. *In:* Managing Saline Water. Proceedings of the International Conference. Texas Tech. Univ., Lubbock, Texas. pp. 199-210.

- Food and Agriculture Organization (2012). World Water Day 2012 Celebration. Un Conference Centre, Bangkok, 22 March 2012. Available online:http://www.Fao.Org/asiapacific/rap/home/meetings/list/detail/en/?Meeti ngs\_id=637&year=2012 (accessed on 20 August 2016).
- Food and Agriculture Organization. (2009) Declaration of the World Summit on Food Security. Rome, Italy, 16–18 November 2009. Available nline:http://www.Fao.Org/wsfs/world-summit/wsfs-challenges/en/
- Food and Agriculture Organization.(1988). Salt-Affected Soils and Their Management. Available online: <u>http://www.fao.org/docrep/ x5871e/</u> <u>x5871e00.htm</u>
- Franco, O.L., Filho, J.E., Prisco, J.T. and Filho, E.G. (1999). Effects of CaCl<sub>2</sub> on growth and osmoregulator accumulation in NaCl stressed cowpea
- Gain, P., Mannan, M.A., Pal, P.S., Hossain, M.M. and Parvin, S. (2004). Effect of salinity on some yield attributes of rice. *Pakistan J. Biol. Sci.* 7(5): 760-762.
- Girija, C., Smith B.N. and Swamy, P.M. (2002). Inter-active effects of sodium chloride and calcium chloride on the accumulation of proline and glycinebetaine in peanut (*Arachis hypogaea* L.). *Envi. & Exper. Bot.* **47**: 1-10.
- Grattan, S.R., Zeng, L., Shannon, M.C. and Roberts, S.R. (2002). Rice is more sensitive to salinity than previously thought. *Calif. Agric.* **56**(6): 189-195.
- Hermans, C., Bourgis, F., Faucher, M., Strasser, R.J., Delrot, S., and Verbruggen, N. (2005). Magnesium Deficiency in Sugar Beets Alters Sugar Partitioning and Phloem Load- ing in Young Mature Leaves, *Planta*, 220(4) : pp. 541-549.

- Hoang, T.M.L., Williams, B., Khanna, H., Dale, J. and Mundree, S.G. (2014).Physiological basis of salt stress tolerance in rice expressing the antiapoptotic gene SfIAP. Funct. Plant Biol. 41, 1168–1177.
- Hossain, M. and Fischer, K.S. (1995). Rice research for food security and sustainable agricultural development in Asia: Achievements and future challenges. GeoJournal, **35**, 286–298.
- Jacoby, B. (1999). Mechanisms involved in salt tolerance of plants. *In:* Pessarakli, M. (ed.), Handbook of Plant and Crop Stress (2nd ed.). Marcel Dekker, New York. pp. 97-123.
- Karim, Z., Hussain, S.G. and Ahmed, M. (1990). Salinity problems and crop intensification in the coastal regions of Bangladesh, BARC, *Soil Publ.* 3(3):3-6.
- Khan, H.R., Faiz, S.M.A., Islam, M.N., Adachi, T. and Ahmed, I.U. (1997). Effects of salinity, gypsum and Zn on mineral nutrition of rice. *Int. J. Trop. Agric.* 10(2): 147-156.
- Khatun, S., Rizzo, C.A. and Flowers, T.J. (1995). Genotypic variation in the effect of salinity on fertility in rice. *Plant Soil.* **173**(2): 239-250.
- Khush, G. (2005). What it will take to feed 5.0 billion rice consumers in 2030. Plant Mol. Biol., **59**, 1–6.
- LaHaye, P.A. and Epstein, E. (1971). Calcium and salt tolerance by bean plants. *Physiol. Plant.*, **25**: 213–8.
- Lauchli, A., (1990). Calcium, salinity and the plasma membrane. *In:* R.T. Leonard,P.K. Hepler (Eds.), *Calcium in plant growth and development*, pp. 26–35.American Society of Plant Physiologist, Rockville, M.D. Aslam, M.N.,

Muhammad, R.H., Qureshi, J., Akhtar and Ahmed, Z. (2000). Role of Ca2+ in salinity tolerance of rice. Symp. *On Integ. PlantManage*. No. 8-10 (1998), Islamabad.

- Linares, O.F. (2002) African rice (*Oryza glaberrima*): History and future potential. Proc. Natl. Acad. Sci. USA, **99**.16360–16365.
- Lynch, J. and Lauchli, A. (1988a). Salt stress disturbs the calcium nutrition of barley (*Hordeum Vulgare* L.). *New Phytol.*, **99**: 345–54.
- Maloo, S.R. (1993). Breeding and screening techniques for salt tolerance in crop plants. *In:* Somani, L. L. and K. L. Totawat (eds.), Management of Salt-Affected Soils and Waters. Agrotech Publishing Academy, Udaipur-313001, India. pp. 321-357.
- Manneh, B. (2004). Genetic Physiological and Modeling Approaches towards Tolerance to Salinity and Low Nitrogen Supply in Rice (*Oryza sativa* L.). PhD. Thesis, Wageningen University, Wageningen, The Netherlands. 206.
- Marschner, H. (1995). "Mineral Nutrition of Higher Plants," 2<sup>nd</sup> Edition, Academic Press, London.
- Martinez, V. and Lauchli, A. (1993). Effect of Ca<sup>2+</sup>z on the salt stress response of barley roots as observed by in vivo 31P-nuclear magnetic resonance and *in vitro* analysis. *Planta.*, **1909**: 519–24.
- Mengel, K. and Kirkby, E.A. (1987). "Principles of Plant Nutri- tion," 4th Edition, International Potash Institute, Switzer- land.
- Mondal, M.K., Bhuiyan, S.I., and Franco, D.T. (2001). Soil salinity reduction and production of salt dynamics in the coastal rice lands of Bangladesh. *Agric*.

*Water Manage.* **47:** 9-23

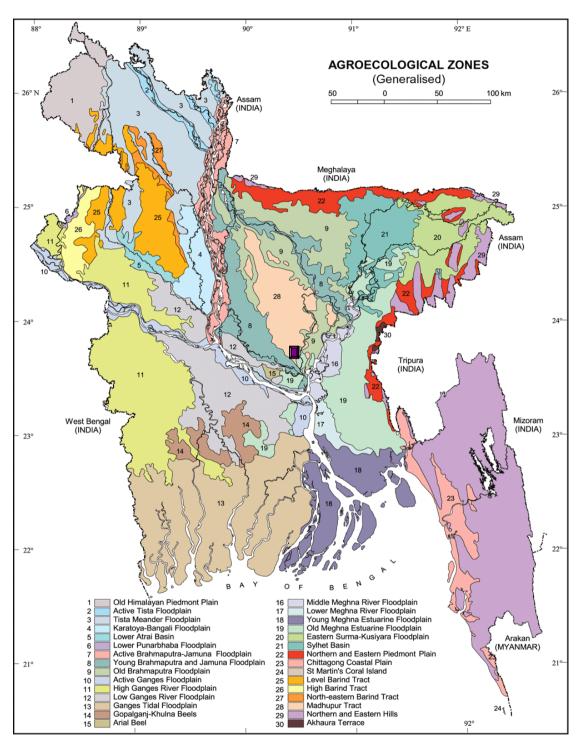
- Muhammad, N. (1998). Salt tolerance of rice (*Oryza sativa*) as affected by nutrient supply. *Ph.D. Thesis*, Univ. Agric. Faisalabad. Pakistan.
- Munns, R. (2002). Comparative physiology of salt and water stress. *Plant Cell Environ.* **25**: 239-250.
- Munns, R. (2011). Plant adaptations to salt and water stress: Differences and commonalities. In Advances in Botanical Research; Ismail, T., Ed.; Elseiver: Wyman Street, Waltham, MA, USA, Volume 57, pp. 1–32.
- Munns, R. Genes and salt tolerance: Bringing them together. New Phytol. (2005), **167**, 645–663.
- Munns, R., Tester, M. (2008) Mechanisms of salinity tolerance. Annu. Rev. Plant Biol.,**59**, 651–681.
- Murshed, M.G., Hossain, M.A., Tareq, M.Z. & Ashrafuzzaman, M. (2008) Short-term effects of salinity on seedling growth of rice. *Bangladesh J. of envi. Sci.***14**(1): 66–70.
- Rengasamy, P. (2006).World salinization with emphasis on Australia. J. Exp. Bot., 57, 1017–1023.
- Saleque, M.A., Choudhury, N.N., Rezaul-Karim, S.M. and Panaullah, G.M. (2005). Mineral nutrition and yield of four rice genotypes in the farmers' fields of salt affected soils. *J. Plant Nutr.* 28(5): 865-875.
- Shabala, S. and Hariadi, Y. (2005). "Effects of Magnesium Avail- ability on the Activity of Plasma Membrane Ion Trans- porters and Light-Induced Responses from Broad Bean Leaf Mesophyll," *Planta*, **221**(1): pp.56-65.

- Shaul, O. (2002). Magnesium Transport and Function in Plants: the Tip of the Iceberg, *Biometals*, **15**(3): 309-323.
- Shrivastava, P.; Kumar, R. Soil salinity: A serious environmental issue and plant growth promoting bacteria as one of the tools for its alleviation. *Saudi J. Biol. Sci.* 22, 123–131.
- SRDI (2001). Soil salinity in Bangladesh. Soil Resource Development Institue, Ministry of Agriculture, Dhaka, Bangladesh.
- SRDI (2010). Saline soils of Bangladesh. Soil Resource Development Institue. SRMAF Project, Ministry of Agriculture, Dhaka, Bangladesh.
- Tavakkoli, E., Rengasamy, P. and Mcdonald, G.K. (2010). High concentrations of Na<sup>+</sup> and Cl<sup>-</sup> ions in soil solution have simultaneous detrimental effects on growth of faba bean under salinity stress. *J. Exp. Bot.* **61**: 4449-4459.
- Tester, M., Langridge, P. (**2010**). Breeding technologies to increase crop production in a changing world. Science, **327**, 818–822.
- Umali, D.L. (1993) Irrigation-Induced Salinity a Growing Problem for Development and the Environment; The Word Bank: Washington, DC, USA.
- United States Department of Agriculture. Bibliography on Salt Tolerance. Fibres, Grains and Special Crops; Brown, G.E., Jr., Ed.; Salinity Laboratory United States Department of Agriculture Research Service: Riverside, CA, USA. Availableonline:http://www.Ars.Usda.Gov/services/docs.Htm?Docid=8908.
- UNPF (The United Nations Population Fund. Linking Population, Poverty and Development). 2016Available online: <u>http://www.unfpa.org/pds/trends.htm</u>

- Vaidyanathan, H., Sivakumar, P., Chakrabarty, R. & Thomas, G. (2003) Scavenging of reactive oxygen species in NaCl-stressed rice (*Oryza sativa* L.) differential response in salt-tolerant and sensitive varieties. *Plant Sci.* 165: 1411–1418.
- Vaughan, D.A., Lu, B.R., Tomooka, N. (2008). The evolving story of rice evolution. Plant Sci., **174**, 394–408.
- Virmani, S.S., Ilyas-Ahmed, M. (2008). Rice breeding for sustainable production. InBreeding Major Food Staples; Kang, M., Priyadarshan, P.M., Eds., Wiley:Hoboken, NJ, USA.
- Wassmann, R., Hien, N.X., Hoan, C.T. & Tuong, T.P. (2004) Sea level rise affecting the Vietnamese Mekong Delta: water elevation in the flood season and implications for rice production. *Climatic Change* 66: 89–107.
- Wu, G.Q. and Wang, S.M. (2012). Calcium regulates K+/Na+ homeostasis in rice (*Oryza sativa* L.) under saline conditions. Plant soil environ., 58(3): 121–127.
- Yeo, A.R. (1994). Physiological criteria in screening and breeding. *In:* Yeo, A.R. and Flowers, T.J. (eds.), Soil Mineral Stresses, Approaches to Crop Improvement. *Springer Verlag, Berlin.* pp. 37-60.
- Yousof, F.I. (2014). Effect of rice seed priming with calcium chloride (CaCl2) on germination and seedlings vigor under salinity stress. J. Plant Production, Mansoura Univ., 4: 523 – 535.
- Zafar, S., Ashraf, M.Y., Sarwar, G., Mahmood, S., Kausar, A. and Ali, I. (2004). Variation in growth and ion uptake in salt tolerant and sensitive rice cultivars under NaCl salinity. Asian J. of Plant Sci. **3**: 156-158.

- Zeng, L., and Shannon, M.C. (2000). Salinity effects on seedling growth and yield components of rice. *Crop Sci.* **40**: 996-1003.
- Zeng, L., Shannon, M.C. and Grieve, C.M. (2002). Evaluation of salt tolerance in rice genotypes by multiple agronomic parameters. *Euphytica*. **127**: 235-245.

APPENDICES Appendix I. Map showing the experimental sites under study



**The experimental site under study**