# ECO-FRIENDLY MANAGEMENT OF SUCKING AND OTHER FOLIAR INSECT PESTS OF BRINJAL

### SANJUKTA BISWAS



# DEPARTMENT OF ENTOMOLOGY SHER-E-BANGLA AGRICULTURAL UNIVERSITY

# SHER-E-BANGLA NAGAR, DHAKA-1207, BANGLADESH

JUNE, 2018

# ECO-FRIENDLY MANAGEMENT OF SUCKING AND OTHER FOLIAR INSECT PESTS OF BRINJAL

BY

# SANJUKTA BISWAS

# **REGISTRATION NO. 12-05146**

A Thesis

Submitted to the Department of Entomology, Faculty of Agriculture, Sher-e-Bangla Agricultural University, Dhaka in partial fulfillment of the requirements for the degree of

# MASTER OF SCIENCE (MS) IN ENTOMOLOGY

### **SEMESTER: JANUARY-JUNE, 2018**

**Approved by:** 

Prof. Dr. Md. Razzab Ali

Supervisor Department of Entomology SAU, Dhaka Dr. Kabir Uddin Ahmed Co-supervisor Chief Scientific Officer

**Planning and Evaluation Division** 

**BARC**, Farmgate, Dhaka

Prof. Dr. S. M. Mizanur Rahman Chairman Examination Committee



**DEPARTMENT OF ENTOMOLOGY** 

Sher-e-Bangla Agricultural University (SAU) Sher-e-Bangla Nagar, Dhaka-1207

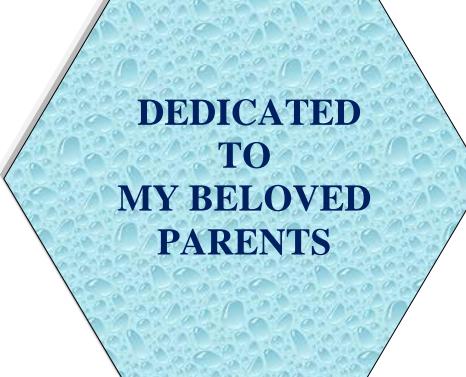
# CERTIFICATE

This is to certify that thesis entitled "ECO-FRIENDLY MANAGEMENT OF SUCKING AND OTHER FOLIAR INSECT PESTS OF BRINJAL" submitted to the Faculty of Agriculture, Sher-e-Bangla Agricultural University (SAU), Dhaka in partial fulfillment of the requirements for the degree of MASTER OF SCIENCE (MS) IN ENTOMOLOGY, embodies the result of a piece of bona fide research work carried out by SANJUKTA BISWAS, Registration no. 12-05146 under my supervision and guidance. No part of the thesis has been submitted for any other degree or diploma.

I further certify that such help or source of information, as has been availed of during the course of this investigation has duly been acknowledged.



Dated: June, 2018 Place: Dhaka, Bangladesh Prof. Dr. Md. Razzab Ali Supervisor Department of Entomology SAU, Dhaka



### ACKNOWLEDGEMENT

At first, the author takes the opportunity to express her deepest sense of gratefulness to Almighty Allah enables the author to complete her research work for the degree of Master of Science (MS) in Entomology.

The author really does not have adequate words to express her heartfelt sense of gratification, ever indebtedness and sincere appreciation to her benevolent teacher and research supervisor, Dr. Md. Razzab Ali, Professor, Department of Entomology, Sher-e-Bangla Agricultural University, Dhaka-1207, for his constant help, scholastic guidance, planning experiment, valuable suggestions, timely and solitary instructive criticism for successful completion of the research work as well as preparation of this thesis.

It is a great pleasure for the author to express her sincere appreciation, profound sense, respect and immense indebtedness to her respected co-supervisor, Dr. Kabir Uddin Ahmed, Chief Scientific Officer, Bangladesh Agricultural Research Council, Farmgate, Dhaka for providing her with all possible help during the period of research work and preparation of the thesis.

The author would like to express her deepest respect and boundless gratitude to the Chairman and all the teachers of the Department of Entomology, Sher-e-Bangla Agricultural University, Dhaka-1207 for their sympathetic co-operation and inspiration throughout the course of this study and research work.

Cordial thanks are also due to all laboratory staff and field workers of SAU farm for their cooperation to complete her research work in the field.

The author would like to express her last but not least profound and grateful gratitude to her beloved parents, friends and all of her relatives for their inspiration, blessing and encouragement that opened the gate of her higher studies in her life.

Date: June, 2018 SAU, Dhaka The Author

### ECO-FRIENDLY MANAGEMENT OF SUCKING AND OTHER FOLIAR INSECT PESTS OF BRINJAL

#### BY

#### SANJUKTA BISWAS

# ABSTRACT

A field experiment was conducted in the experimental field of Sher-e-Bangla Agricultural University to find out the effective environment friendly management practice(s) of brinjal, cultivated during Rabi season (November, 2017 to March, 2018). The experiment was laid out in Randomized Complete Block Design (RCBD) with three replications. The experimental treatments were T<sub>1</sub> (spraying of neem oil @ 3 ml neem oil and 10 ml trix mixed with 1 liter of water @ 7 days interval); T<sub>2</sub> (spraying of neem seed kernel extract @ 5.0 ml/L of water at 7 days interval); T<sub>3</sub> (spraying of bioneem plus @ 3.0 ml/L of water at 7 days interval); T<sub>4</sub> (spraying of Marshal 25 EC@ 3.0 ml/L of water at 7 days interval); T<sub>5</sub> (spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval); T<sub>6</sub> (spraying of Imitaf 20 SL @ 0.1 ml/L of water at 7 days interval); T<sub>7</sub> (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) and T<sub>8</sub> (untreated control). Treatment T<sub>7</sub> contributed to reduce the highest number of leaf hopper, aphid, epilachna beetle, leaf roller, percent leaf infestation and plant infestation by leaf hopper, aphid, epilachna beetle, leaf roller were 5.72 leaf hopper/plant, 2.79 aphid/plant, 1.14 epilachna beetle/plant, 2.12 leaf roller/plant, 13.45%leaf/ five plant, 12.03 %leaf/ five plants, 9.20% leaf/five plants, 5.66% leaf/five plants, 13.89% plant/plot, 13.89% plant/plot, 8.33% plant/plot, 8.33% plant/plot respectively. Similarly, T<sub>1</sub> also contribute to reduce the number of leaf hopper (6.91), aphid (3.89), epilachna beetle (2.03), leaf roller (3.03), percent infestation of leaf and plant respectively by them. According to eco-friendly management  $T_1$  showed the highly incidence of beneficial arthropods like lady bird beetle (5.21 lady bird beetle/plant), field spider (3.53 field spider/plant), ants (4.05 ants/plant) etc. in the brinjal field throughout the growing season. T<sub>7</sub> showed best performance for high single fruit weight (39.00 gm) and yield (25.57 ton/ha) of brinjal. T<sub>1</sub> also showed more or less similar performance in case of single fruit weight (34.66 gm) and yield (24.59 ton/ha). Considering the environmental hazard and effect of incidence of beneficial arthropods T<sub>1</sub> was the best treatment against sucking and other foliage insect pests of brinjal.

# **TABLE OF CONTENTS**

| CHAPTER |     | TITLE                         | PAGE         |
|---------|-----|-------------------------------|--------------|
|         |     | ACKNOWLEDGEMENT               | i            |
|         |     | ABSTRACT                      | ii           |
|         |     | TABLE OF CONTENTS             | iii          |
|         |     | LIST OF TABLES                | iv           |
|         |     | LIST OF FIGURES               | $\mathbf{V}$ |
|         |     | LIST OF ABBREVIATIONS AND     | vi           |
|         |     | ACRONYMS                      |              |
| CHAPTER | Ι   | INTRODUCTION                  | 01           |
| CHAPTER | II  | <b>REVIEW OF LITERATURE</b>   | 05           |
| CHAPTER | III | MATERIALS AND METHODS         | 26           |
| CHAPTER | IV  | <b>RESULTS AND DISCUSSION</b> | 35           |
| CHAPTER | V   | SUMMARY AND CONCLUSION        | 83           |
| CHAPTER | VI  | REFERENCES                    | 90           |
| CHAPTER | VII | APPENDICES                    | 102          |

# LIST OF TABLES

| TABLE<br>NO. | NAME OF THE TABLES  | PAGE<br>NO. |
|--------------|---|-------------|
| 1            | Effect of management practices of number of leaf hopper<br>on fully opened leaves per plant                 | 36          |
| 2            | Effect of management practices on infestation of fully opened leaves by leaf hopper per five plants         | 38          |
| 3            | Effect of management practices of infestation of plants by leaf hopper per plot                             | 40          |
| 4            | Effect of management practices of number of aphid on fully opened leaves per plants                         | 41          |
| 5            | Effect of management practices on infestation of fully opened leaves by aphid per five plants               | 43          |
| 6            | Effect of management practices of infestation of plants by aphid per plot                                   | 45          |
| 7            | Effect of management practices of number of epilachna<br>beetle on fully opened leaves per plant            | 47          |
| 8            | Effect of management practices on infestation of fully<br>opened leaves by epilachna beetle per five plants | 49          |
| 9            | Effect of management practices on infestation of plants by<br>epilachne beetle per plot                     | 51          |
| 10           | Effect of management practices against number of leaf<br>roller on fully opened leaves per five plants      | 53          |
| 11           | Effect of management practices of infestation of fully<br>opened leaves by leaf roller per five plants      | 55          |
| 12           | Effect of management practices against infestation of plants<br>by leaf roller per plot                     | 57          |
| 13           | Effect of management practices on incidence of lady bird beetle per plants                                  | 59          |
| 14           | Effect of management practices on incidence of field spider<br>per plot                                     | 61          |
| 15           | Effect of management practices on incidence of ants per plot  | 62          |
| 16           | Effect of management practices on fruit length of brinjal during growing season                             | 64          |
| 17           | Effect of management practices on fruit girth of brinjal during growing season                              | 66          |
| 18           | Effect of management practices on fruit girth of brinjal during growing season                              | 68          |
| 19           | Effects of management practices on number of branch per<br>plant, plant height per plot and yield           | 69          |

| FIGURE<br>NO. | TITLE  | PAGE<br>NO. |
|---------------|--|-------------|
| 1             | Relationship between number of leaf hopper and yield of                      | 70          |
|               | brinjal  |             |
| 2             | Relationship between number of aphid and yield of brinjal                    | 71          |
| 3             | Relationship between mean number of epilachna beetle and yield of brinjal    | 72          |
| 4             | Relationship between number of leaf roller and yield of brinjal              | 73          |
| 5             | Relationship between percent leaf infestation by leaf hopper<br>and yield    | 74          |
| 6             | Relationship between percent leaf infestation by aphid and yield             | 75          |
| 7             | Relationship between percent leaf infestation by epilachna beetle and yield  | 76          |
| 8             | Relationship between percent leaf infestation by leaf roller<br>and yield    | 77          |
| 9             | Relationship between percent plant infestation by leaf hopper and yield      | 78          |
| 10            | Relationship between percent plant infestation by aphid and yield            | 79          |
| 11            | Relationship between percent plant infestation by epilachna beetle and yield | 80          |
| 12            | Relationship between percent plant infestation by leaf roller<br>and yield   | 81          |
| 13            | Relationship between single fruit weight and yield                           | 82          |

# LIST OF FIGURES

# LIST OF ABBREVIATIONS AND ACRONYMS

| Abbreviation | Full meaning                                    |
|--------------|---|
| BADC         | Bangladesh Agricultural Development Corporation |
| BARI         | Bangladesh Agricultural Research Institute      |
| BBS          | Bangladesh Bureau of Statistics                 |
| BCPC         | British Crop Production Council                 |
| CV           | Coefficient of variation                        |
| °C           | Degree Celsius                                  |
| d.f.         | Degrees of freedom                              |
| et al.       | And others                                      |
| EC           | Emulsifiable Concentrate                        |
| FAO          | Food and Agriculture Organization               |
| gm           | Gram  |
| ha           | Hectare   |
| IPM          | Integrated Pest Management                      |
| CRSP         | Collaborative Research Support Program          |
| J.           | Journal   |
| Kg           | Kilogram  |
| LSD          | Least Significant Difference                    |
| mg           | Milligram                                       |
| ml           | Milliliter                                      |
| MP           | Muriate of Potash                               |
| %            | Per cent  |
| RCBD         | Randomized Complete Block Design                |
| SAU          | Sher-e-Bangla Agricultural University           |
| TSP          | Triple Super Phosphate                          |
| WP           | Wetable Powder                                  |

# **CHAPTER I**

# **INTRODUCTION**

Brinjal or eggplant (*Solanum melongena* L.) is the most common, popular and principal vegetables in Bangladesh and other parts of the world (Nonnecke, 1989). Brinjal is the second most important vegetables crops after potato in relation to its total production (Anon., 1996). This useful crop is grown year round in Bangladesh and covers 48679 ha with a production of 507000 tons (BBS, 2017) with about 25.4% of the total vegetable area of the country. Brinjal is grown all over areas and seasons of Bangladesh. But the brinjal is intensively grown in winter season in Jashore, Mymensingh, Narsingdi, Cumilla, Savar, Bogura, Jamalpur, Dinajpur and Rajshahi districts of Bangladesh.

Brinjal is well known for its nutritive value as a source of carbohydrate, proteins, minerals and vitamin (FAO, 1995). It is also a good source of dietary fiber and folic acid, and is very low in saturated fat, cholesterol and sodium. It is a native to India and is extensively grown in all the Southeast Asian countries. Brinjal is one of the most important vegetables in South Asia (India, Nepal and Srilanka). This region of South Asian accounts for almost 50% of the world area under brinjal cultivation (Alam *et al.*, 2003). Brinjal is grown in Bangladesh throughout the year including the summer season, when the supply of vegetables in the market is scarce. Thus, the farmers find it as a cash crop, which serves as a source of continuous flow of income (FAO, 2003). Sales of eggplant throughout the prolonged harvest season provide farmers with valuable cash income (Alam *et al.*, 2003).

Brinjal is a high income generating crop and its price reaches up to Tk.80 per kilogram during the month of Ramadan in Bangladesh. Due to various uses of brinjal, it is liked by rich and poor, urban and rural class people. The importance of brinjal to the farmers stems from its reasonably consistent and high yields of about 19 tons/ha per growing season. The crop is relatively hard and can withstand adverse conditions better than other crops. It can also be chopped and regrown as a perennial crop. Brinjal is extensively grown in kitchen and commercial gardens in both rabi and kharif season in Bangladesh, especially in the hot humid monsoon season when other vegetables are in short supply. Brinjal is practically the only vegetable that is available at an affordable price for rural and urban consumers. It is cultivated largely on small family owned farms, where weekly sales of it brings in a readily cash income. The crop is infested by various arthropods pest species in the field. El-Shafie (2001) recorded 28 species of insect pests under 7 different insect orders from the brinjal ecosystem in Sudan. Latif (2007) observed 20 species of pest under 6 different orders, jassid was the second most common in the field after brinjal shoot and fruit borer. Srinivasan (2009) reported that, eggplant production is severely constrained by several insect and mite pests. The major pests include eggplant shoot and fruit borer, leaf hopper, whitefly, thrips, aphid, spotted beetles, leaf roller, stem borer, blister beetle, red spider mite, and little leaf disease.

The leafhopper sucks the nutrient sap from the xylem and severe infestation results in crinkling of leaves, hopper burn and cupping up symptoms (Anand *et al*, 2013). Leafhopper nymphs and adults sucks the sap or juice of the plant; however, damage to the crop is brought about not so much by its direct feeding but rather by the plant reaction to the toxin in its saliva (Barroga and Bernardo, 1993). Damage symptom due to the toxic saliva, which is slight yellowing of leaf margins become noticeable even at low population density of 1-2 leafhopper (Navasero, 2003). Aphid causes cupping, distortion of leaves and stunted growth of plants. It suck cell sap from the leaves and secrete honey dew, which causes fungal diseases (Cueva, *et al.*, 2015). Adult and larva of epilachna beetle feed on leaves by scraping the surface cells between veins leaving

marks, which are initially C-shaped that later on result in irregularly-shaped holes or strips. High level of infestation severely damages the leaves, giving them a skeletonized or lace-like coupled with slight yellowish to brownish appearance due to drying of affected tissues. Leaf roller suck the cell sap from the underside of the leaves and curl leaves upward and role that and stay in that. Farmers spray synthetic insecticides four to six times for managing these sucking pests, resulting in the reduction of natural enemies and beneficial organisms. Even though, neonicotinoids are widely used for managing the homopteran insect pest, very little work on their side effects on natural enemy has been carried out (Cloyd and Bethke, 2011).

The management of these sucking and foliar pests through various non-chemical method namely, cultural, mechanical, biological and host plant resistant etc. was limited throughout the world. Management practices in Bangladesh and other countries are still limited to frequent spray of toxic chemical pesticides (Alam, 2005; Anon., 2005; Misra and senapati, 2003; Singh and choudhary, 2001; Bhargava et al., 2001; Ali and Karim, 1994; Yadgirwar et al., 1994; Singh et al., 1991). Farmers spray synthetic insecticides four to six times for managing these sucking pests, resulting in the reduction of natural enemies and beneficial organisms. Even though, neonicotinoids are widely used for managing the homopteran insect pest, very little work on their side effects on natural enemy has been carried out (Cloyd and Bethke, 2011). The insecticides used mostly belong to organophosphates, carbamates, and synthetic pyrithroides. Bangladeshi farmers usually apply six to eight schedule based insecticide sprays against this pest throughout the season. But this kind of insect pest control strategy relying solely on chemical protection had got many limitations and undesirable side effects (Husain 1993, 1984) and this in the long run led to many insecticides related complications (Frisbie, 1984) such as direct toxicity to beneficial insect, fishes and other non-target organism (Munakata, 1997; Goodland et al., 1985; Pimentel,

1981), human health hazards (Bhaduri *et al.*, 1989) resurgence of pests (Husain, 1993; Luckmann and Metcalt, 1975) out-break of secondary pest (Hagen and Franz, 1973) and environmental pollution (Fiswick, 1988; Kavadia *et al.*, 1984).

To overcome the hazards of chemical pesticides, botanicals such as neem seed kernel extract, neem oil, soap water are now used in many developed and developing countries for combating this pest infestation with the aim of increasing crop yield (Hossain *et al.*, 2003; Mote and Bhavikatti, 2003; Singh and Kumar, 2003; Rao and Rajendran, 2002; Gahukar, 2000; Lawrence *et al.*, 1996). Shrestha *et al.*, (2010) suggested use of neem products and lantana products to protect plants against aphids. Neem extract, neem oil, neem seed carnel etc. are also effective to control epilachna beetle, leaf hopper and other sucking insects in brinjal field. To use these botanicals human health hazard become low and incidence of beneficiary insects remain hazard free, so that, they can control the insect pest of brinjal keeping the environment sound.

Keeping this perspective in view of the experiment was undertaken against sucking and foliage insects like leafhopper, aphid, epilachna beetle, leaf roller etc. to fulfill the following objectives:

- To find out the level of infestation caused by leafhopper and other sucking pest of brinjal in filed condition
- To reduce the infestation of leafhopper and other sucking pest of brinjal using botanicals and other insecticides in field condition and
- To find out the impact of botanicals and other chemical insecticides of beneficial arthropods in the field of brinjal during managements of leafhopper and other sucking pest of brinjal

4

# **CHAPTER II**

### **REVIEW OF LITERATURE**

Sucking insect pests are very much dangerous for most of the vegetable crops. Aphid, leafhopper, epilachna beetle and leaf roller are considered as major sucking insect pests of brinjal, which cause significant damage to crop every year. The incidence of those insects occurs sporadically or in epidemic form throughout Bangladesh and affecting adversely the quality and yield of the crop. In the favorable weather severe infestation may occur and total crop may be damaged. Literatures regarding their population dynamics and management in brinjal are scanty. However, review of the available literatures relevant to the present study is presented below under the following sub-headings.

#### 2.1. Sucking and foliage insect pests of brinjal

#### 2.1.1. Leafhopper

#### 2.1.1.1. Scientific classification

Kingdom: Animalia

Phylum: Arthropoda

Class: Insecta

Order: Hemiptera

Family: Cicadellidae

Genus: Amrasca

Specis: A. biguttula biguttula

#### 2.1.1.2. Origin and distribution

Although found throughout much of the United States east of the Rocky Mountains, the potato leafhopper only overwinters along the Gulf Coast. The insect undergoes mass movements northward in the spring and early summer and becomes established in many areas of the country. The potato leafhopper is generally distributed northward by wind. Although the potato leafhopper does not overwinter in northern areas, it may complete several generations in these areas.

It occurs in several countries including India, Bangladesh, China, Myanmar, North Africa, Pakistan, Philippines, Sri Lanka, and Taiwan. Relatively dry and humid weather favors population build-up (Padwal *et al*, 2016).

#### **2.1.1.3.** Host range

The potato leafhopper feeds on a variety of plant species and has been reported to feed on nearly 200 kinds of plants. Flowers attacked by the potato leafhopper include dahlia, rose, and sunflower. Ornamental trees that are hosts for this leafhopper are Chinese chestnut, elm, English walnut, flowering Japanese cherry, hickory, locust, oak, and redbud. Flowering fruit trees, e.g., crabapple, also have been reported as hosts of potato leafhopper. A few of the economic plants that are infested with this insect are alfalfa, apple, eggplant, peanut, potato, soybean, and sweet potato. Leafhopper is widely distributed in India and most destructive to American cotton in the north-western region. Besides cotton it also feeds, potato, brinjal and some wild plant like hollyhock, kangi buti, etc. (Atwal, 1986). The insect also invades cotton, lady's finger, tomato and many other malvaceous and solanaceous plants (Alam, 1969). Jacob *et al.* (2000) identified *A. devastans* on castor bean, which attained at pest status. A survey report in Madhya Pradesh, India revealed that *A. devastans* infested potatoes in that area (Dharpure, 2003). Mamun (2006) reported that leafhopper prefers tomato, sweet gourd, country bean, brinjal, okra and cotton as host.

#### 2.1.1.4. Seasonal abundance

6

Population of jassid varies in different time of the year. Alam *et al.* (2005) stated that jassid population was higher during the dry period especially mid February to mid April and number of jassid per leaf reached to its maximum (15.41) peak during the month of April. During long dry period especially in the month of February-March jassid became a serious problem for brinjal cultivation (Alam *et al.*, 2006). Mall *et al.* (1992) described that seasonal incidence of jassid was more prevalent during vegetative phase of the crop up to the 3rd week of September when the average temperature and humidity were more than 28°C and 80 percent respectively.

Early plantings are more injured by the cicadellidae than the late planting. The pest breed practically throughout the year but during the winter month only adult was found on plants such as potato, brinjal, tomato etc. In spring they migrate to okra and started breeding. Particularly the American cotton was very susceptible to this pest (Nair, 1986). The cotton jassid was formally considered to be on early season pest attacking plant in Bangladesh (Bohlen, 1984). Ali (1987) reported that jassid had been found to attack plants throughout the season. The incidence of jassid on brinjal planted at various dates from 20 July to 20 December was higher an early planted crop than on late planted crops (Borah, 1995). Seasonal abundance of cotton jassid on okra was dependent on meteorological parameters. Jassid population was maximum during middle of April (30.00 nymph/leaf) to last week of May (37.5 nymph/leaf). High temperature (30.36°C) evening relative humidity (below 80%) and low rainfall period coupled with bright sunshine hours were favourable for the development of cotton jassid population (Inee et al., 2000). Muthukumar and Kalyanasundaram (2003b) observed that jassid had a negative association with minimum temperature and rainfall when investigation on the seasonal incidence of jassid (Amrasca biguttula biguttula) population on okra and their correlation with abiotic factors were carried out kharif 1990 in the semiarid region in India. The infestation of jassid

started in the fourth week of July and reached peak in the second and fourth weeks of September respectively (Kumawat *et al.*, 2000). Studies on the seasonal incidence of jassid on cotton under rainfed conditions were conducted at Bharuch, and Gujarat in India, during 1979-80 and 1981-82. Results showed that population of *Amrasca biguttula biguttula* ranged from 0.59 to 2.78 per plant recorded in the second fortnight of November (Patel and Patel, 1998).

The spatial distribution of A. biguttula biguttula was studied in upland cotton in India. Environmental heterogeneity at low population in July and innate behaviour at high population were responsible for aggregated dispersion in the species of hemiptera (Singh et al., 1990). Ali and Karim (1991) investigated the influence of cotton plant age on the seasonal abundance of A. biguttula. They found that the insect remained below the economic threshold level of 1 insect/leaf for up to 35 days of plant age in kharif cotton and 65 days of plant age in rabi cotton. Most of the cicadellids were found in 35 to 75 days old cotton plants in kharif and 65 to 130 days old cotton plants in the rabi season. Cotton grown in the kharif season was more vulnerable to insect attack than cotton grown in the rabi season. The population of leaf hopper on brinjal was positively correlated with average maximum-minimum temperature, relative humidity and total rainfall (Shukla, 1989). Observation on the jassid population was made from the second week of July up to the third week of September. The insect population increased from July to August. The maximum activity of the insect occurred from the 1stweek to the middle of August. After this period, the jassid population gradually declined, probably due to the slight increase in atmospheric temperature and RH, maximum crop damage coincidence with the maximum activity of the pest (Poonia, 2005). Investigations on the seasonal incidence of jassid population on okra with abiotic factors were carried out during kharif 1996 in the semiarid region of Rajasthan, India. The infestation of jassid started in the 4th week of July and reached peaks in the 2nd and 4th weeks of September, respectively (Kumawat *et al.*, 2000).

#### 2.1.1.5. Biology of leafhopper

The adults mated two days after emergence and the eggs were laid two to seven days after copulation (Nair, 1986). Eggs were laid on the leaves of food plants and are hatched in about a week (Alam, 1969). Eggs were laid singly within leaf veins in the paranchymatous layer between the vascular bundles and the epidermis on the upper leaf surface. An average of 15 eggs (with a maximum of 29) was laid per female. Mature leaves (35-45 days old) were prefened for egg deposition; curved, greenish-yellow, eggs (0.7-0.9 x 0.15-0.2 mm) were laid, the egg period last for 4- 11 days (Nair 1986). The females deposited slender white eggs within the stems and larger veins of the leaves and hatching period was 6-9 days (Davidson, 1987). A female laid 25-30 eggs of 1:4 eggs per day, which were hatched in 4 to 11 days (Nair, 1986).

Nymphs were pale green, wedge shaped, 0.5-2.0 mm long, have a characteristics crab like, sideways movement when disturbed. They were confined to the under surface of leaves during the day time but found anywhere on the leaves at night. The nymphal period varied from 7 to 21 days depending on food supplies and temperature they passed through six stages of growth during nymphal period (Atwal, 1986). Another study revealed that they became full grown in seven days in autumn and 25 days in winters. Nair (1986) reported the five nymphal instars completed in 19-21days. Bohlen (1984) stated five nymphal instars and the nymphs resembled the adult but had no wings. The nymphs were smaller than the adult but wingless. Nymphs were found on the underside of leaves (FAO, 2003). The adult were small, elongate, wedge shaped, about 2.5 mm long, body pale green semi-transparent wings very active having aside way walk

like the nymph, but quick to hop and fly when disturbed. The adult of the summer brood were greenish yellow in color and those of the winter broad radish. Unmated adults lived for 3 month or more, when mated, they lived five weeks in summer and seven weeks in winter. Life cycle was completed in 15-46 days in the different seasons and up to eleven generation was completed in a year (Nair, 1986). They were also attracted to light at night (Atwal, 1986). Adults were usually less than 13 mm long with slender, tapered bodies of various colors from bright grey to yellow green with shiny wings and had two distinct black spot at distal found on the foliage in large numbers and moved around by jumping but flew very rapidly when disturbed. The adults were found on the under sides of the leaves (FAO, 2003).

#### 2.1.1.6. Nature of damage

Jassid, *Amrasca devastans* infestation was manifested by some characteristic symptoms. The primary symptom was characterized by leaf edge curling and the secondary symptom was characterized by leaf edge curling along with leaf edge and vein colouring and drying of the leaves. From the initial infestation, the symptoms developed in sequence leading to hopper burn and shedding of leaves in severe cases of infestations, which ultimately caused the retraction of plant growth and reduction of yield (Afzal and Ghani, 1953). Nair (1986) reported that the nymphs and adults of *A. biguttula biguttula* attacked host leaves at all stages of development. The adults and nymphs feed on the sap and injected saliva into the tissues, which caused toxemia and injury of the leaves. The edges of the infested leave turned pale-green, then yellow and finally brick red brown in colour. The colour changes were accompanied by severe crinkling and curling of the leaf. The whole leaf gradually dried up and dropped. The plant became stunted and quality of fruit was also affected. El-Tom (1987) reported that cotton jassid; *A. biguttula biguttula* was one of the key pests of cotton and in the major factor limiting cotton yield in

Bangladesh. This pest caused more than 50% reduction of seed cotton yield in some cotton genotypes (Bhat *et al.*, 1984).

The jassid while sucking the plant sap injected some toxic substances with saliva into the cotton plants. Time required to development characteristic jassid damage symptoms in cotton plants were found positively corelated with age of the plant. The younger plants were found susceptible to jassid attack than the older plants (Nayer *et al.*, 1984). As the plants grew older they became less susceptible to jassid infestation (Ali, 1990). Rote *et al.* (1985) reported a significant positive correlation between jassid damage symptoms and jassid population levels on the plant. Yield losses of cotton due to sucking pests (*Amrasca biguttula biguttula*) were evaluated during the rainy season of 1985 and 1986 in Karnataka, India. The average yield loss was 46.41% (Panchabhavi *et al.*, 1990).

#### 2.1.2. Epilachna beetle

#### 2.1.2.1. Scientific classification

Kingdom: Animalia

Phylum: Arthropoda

Class: Insecta

Order: Coleoptera

Family: Coccinellidae

Genus: Epilachna

Specis: E. vigintioctopunctata

#### 2.1.2.2. Origin and distribution

South Canada, USA, Mexico, Guatemala, Africa and South East Asia. It occurs in Russia, China, Japan, and Korea. This species is native to southeastern Asia, primarily India, but has been

accidentally introduced to other parts of the world, including Australia and New Zealand. It has also been recorded from Brazil and Argentina, beginning in 1996 (CSIRO, 2005).

#### 2.1.2.3. Host range

Brinjal, potato, tomato, cucurbitaceous plants, wild solanaceous plants.

#### 2.1.2.4. Natural abundance

This species causes damage to agricultural crops primarily in the family Solanaceae, especially potatoes; other crops include pumpkin, turnips, radishes, beans and spinach.

#### 2.1.2.5. Life cycle

Egg period: 2-4 days. Cigar shaped, laid in clusters on lower leaf surface, yellow; 120-460 eggs/female. Grub: 10-35 days. Yellowish bearing six rows of longitudinal spines. Pupa: 5-6 days. Yellowish with spines on posterior part; anterior portion being devoid of spines. Pupates on the stem or leaves. Adult *E. dodecastigma*: Copper-coloured, 6 spots / elytra *E. demurille*: Dull appearance, light copper coloured and six black spots surrounded by yellowish area on each elytra. *E. vigintioctopunctata*: 14 spots on each elytra, deep red. Total life period: 20-50 days. 7 generations / year.

The orange and black spotted adults are about 7-10 millimetres long. The head, prothorax (first part of the middle body) and elytra (wing covers) are covered with short fine hairs. The elytra are covered with 28 spots. The size and shape of the spots is variable, but only the pairs of spots by the mid line of the second and fourth transverse rows may join each other. The underside of the ladybird orange-brown and black. There are three pairs of orange-brown legs. Under the elytra is a pair of wings used for flying. The small head is mainly pale orange and has a pair of compound eyes and two short antennae. The antennae are orange-brown.

Female ladybirds lay clusters of yellow eggs near infestations of prey. A larva hatches from each egg. There are four larval instars (stages). As the larva grows, it moults (changes skin). The newly hatched larva is pale yellow and covered with tubercles with long seta. The body remains yellow and the tergites, tubercles, setae and legs become dark grey. There are three pairs of legs. Larvae also use the tip of the abdomen for holding onto the substrate on which they are walking. The tip of the abdomen also holds the larva to the surface during moulting both to another larval instar and to a pupa. When the fourth larval instar is fully grown, it attaches itself to a sheltered place on a plant. The spiny skin of the larva remains attached to the base of the pupa. The pupa is covered in black setae. It is black except for the pale inter-segmental membranes. There are prominent white tubular abdominal spiracles, openings to the air ducts (trachea). Adults hatch from pupae and mate. The length of time of each life stage depends on temperature, being shorter at higher temperatures (Martin, 2018).

#### 2.1.2.6. Nature of damage

Both adult and grubs scrap the lower epidermis of leaves in characteristic manner leaving behind stripes of uneaten areas. The leaves give a stifled appearance. In severe infestation all leaves may be eaten off leaving only the veins intact (Skeletonization) and plants may wither.

#### 2.1.3. Aphid

#### 2.1.3.1. Scientific classification

Kingdom: Animalia Phylum: Arthropoda Class: Insecta Order: Hemiptera

Family: Aphididae

Genus: Aphis

Specis: A. fabae

#### 2.1.3.2. Origin and distribution

Aphids are distributed worldwide, but are most common in temperate zones. In contrast to many taxa, aphid species diversity is much lower in the tropics than in the temperate zones (Zyla *et al.*, 2017). They can migrate great distances, mainly through passive dispersal by riding on winds. For example, the currant lettuce aphid, *Nasonovia ribisnigri*, is believed to have spread from New Zealand to Tasmania in this way (Pip Courtney, 2005). Aphids have also been spread by human transportation of infested plant materials.

Winged aphids may also rise up in the day as high as 600 m where they are transported by strong winds (Berry and Taylor, 1968; Isard *et al.*, 1990). For example, the currant-lettuce aphid, *Nasonovia ribisnigri*, is believed to have spread from New Zealand to Tasmania around 2004 through easterly winds (Hill, L. 2012). Aphids have also been spread by human transportation of infested plant materials, making some species nearly cosmopolitan in their distribution (John, *et al.* 2009).

The black bean aphid may have originated in Europe and Asia, but it is now one of the most widely distributed species of aphids. It is found throughout temperate areas of Western Europe, Asia, and North America and in the cooler parts of Africa, the Middle East, and South America (AphID, 2012). In the warmer parts of its range, apterous individuals can survive the winter and they may continue to reproduce asexually all year round (HYPP, 2013). It is known to be migratory (Johnson, C. G., 1963).

#### 2.1.3.3. Host range

The black bean aphid can feed on a wide variety of host plants. Its primary hosts on which the eggs overwinter are shrubs such as the spindle tree (*Euonymus europaeus*), Viburnum species, or the mock-orange (*Philadelphus species*). Its secondary hosts, on which it spends the summer, include a number of crops including sugar beets, spinach, beans, runner beans, celery, potatoes, sunflowers, carrots, artichokes, tobacco, and tomatoes. It colonize more than 200 different species of cultivated and wild plants. Among the latter, it shows a preference for poppies (*Papaver species*), burdock (*Arctium tomentonum*), fat-hen (*Chenopodium album*), saltbush (*Atriplex rosea*), chamomile (*Matricaria chamomilla*), thistles (*Cirsium arvense*) (Berim, M. N., 2009), and docks (*Rumex* spp.) (RIR, 2013).

Two conflicting factors are involved in host preferences, the species and the age of the leaf. Offered spindle and beet leaves on growing plants throughout the year, winged aphids moved from one to the other depending on the active growth state of each and the senescence of each host plant. Thus, in late summer and autumn, the beet leaves were old and unattractive to the aphids in comparison with the leaves of the spindle, whereas in spring, the young unfolding leaves of the beet were more attractive than those of the spindle (Kennedy and Booth, 1951).

#### 2.1.3.4. Life cycle

The black bean aphid has both sexual and asexual generations in its life cycle. It also alternates hosts at different times of year. The primary host plants are woody shrubs, and eggs are laid on these by winged females in the autumn. The adults then die and the eggs overwinter. The aphids that hatch from these eggs in the spring are wingless females known as stem mothers. These are able to reproduce asexually, giving birth to live offspring, nymphs, through parthenogenesis (Chinery and Michael, 1993). The lifespan of a parthenogenetic female is about 50 days and during this period, each can produce as many as 30 young (Berim, M. N., 2009). The offspring

are also females and able to reproduce without mating, but further generations are usually winged forms. These migrate to their secondary host plants, completely different species that are typically herbaceous plants with soft, young growth (HYPP, 2013; Chinery and Michael, 1993; Berim, M. N., 2009).

Further parthenogenesis takes place on these new hosts on the undersides of leaves and on the growing tips. All the offspring are female at this time of year and large populations of aphids develop rapidly with both winged and wingless forms produced throughout the summer. Winged individuals develop as a response to overcrowding and they disperse to new host plants and other crops. By midsummer, the number of predators and parasites has built up and aphid populations cease to expand (RIR, 2013). As autumn approaches, the winged forms migrate back to the primary host plants. Here, both males and sexual females are produced parthogenetically, mating takes place, and these females lay eggs in crevices and under lichens to complete the lifecycle. Each female can lay six to ten black eggs which can survive temperatures as low as  $-32 \,^{\circ}C$  ( $-26 \,^{\circ}F$ ) (HYPP, 2013; Chinery and Michael, 1993; Berim, M. N., 2009). More than 40% of the eggs probably survive the winter, but some are eaten by birds or flower bugs, and others fail to hatch in the spring (Way and Banks, 1964).

#### 2.1.3.5. Nature of damage

The black bean aphid is a major pest of sugar beet, bean, and celery crops, with large numbers of aphids cause stunting of the plants. Beans suffer damage to flowers and pods which may not develop properly. Early-sown crops may avoid significant damage if they have already flowered before the number of aphids builds up in the spring (RIR, 2013). Celery can be heavily infested. The plants are stunted by the removal of sap, the stems are distorted, harmful viruses are transmitted, and aphid residues may contaminate the crop (Godfrey and Trumble, 2009). As a

result of infestation by this aphid, leaves of sugar beet become swollen, roll, and cease developing. The roots grow poorly and the sugar content is reduced. In some other plants, the leaves do not become distorted, but growth is affected and flowers abort due to the action of the toxic saliva injected by the aphid to improve the flow of sap (HYPP, 2013).

To obtain enough protein, aphids need to suck large volumes of sap. The excess sugary fluid, honeydew, is secreted by the aphids. It adheres to plants, where it promotes growth of sooty molds. These are unsightly, reduce the surface area of the plant available for photosynthesis and may reduce the value of the crop. These aphids are also the vectors of about 30 plant viruses, mostly of the non-persistent variety. The aphids may not be the original source of infection, but are instrumental in spreading the virus through the crop (RIR, 2013). Various chemical treatments are available to kill the aphids and organic growers can use a solution of soft soap (Godfrey and Trumble, 2009).

#### 2.1.4. Leaf roller

#### 2.1.4.1. Scientific classification

Kingdom: Animalia

Phylum: Arthropoda

Class: Insecta

Order: Lepidoptera

Family: Tortricidae

Genus: Archips

Specis: A. semiferanus

#### 2.1.4.2. Origin and distribution

*Archips semiferanus* is found in the eastern United States and adjoining portions of southeastern Canada. It has been found in US states including Connecticut, Massachusetts, New York, Pennsylvania, Texas, Virginia, and West Virginia (USDAFS, 1998; Drees, *et al.*, 2009). They may have been introduced to the United Kingdom (Walker, 1863). In 2001 recent outbreaks had occurred in Cambria, Cameron, Clearfield, Clinton, and Warren counties in Pennsylvania (John, Y. 2001).

#### 2.1.4.3. Host range

Fruit tree leaf roller feeds on a wide variety of deciduous trees and shrubs are hosts, including apple, crabapple, honey locust, ash and linden. Oak leaf roller is associated with Gambel oak; boxelder leaf roller with boxelder.

#### 2.1.4.4. Life cycle

Oak leaf roller moths lay their eggs in July each year, in groups of 40 to 50. The female covers the eggs with hairs from her body; they are deposited on "the base of large branches and rough bark patches on both tree trunks and limbs" (Talerico *et al.*, 1978). The flat egg masses are white-gray in color and oval shaped, and are about 4.8 millimetres (0.19 in) across. The eggs overwinter and hatch in spring of the next year. The larvae (or caterpillars) emerge in April and initially eat the buds of oak trees and young leaves inside them (Talerico *et al.*, 1978; USDAFS, 2011).

When fully grown, the larvae are between 25 to 29 millimetres (0.98 to 1.14 in) long with a body that can be yellow-green or darker shades of green. Other identifying characteristics in the larvae include pale legs and a head that is either black or has "a dark eye patch or a dark bar" (Talerico *et al.*, 1978; USDAFS, 2011; USDAFS, 1998). The larvae feed and nest inside leaves which they

18

have rolled or folded until they are ready to pupate in mid-June. The larvae pupate in cocoons which are found inside the rolled leaves or in "bark crevices" (USDAFS, 2011).

After a week or two in the pupal stage, the adult moths emerge in late June or early July. The moths are small with a wingspan of 18 to 22 millimetres (0.71 to 0.87 in); the wings have a characteristic bell shape (USDAFS, 1998). Wing color can vary considerably. Forewings are a mixture of "creamy brown and gray" with gray found at the wingtips (Talerico *et al.*, 1978). The forewings have a darker band of brown or gray crossing obliquely. The adults mate and lay eggs to start the next generation. The moths produce only one generation annually (Talerico *et al.*, 1978; USDAFS, 2011).

In Texas, the timing of the various stages of the life cycle starts earlier, and other differences in behavior are seen. Since spring comes earlier in Texas, the eggs are laid in May and hatch in mid-March of the next year. The larvae can be dislodged from trees and dangle beneath them from silk threads. Although the larvae cannot harm humans, most people in Texas will avoid walking under oak trees to avoid them. The pupae of oak leaf rollers in Texas are also found on branch tips and weeds near the tree (Drees, *et al.* 2009).

#### 2.1.4.5. Nature of damage

Larvae are active early in the season and chew leaves of a wide variety of plants. Older larvae have the habit of curling over the edge of leaves and fastening with silk to create a rolled leaf shelter. Damage by leaf rollers is usually transitory and mostly cosmetic. However, oak leaf roller, often in combination with species such as the oak looper and speckled green fruit worm, have caused episodes of extensive defoliation to native oak stands.

The caterpillars, which are range from pale to dark green have a black head. They are usually found within the folded leaves where they feed, chewing in a skeletonizing manner. When

19

disturbed they can move vigorously and often will drop out of the leaf on a strand of silk. On fruit trees larvae of the fruit tree leaf roller may chew pits in developing fruit causing them to prematurely drop or grow in a distorted manner.

#### 2.2. Management

The management of (*Amrasca biguttula biguttula*) through various non-chemical method namely, cultural, mechanical, biological and host plant resistant etc. was limited throughout the world. The research work on non-chemical control measure of this insect pest was also scanty. The farmers of Bangladesh usually apply six to eight schedule based insecticide sprays against this insect pests throughout the season. But this kind of insect pests control strategy relying solely on chemical protection had got many limitations and undesirable side effects (Hussain 1993, 1984) and this in the long run led to many insecticides related complications (Frisbie, 1984) such as direct toxicity to beneficial insects, fishes and other nontarget organism (Munakata, 1997; Goodland *et al.*, 1985, Pimentel, 1981), human health hazards (Bhaduri *et al.*, 1989) resurgence of pests (Husain, 1993; Luckmann and Metcalf, 1975) outbreak of secondary pest (Hagen and Franz, 1973) and environmental pollution (Fishwick, 1988; Kavadia *et at.*, 1984). To overcome these problems botanical insecticide soaprvater and water are now used in many developed and developing countries for combating this pest infestation with the aim of increasing crop yield.

*Verticillium lecanii*. In sustainable agriculture, prevention strategies are one of the most important tactics that growers can employ to avert aphid infestation. These include cultural techniques such as use of physical barriers, removal of crop in space and time, mulching, crop rotation, border crops and cover crops. Both synthetic and living mulches have been shown to reduce population of alate aphids reaching/landing on plants and hence reducing the incidence of aphid-transmitted viruses. Crops receiving high levels of nitrogen are more susceptible (attractive) to aphids; therefore, slow release fertilizers may be an alternative to avoid high aphid infestations. Aphids receive visual cues to land on crops when there is a clearly defined contrast in color between tilled bare soil and the lush foliage of crops. Living mulches reduce the contrast between the bare ground and the plant foliage so aphids do not detect their host. These mulches provide additional feeding sites for viruliferous aphids (aphids carrying virus) around the crop and hence reduce the incidence and spread of aphid-borne non-persistently transmitted viruses (Toba *et al.*, 1977).

#### 2.2.1. Plant extracts for sucking insect pests management

Johnson (2001) assigned that, mealybugs can be controlled using the biocontrol agent, e.g. Botanical pesticides are the most cost effective and environmentally safe inputs in Integrated Pest Management (IPM) strategies. There were about 3000 plants and trees with insecticidal and repellant properties in the world, and India was home to about 70 percent of this floral wealth (Narayanasamy, 2002). He stated the use of more than 450 botanical derivatives used in traditional agricultural system and neem was one of the well-documented trees, and almost all the parts of the tree had been found to have insecticidal value. The neem seed kernel extract, neem oil, extracts from the leaves and barks had all been used since ancient times to keep scores of insect pests away. A number of commercial neem based insecticides were now available and they had replaced several toxic chemical insecticides. The extracts were of particular value in controlling the sucking and chewing insect pests. The young caterpillars devouring the tender leaves were well managed by the botanical insecticides. The plant materials should be thoroughly washed before preparing the extract and the right quantity should be used. Pink

mealybug infests the mulberry plants and cause Tukra diseases that leads to qualitative loss of leaves. Hence a study was carried out to evaluate the efficacy of various indigenous native plant extracts for their repellency property against pink mealybug, *Maconellicoccus hirsutus* (Green) at the Tamil Nadu Agricultural University, Coimbatore. The native botanicals such as Andrographis leaf extract, Leucas leaf extract, Neem seed kernel extract, vitex leaf extract, fish oil rosin soap, ocimum leaf extract and lawsonia leaf extract at different dose levels viz., 1, 2, 4, 8 and 10 percent respectively. After 48 hours (Hour of release) the highest repellency was recorded in case of Andrographis leaf extract (99.0%) followed by Leucas leaf extract and NSKE (99.0%). Vitex leaf extract and FORS showed on par results among various treatments. The ocimum leaf extract (90.1%) also recorded a moderate repellent effect and the least repellency was recorded in case of Lawsonia leaf extract (81.3%). Similar trend was recorded during 24 hour of release also. As the dose increases the repellent effect also increased irrespective of the native botanical extracts against mealybugs (Sathyaseelan and Bhaskaran, 2010).

There are various insecticides that can be used to control aphids. Nowadays, there are many plant extracts and plant products that are eco-friendly and control aphids as effectively as chemical insecticides. Shreth *et al.*, suggested use of neem products and lantana products to protect plants against aphids (Chongtham *et al.*, 2009). For small backyard infestations, simply spraying the plants thoroughly with a strong water jet every few days is sufficient protection for roses and other plants.

With the continued robust growth of the global bio-pesticide market, Azadirachtin is uniquely positioned to become a key insecticide to expand in this market segment. In the USA, actual or impending cancellation of some organophosphate and Carbamate insecticides that had either lost patent protection or were not being re-registered in many markets because of the food quality

22

protection Act of 1996, had opened new opportunities for bio-pesticides and reduced risks of pesticides in general. The broad-spectrum activity of Azadirachtin at low use rates (125-140g a.i. ha<sup>-1</sup>) coupled with the insect growth regulator activity (in all larval /nymphal instars including the pupal stages) and unique mode of action (ecdysone disruptor) made Azadirachtin an ideal candidate for insecticide resistance, integrated pest control and organic pest control programs. The pest control potential demonstrated by various extracts and compounds isolated from the kernels and leaves of the neem plant (Azadirachta indica, (Meliaceae)) seem to be of tremendous importance for agriculture in developing countries. Laboratory and field trial data had revealed that neem extracts were toxic to over 400 species of insect pests; some of which had developed resistance to conventional pesticides, e.g. sweet potato whitefly (Bemisia tabaci Genn. Homoftera: Aleyrodidae), the diamond back moth (Plutella xylostella L. Lepidoptera: Plutellidae) and cattle ticks (Amblyomma cajennense F. Acarina: Ixodidae and Tsoaphilus microplus Canestrini. Acarina: Ixodidae). The compounds isolated from the neem plant manifested their effects on the test organisms in many ways, e.g. as antifeedants, growth regulators, repellents; toxicants and chemosterilants. This review strived to assess critically the pest control potential of neem extracts and compounds for their use in the tropics. This assessment was based on the formulation, stability and phytotoxicity information available on the wide range of pests against which neem extracts and compounds had proven to be toxic, toxicity to non-target organisms, e.g. parasitoids, pollinators, mammals and fish. (Lawrence et al., 1996).

Azadirachtin had been exempted from residue tolerance requirements by the US environmental protection agency for food crop applications. It exhibited good 20 efficacy against key pests with minimal to no impact on non-target organisms. It was also compatible with other biological

23

control agents and had a good fit into classical integrated pest management programs (John and Immaraju, 1997). Products derived from leaves and kernels of neem (Azadirachta indica) are becoming popular in plant protection programs for cotton, mainly because synthetic pesticides have several undesirable effects. Neem products acted both as systemic and as contact poisons and their effects were antifeedant, toxicological, repellent, sterility inducing or insect growth inhibiting. Furthermore, neem products appeared to be environmentally safe and IPM compatible and had the potential to be adopted on a broad scale, together with other measures, to provide a low cost management strategy (Hillocks, 1995; Gahukar, 2000). Indigenous plant materials were cheaper and hazard free in comparison to chemical insecticide (Saxena et al., 1992). These were also easily available in everywhere in our country. Ofori and Sackey (2003) reported that acetylic, aqueous neem seed extract reduced the Amrasca biguttula on okra. The biological control agents Bacillus thuringiensis (Bt; Delfin 85 WG) at 0.04% and Trichograma chilonis at 60000/ha and insecticides Azadirachtin (Econeem) at, 0.0006%, Lufenuron (Match 5EC) at 0.005%, Avermectin (Vertimec [Abamectin] at 0.0004%, Monocrotophos 36SL (Monocil) at 0.05%, Spark 36EC (Detramethrin IEC + Triazophos 35EC) at 0.05%, Bulldock star 262.5EC (Beta-cyfluthrin12.5EC + Chlorpyrifos 250 and Nurelle-D.505. 55EC Cypermethrin 5 + Chlorpyrifos 50) at 0.05% were tested in a field trial in Rahuri, Maharashtra, India, ;ring the kharif season of 2000 against pest complex of brinjal. Azadirachtin was moderately effective against the sucking pest including Bemisia tabaci, Aphis gossypii, Amrasca biguttula biguttula (Mote and Bhavikatti, 2003).

The joint action potential of methanoic extract of neem seed kernel (*Azadirachta indica*) in combination with methanolic extracts of two other botanical, viz. sweet flag (*Acorus calamus*) and *Pongamia glabra* (*P. pinnata*) against *Ammrasca devastans* at 1:1:1,2:1:1 and 3:1:1 (v/v)

ratio were studied. This combination at 0.42% concentration gave superior control of A. devastans (Rao and Rajendran, 2002). An experiment was conducted with okra in India to determine the efficacy of neem based pesticide against the cotton jassid, A. biguttula. The treatments comprised Endosulfan at 0.07%, A Chook at 3% Neemarin at 0.7%, neem seed kernel extract (NSKE) at 1%, NSKE at 3% with an untreated control. Endosulfan followed by A Chook and NSKE (3%) were most effective in controlling the okra jassid. A Chook treated plots gave the highest yield of 50.06 q/ha and significantly superior to other treatments. However on the basis of cost benefit ratio NSKE (3%) ranked first (Singh and Kumar, 2003). Schneider and Madel (1992) reported that the treatments of neem seed kernel extract (NSKE) did not show a significant reduction in parasitization rate of fecundity of larval parasitoid, Diadegma semiclausum. The aqueous NSKE had no adverse effects on D. semiclausum following dfuect contact. Patel and Patel (1998) reported that application of Quinalphos and Triazophos resulted in a resurgence of A. biguttuta on okra and abergine (Brinjal), while Endosulfan at 0.07% and Repelin (based on Azadirachta indica) 1% were highly effective. Nandagopal and Soni (1992) observed that in India neem oil was least persistant insecticides and caused >50% mortality of jassid only up to 24 hours after application. Different concentrations of soap solution were applied against jassid of cotton. Soap powder (25gm/liter of water) predominantly reduced the pest population during the period and harvested the best yield than other treatments economic return is reasonably satisfied (Hossain et al., 2003).

# **CHAPTER III**

### **MATERIALS AND METHODS**

The experiment on the population dynamics and management of sucking insect pests in brinjal, Homoptera, Cicadellidae was carried out at the experiment field of the Sher-e-Bangla Agricultural University (SAU), Dhaka, Bangladesh during November, 2017 to March, 2018. The materials and methods adopted in this study are discussed in the following sub headings:

#### 3.1. Location

The experimental site was located at the experimental farm of Sher-e-Bangla Agricultural University, Dhaka-1207, during the period from November, 2017 to March, 2018. The experimental field was located at  $90^{\circ}335$ ' east longitude and  $23^{\circ}$  774' north latitude at a height of 4 meter above the sea level. The land was medium high and well drained.

#### 3.2. Climate

The experimental site was situated in the sub-tropical climatic zone, characterized by lower rainfall during the month of November 2017 to March 2018. Monthly maximum and minimum temperature, relative humidity and total rainfall recorded during the period of study at the SAU experimental farm have been presented in the Appendix I. The recorded and calculated as monthly average temperature, relative humidity and rainfall for the crop growing period of experiment were noted from the Bangladesh Meteorological Department (climate division), Agargaon, Dhaka-1207 and has been presented.

#### 3.3. Soil

The soil of the study was silty clay in texture. The area represents the agroecological zone of "Madhupur tract" (AEZ No. 28). Organic matter content was very low (0.82%) and soil pH varied from 5.47 to 5.63.

### **3.4. Design and layout**

The study was conducted considering eight treatments including a control for controlling sucking pest at seedling to harvesting stage. The experiment was laid out in a Randomized Complete Block Design (RCBD) with three replications in the field of the Entomology Department. The whole field was divided three blocks of equal size and each block was sub divided into nine plots. The unit plot size was  $3m \times 2m$  accommodating twelve pits per plot. The distance between row to row was 100 cm and that of the plants to plants was 70 cm.

# **3.5. Land preparation**

The soil of the experimental field was well prepared thoroughly followed by plowing and cross plowing, leveling and laddering to have a good tilth. All weeds and debris of previous crops were removed and land was finally prepared with the addition of basal dose of well decomposed cow dung. The plots were raised by 10 cm from the soil surface keeping the drain around the plots.

### **3.6.** Manuring and fertilization

The following doses of manure and fertilizers were applied as per recommendation of Rashid (1999) for brinjal.

| Manure/ Fertilizers          | Dose per ha |
|------------------------------|-------------|
| Cow-dung                     | 10 tons     |
| Urea                         | 360 Kg      |
| Triple Super Phosphate (TSP) | 150 Kg      |
| Muriate of Potash            | 250 Kg      |

The full dose cow-dung and TSP were applied as basal dose during final land preparation. Onethird of the MP and urea were applied in the pits one week before transplanting and rest of the MP and urea were applied as the top dressing at 21, 35 and 50 days after transplanting.

## 3.7. Raising of seedling and transplanting

Brinjal seed (Vatiety: BARI brinjal-1, Uttara) were collected from BARI, Gazipur, Dhaka. A small seedbed measuring  $5m \times 1m$  was prepared and seeds were sown in the nursery bed at SAU Entomology field on 17 November 2017. Standard seedling raising practice was followed (Rashid, 1999). The plots were lightly irrigated regularly for ensuring seed proper development of the seedlings. The seedbed was mulched for ensuring proper seed germination, proper growth and development of the seedlings. Thirty-days-old healthy seedlings were transplanted in polybag for hardening. After twenty days that seedlings were transplanted on 29 December 2017 in the experimental field.

## **3.8. Intercultural operations**

### 3.8.1. Gap filling

At the time of transplanting a few seedlings were transplanted in the border of the experimental plots for gap filling. Very few numbers of seedlings were damaged after transplanting and such seedling were replaced by healthy seedlings from the same planted earlier on the border of the experimental plot. The seedlings were transplanted with a mass of soil roots to minimize the transplanting shock.

### **3.8.2 Irrigation**

After transplanting light irrigation was given to each plot. Supplementary irrigation was applied at an interval of 2-3 days. Stagnant water was effectively drained out at the time of over irrigation. The urea was top dressed in three splits as mentioned earlier.

### 3.8.3 Weeding

Weeding was done as and when necessary to break the soil crust and to keep the plots free from weeds. First weeding was done after 20 days of planting and the rest were carried out at an interval of 15 days to keep the plot free from weeds.

# **3.8.4** Earthing up

Earthing up was done in each plot to provide more soil at the base of each plant. It was done 40 and 60 days after transplanting.

### **3.9.** Treatment for control measures

The experiment was evaluated to determine the efficacy of different botanical products and some chemical insecticides to compare with each other in considering the less hazardous but effective control measures against major insect pests, such as leaf hopper, aphid, epilachna beetle, leaf roller etc. of brinjal. The botanical based treatments and chemical insecticides as well as their doses to be used in the study are given bellow:-

 $T_1$  = Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval

 $T_2$ = Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval

 $T_3$ = Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days interval

 $T_4$ = Spraying of Marshal 25 EC@ 3.0 ml/L of water at 7 days interval

T<sub>5</sub>= Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval

 $T_6$ = Spraying of Imitaf 20 SL @ 0.1 ml/L of water at 7 days interval

 $T_7$ = Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval

 $T_8$ = Untreated control.

### **3.10.** Treatment preparation

### 3.10.1. Neem oil

The Neem oil was collected from Chawkbazar, Dhaka and the trix liquid detergent was collected from the local market of Agargoan bazaar, Dhaka. All sprays were made according to the methods described earlier. For each neem oil application, 15 ml neem oil (@ 3.0 ml/L of water i.e. 0.3% per 5 liter of water was used. The mixture within the spray machine was shacked well and sprayed on the upper and lower surface of the plants of the treatment until the drop run off from the plant. Three liters spray material was required to spray in three plot of each replication.

## 3.10.2. Neem seed kernel

The mature and dried neem seeds were collected from the neem tree found in the Horticulture garden of SAU. Then seeds were roasted by electric oven. Then the seed kernel was separated and taken into the electric blender for blending. 250 gm of neem seed kernel powder was taken into a beaker and 250 ml water was added into the beaker. Then the beaker was shaken by electric stirrer for mixing up thoroughly the mixture. The aqueous mixture then filtered using Whatmen paper filter and preserved the aqueous extracts of neem seed kernel in the refrigerator at  $4^{\circ}$ c for spraying in the field.

### **3.10.3. Bioneem plus**

The bioneem plus was collected from Chawkbazar, Dhaka and the trix liquid detergent was collected from the local market of Agargoan bazaar, Dhaka. All sprays were made according to the methods described earlier. For each bioneem plus application, 15 ml bioneem plus (@ 3.0 ml/L of water i.e. 0.3% per 5 liter of water was used. The mixture within the spray machine was shacked well and sprayed on the upper and lower surface of the plants of the treatment until the

drop run off from the plant. Three liters spray material was required to spray in three plot of each replication.

# **3.11. Treatment application**

- T<sub>1</sub>: Neem oil @ 3.0 ml/L of water was sprayed at 7 days interval. Under this treatment, neem oil was applied @ 15 ml /5L of water mixed with trix liquid detergent @ 10 ml (1%) to make the oil easy soluble in water. After proper shaking, the prepared spray was applied with a high volume knap-sack sprayer at 7 days intervals commencing from 20 DAT.
- T<sub>2</sub>: Neem seed kernel extract @ 3.0 ml/L of water was sprayed at 7 days. Under this treatment, neem seed kernel extract was applied @ 15 ml /5L of water. After proper shaking, the prepared spray was applied with a high volume knap-sack sprayer at 7 days intervals commencing from 20 DAT.
- T<sub>3</sub>: Bioneem plus @ 3.0 ml/L of water was sprayed at 7 days interval. Under this treatment, bioneem plus was applied @ 15 ml /5L of water mixed with trix liquid detergent @ 10 ml (1%) to make the oil easy soluble in water. After proper shaking, the prepared spray was applied with a high volume knap-sack sprayer at 7 days intervals commencing from 20 DAT.
- T<sub>4</sub>: Marshal 25 EC @ 3.00 ml/L of water was sprayed at 7 days interval. For this treatment 15.0 ml of insecticides per 5 liter of water was mixed and sprayed at 7 days intervals commencing from 20 DAT.
- T<sub>5</sub>: Actara 25 WG @ 0.2 gm/L of water was sprayed at 7 days interval. For this treatment 1.0 gm of insecticides per 5 liter of water was mixed and sprayed at 7 days intervals commencing from 20 DAT.

- T<sub>6</sub>: Emitaf 20 SL @ 0.1 ml/L of water was sprayed at 7 days interval. For this treatment 1.0 gm of insecticides per 5 liter of water was mixed and sprayed at 7 days intervals commencing from 20 DAT.
- T<sub>7</sub>: Ripcord 20 EC @ 1.0 ml/L of water was sprayed at 7 days interval. For this treatment 5.0 ml of insecticides per 5 liter of water was mixed and sprayed at 7 days intervals commencing from 20 DAT.

T<sub>8</sub>: Untreated control. There was no any control measure was applied in brinjal field.

### **3.12. Data collection**

Data were collected some pre-selected parameters like number of leaves and branches per plant, number of fruits/plant, fruit length and diameter, weight of individual fruits and fruit yield of brinjal. The number of aphid, leaf hopper, epilachna beetle and leaf roler was counted from treated and untreated plots of brinjal throughout the cropping season starting from 20 days after transplanting. Adults and nymphs of sucking insects were counted from a 30 random sample of five plants taken from each plot. Five leaves were chosen randomly from each plant, two from the bottom (older leaves), one from the middle and two from the top (younger leaves). The lower surface of the leaf was thoroughly examined for the presence of insects. Counts were made before 08.00 hr (Bangladesh local time) to avoid the excessive mobility of the adult insects after this time, but nevertheless, the migration of the fast moving and mobile adults from one plot to the other could not be totally avoided. The data were pooled over the cropping season and the population density was expressed as number of individuals per five leaves of the plant selected from each plot and tagged. The selected plants were observed regularly at weekly intervals in the morning. Healthy and infested leaves and number of branches per/plant were counted for estimating the infestation intensity. The data were converted to mean healthy and infested leaves

and number of braches per plant. Fruits were harvested at 7 days intervals and the number of fruits was recorded for each plot. Twenty fruits were selected randomly from each plot; length, diameter and weight of the selected fruit were recorded at each harvest. The data were pooled over the cropping season; length, diameter and weight of individual fruit were estimated. The total weight of fruit was recorded every harvest, right from the beginning of the first harvest and continued until the end of the growing season. The cumulative yield of fruits per plot from 12 harvests was calculated and it was then expressed as t/ha. The population of spiders and lady bird beetles were counted by randomly selected five branches from 5 plants of each plot at weekly interval. Assessment of treatment effects the performance profile of each treatment was judged by the reduction of the insect population densities in the treated plots and it was further confirmed by the comparison to yield contributing characters such as number of leaves and branches per plant, number of fruits/plant, fruit length and diameter, weight of individual fruits and yield obtained in each case at the end of the cropping cycle.

### The infested leaves were calculated by the following procedure:

Number of infested leaves was counted from total leaves per five plants and percent leaf infestation by sucking and foliage insect pests of brinjal were calculated as follows:

% Infestation of leaves by number =  $\frac{Number \ of \ infested \ leaves}{Total \ number \ of \ leaves} \times 100$ 

### The infested plants were calculated by the following procedure:

Number of infested plants was counted from total plot and percent plant infestation by sucking and foliage insect pests of brinjal were calculated as follows:

% Infestation of plants by number =  $\frac{Number \ of \ infested \ plants}{Total \ number \ of \ plants} \times 100$ 

# Percent edible fruit weight calculated by the following procedure:

Percent edible fruit weight of total infested fruit weight infestated by sucking and foliage insect

pests of brinjal were calculated as follows:

% Edible fruit weight =  $\frac{Edible fruit weight}{Total infested fruit weight} \times 100$ 

# Percent non-edible fruit weight calculated by the following procedure:

Percent non-edible fruit weight of total infested fruit weight infestated by sucking and foliage insect pests of brinjal were calculated as follows:

% Non-edible fruit weight =  $\frac{Non-edible \ fruit \ weight}{Total \ infested \ fruit \ weight} \times 100$ 

# Percent reduction of brinjal infestation over control

The number and weight of infested brinjal for each treated plot and untreated control plot were recorded and the percent reduction of brinjal infestation in number and weight was calculated using the following formula:

% Reduction over control =  $\frac{X_2 - X_1}{X_2} \times 100$ 

Where,  $X_1$  = the mean value of the treated plot

 $X_2$  = the mean value of the untreated plot

### 3.13. Statistical analysis

Data were analyzed by using MSTAT-C software for analysis of variance after square root transformation. ANOVA was made by F variance test and the pair comparisons were performed by Duncan Multiple Range Test (DMRT).

# **CHAPTER IV**

# **RESULTS OF DISCUSSION**

The study was conducted to evaluate the effectiveness of botanicals for eco-friendly management of sucking and foliage insect pests of brinjal in the field under the Department of Entomology of Sher-e-Bangla Agricultural University, Dhaka during the period from November 2017 to March 2018. The results have been presented and discussed, and possible interpretations have been given under the following sub-headings:

# 4.1 Infestation of leaf hopper of brinjal

### 4.1.1 Number of leaf hopper per plant

The significant variations were observed among different treatments used for the management practices in terms of number of leaf hopper at different growing stage. At vegetative stage significantly the lowest number of leaf hopper was recorded in  $T_7$  (8.20 leaf hopper/plant), which was followed by  $T_5$  (8.87 leaf hopper/plant),  $T_4$  (9.17 leaf hopper/plant) and  $T_1$  (9.33 leaf hopper/plant). On the other hand, the highest number of leaf hopper was recorded in  $T_8$  (14.27 leaf hopper/plant), which were followed by  $T_3$  (11.30 leaf hopper/plant),  $T_6$  (10.57 leaf hopper/plant) and  $T_2$  (9.70 leaf hopper/plant) respectively. More or less similar trends of number of leaf hopper were also recorded at early fruiting stage, mid fruiting stage and late fruiting stage (Table 1).

In case of mean number of leaf hopper per plant significantly the lowest number of leaf hopper was recorded in  $T_7$  (5.72 leaf hopper/plant), which was followed by  $T_5$  (6.23 leaf hopper/plant),  $T_4$  (6.64 leaf hopper/plant) and  $T_1$  (6.91 leaf hopper/plant) respectively. On the other hand, the highest number of leaf hopper was recorded in  $T_8$  (13.53 leaf hopper/plant), which was significantly different from all other treatments (Table 1). Considering the percent reduction of

incidence of leaf hopper per plant, the highest 57.72% reduction over control was achieved in  $T_7$  followed by  $T_5$  (53.95%) and  $T_4$  (50.92%). On the other hand, the minimum reduction of incidence of leaf hopper over control was found in  $T_3$  (35.48%) followed by  $T_6$  (39.99%). Considering among the nonchemical treatment  $T_1$  showed better performance on the incidence of leaf hopper/plant (6.91) and percent reduction of incidence over control of leaf hopper/plant (48.93) in both (Table 1).

 Table 1: Effect of management practices of number of leaf hopper on fully opened leaves per

 plant

| Treatments            | No. of Leaf hopper per plant |          |          |          |         |                |
|-----------------------|------------------------------|----------|----------|----------|---------|----------------|
|                       | Vegetative                   | Early    | Mid      | Late     | Mean    | Incidence      |
|                       | stage                        | fruiting | fruiting | fruiting |         | reduction over |
|                       |                              | stage    | Stage    | stage    |         | control (%)    |
| $T_1$                 | 9.33 e                       | 8.67 e   | 5.01 e   | 4.57 e   | 6.91 e  | 48.93          |
| T <sub>2</sub>        | 9.70 d                       | 8.83 d   | 5.37 d   | 5.13 d   | 7.26 d  | 46.34          |
| T <sub>3</sub>        | 11.30 b                      | 9.83 b   | 7.17 b   | 6.60 b   | 8.73 b  | 35.48          |
| $T_4$                 | 9.17 f                       | 8.33 f   | 4.87 f   | 4.20 f   | 6.64 f  | 50.92          |
| T <sub>5</sub>        | 8.87 g                       | 7.87 g   | 4.30 g   | 3.87 g   | 6.23 g  | 53.95          |
| T <sub>6</sub>        | 10.57 c                      | 9.33 c   | 6.73 c   | 5.83 c   | 8.12 c  | 39.99          |
| <b>T</b> <sub>7</sub> | 8.20 h                       | 7.13 h   | 3.93 h   | 3.60 h   | 5.72 h  | 57.72          |
| T <sub>8</sub>        | 14.27 a                      | 14.73 a  | 13.23 a  | 11.90 a  | 13.53 a |                |
| CV (%)                | 0.76                         | 0.62     | 1.00     | 1.68     | 0.53    |                |
| LSD (0.05)            | 0.13                         | 0.09     | 0.11     | 0.16     | 0.08    |                |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From the above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) performed best result in reducing number of leaf hopper per plant over control (57.72%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in

terms of reducing the number of leaf hopper over control (48.93%). As a result, the order of rank of efficacy of the treatments applied against brinjal leaf hopper including untreated control in terms of reducing number of brinjal leaf hopper per plant was  $T_7 > T_5 > T_4 > T_1 > T_2 > T_6 > T_3 > T_8$ .

### 4.1.2 Infestation of leaves by brinjal leaf hopper per five plants

The significant variations were observed among different treatments used for the management practices in terms of number of infested leaves by leaf hopper at different growing stage. The highest number of leaves per five plants was recorded in  $T_1$  (59.30 leaves/five plants), which was statistically different from other treatments and followed by  $T_5$  (55.20 leaves/five plants),  $T_8$  (51.70 leaves/five plants) and  $T_4$  (50.30 leaves /five plants). On the other hand, the lowest number of leaves per five plants was recorded in  $T_3$  (40.60 leaves /five plants), which was statistically different from other treatments and followed by  $T_6$  (41.40 leaves /five plants),  $T_2$  (42.47 leaves /five plants) and  $T_7$  (47.10 leaves/five plants) (Table 2).

In case of number of infested leaves, the highest number of infested leaves per five plants was recorded in  $T_8$  (17.00 leaves/five plants), which was statistically different from other treatments and followed by  $T_1$  (14.00 leaves/five plants),  $T_3$  (13.00 leaves/five plants) and  $T_6$  (12.33 leaves /five plants). On the other hand, the lowest number of infested leaves per five plants was recorded in  $T_7$  (6.33 leaves /five plants), which was statistically different from other treatments and followed by  $T_5$  (8.67 leaves /five plants),  $T_4$  (9.33 leaves /five plants) and  $T_2$  (11.33 leaves/five plants) (Table 2).

In case of percent infestation of leaves per five plants, the highest percentage was recorded in  $T_8$  (32.88%) untreated control, which was significantly similar with  $T_3$  (32.02%) and  $T_6$  (29.79%) and followed by  $T_2$  (26.69%). On the other hand, the lowest percentage was recorded in  $T_7$ 

(13.45%), which was significantly similar with  $T_5$  (15.70%) and  $T_4$  (18.56%) and followed by  $T_1$  (23.61%) (Table 2). Considering the percent reduction of number of infested leaves by leaf hopper per plant, the highest 59.09% reduction over control was achieved in  $T_7$  followed by  $T_5$  (52.25%) and  $T_4$  (43.55%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_3$  (2.62%) followed by  $T_6$  (9.40%).

**Table 2:** Effect of management practices on infestation of fully opened leaves by leaf hopper per

 five plants

| Treatments            | Infestation of | Infestation of leaves by leaf hopper per five plants |               |                            |  |  |  |
|-----------------------|----------------|--|---------------|----------------------------|--|--|--|
|                       | No. of total   | No. of infested                                      | % infestation | Infestation reduction over |  |  |  |
|                       | leaves         | leaves   |               | control (%)                |  |  |  |
| $T_1$                 | 59.30 a        | 14.00 b  | 23.61 d       | 28.19                      |  |  |  |
| $T_2$                 | 42.47 f        | 11.33 d  | 26.69 c       | 18.83                      |  |  |  |
| <b>T</b> <sub>3</sub> | 40.60 h        | 13.00 bc   | 32.02 ab      | 2.62                       |  |  |  |
| $T_4$                 | 50.30 d        | 9.33 e   | 18.56 e       | 43.55                      |  |  |  |
| T <sub>5</sub>        | 55.20 b        | 8.67 e   | 15.70 ef      | 52.25                      |  |  |  |
| T <sub>6</sub>        | 41.40 g        | 12.33 cd   | 29.79 b       | 9.40                       |  |  |  |
| $T_7$                 | 47.10 e        | 6.33 f   | 13.45 f       | 59.09                      |  |  |  |
| T <sub>8</sub>        | 51.70 c        | 17.00 a  | 32.88 a       |                            |  |  |  |
| CV (%)                | 0.22           | 7.38   | 7.10          |                            |  |  |  |
| LSD (0.05)            | 0.19           | 1.43   | 2.89          |                            |  |  |  |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments, T<sub>7</sub> (spraying of

Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of infested leaves by

leaf hopper per five plants over control (59.09%). Considering the botanical treatments, T1

(spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in

terms of reducing the number of infested leaves by leaf hopper per five plants over control

(28.19%). As a result, the order of rank of efficacy of the treatments applied against brinjal leaf

hopper including untreated control in terms of reducing number of infested leaves by leaf hopper per five plants was  $T_7 > T_5 > T_4 > T_1 > T_2 > T_6 > T_3 > T_8$ .

### 4.1.3 Infestation plants by leaf hopper

The significant variations were not observed among different treatments used for the management practices in terms of number of total plants at different growing stage. In case of number of infested plants, the highest number of plants per plot was recorded in  $T_8$  (6.33 plants/plot), which was statistically similar with  $T_6$  (5.33 plants/plot),  $T_3$  (5.33 plants/plot) and  $T_4$  (4.33 plants/plot). On the other hand, the lowest number of plants per plot was recorded in  $T_7$  (1.67 plants /plot), which was statistically similar with  $T_1$  (2.67 plants/plot),  $T_2$  (3.67 plants/plot) and  $T_5$  (3.67 plants/plot) (Table 3).

In case of percent infestation of plants per plot, the highest percentage was recorded in  $T_8$  (52.78%) comprised of untreated control, which was significantly similar with  $T_6$  (44.45%),  $T_3$  (44.45%) and  $T_4$  (36.11%). On the other hand, the lowest number of plants per plot was recorded in  $T_7$  (13.89%), which was statistically similar with  $T_1$  (22.22%),  $T_2$  (30.55%) and  $T_5$  (30.55%) (Table 3).Considering the percent reduction of number of leaf hopper per plant, the highest 73.68% reduction over control was achieved in  $T_7$  followed by  $T_1$  (57.90%) and  $T_5$  (42.12%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_3$  (15.78%).

| Treatments     | Infestation of plant | Infestation of plants by leaf hopper per plot |               |                       |  |  |
|----------------|----------------------|---|---------------|-----------------------|--|--|
|                | No. of total         | No. of infested                               | % infestation | Infestation reduction |  |  |
|                | plants               | plants  |               | over control (%)      |  |  |
| T <sub>1</sub> | 12.00 a              | 2.67 de                                       | 22.22 de      | 57.90                 |  |  |
| T <sub>2</sub> | 12.00 a              | 3.67 cd                                       | 30.55 cd      | 42.12                 |  |  |
| T <sub>3</sub> | 12.00 a              | 5.33 ab                                       | 44.45 ab      | 15.78                 |  |  |
| $T_4$          | 12.00 a              | 4.33 bc                                       | 36.11 bc      | 31.58                 |  |  |
| T <sub>5</sub> | 12.00 a              | 3.67 cd                                       | 30.55 cd      | 42.12                 |  |  |
| T <sub>6</sub> | 12.00 a              | 5.33 ab                                       | 44.45 ab      | 15.78                 |  |  |
| T <sub>7</sub> | 12.00 a              | 1.67 e  | 13.89 e       | 73.68                 |  |  |
| T <sub>8</sub> | 12.00 a              | 6.33 a  | 52.78 a       |                       |  |  |
| CV (%)         | 00                   | 14.61   | 14.61         |                       |  |  |
| LSD (0.05)     | 0.05                 | 1.02  | 8.48          |                       |  |  |

Table 3: Effect of management practices of infestation of plants by leaf hopper per plot

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments, T<sub>7</sub> (spraying of

Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of infested plants by

leaf hopper per plot over control (73.68%). Considering the botanical treatments, T<sub>1</sub> (spraying of

Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of

reducing the number of infested plants by leaf hopper per plot over control (57.90%). As a result,

the order of rank of efficacy of the treatments applied against brinjal leaf hopper including

untreated control in terms of reducing number of infested plant by leaf hopper per plot was T<sub>7</sub>>

 $T_1 > T_5 > T_2 > T_4 > T_6 > T_3 > T_8.$ 

### 4.2 Infestation of aphid of brinjal

# 4.2.1 Number of aphid per plant

The significant variations were observed among different treatments used for the management practices in terms of number of aphid at different growing stage. At vegetative stage, the lowest number of aphid was recorded in  $T_7$  (3.70 aphid/plant), which was followed by  $T_5$  (4.20 aphid/plant),  $T_4$  (4.40 aphid/plant) and  $T_1$  (4.83 aphid/plant). On the other hand, the highest number of aphid was recorded in  $T_8$  (8.00 aphid/plant), which was followed by  $T_3$  (6.20 aphid/plant),  $T_6$  (5.73 aphid/plant) and  $T_2$  (5.23 aphid/plant). More or less similar trends of number of leaf hopper were also recorded at early fruiting, mid fruiting stage and late fruiting stage (Table 4).

In case of mean number of leaf hopper per plant, the highest number of aphid was recorded in  $T_8$  (7.65 aphid/plant) comprised of untreated control, which was significantly different from all other treatments and followed by  $T_3$  (5.54 aphid/plant),  $T_6$  (5.06 aphid/plant) and  $T_2$  (4.38 aphid/plant). On the other hand, the lowest mean leaf infestation by number was recorded in  $T_7$  (2.79 aphid/plant), which was significantly different from all other treatments and followed by  $T_5$  (3.27 aphid/plant),  $T_4$  (3.61 aphid/plant) and  $T_1$  (3.89 aphid/plant) (Table 4). Considering the percent reduction of number of aphid per plant, the highest 63.53% reduction over control was achieved in  $T_7$  followed by  $T_5$  (57.25%) and  $T_4$  (52.81%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_3$  (27.58%) followed by  $T_6$  (33.86%).

Table 4: Effect of management practices of number of aphid on fully opened leaves per plants

| Treatments            | No. of aphids per plant |          |          |          |        |                |
|-----------------------|-------------------------|----------|----------|----------|--------|----------------|
|                       | Vegetative              | Early    | Mid      | Late     | Mean   | Incidence      |
|                       | stage                   | fruiting | fruiting | fruiting |        | reduction over |
|                       |                         | stage    | Stage    | stage    |        | control (%)    |
| <b>T</b> <sub>1</sub> | 4.83 e                  | 4.17 e   | 3.73 e   | 2.83 e   | 3.89 e | 49.15          |

| T <sub>2</sub> | 5.23 d | 4.87 d | 4.13 d | 3.27 d | 4.38 d | 42.75 |
|----------------|--------|--------|--------|--------|--------|-------|
| T <sub>3</sub> | 6.20 b | 6.13 b | 5.17 b | 4.63 b | 5.54 b | 27.58 |
| T <sub>4</sub> | 4.70 f | 3.83 f | 3.27 f | 2.63 f | 3.61 f | 52.81 |
| T <sub>5</sub> | 4.20 g | 3.50 g | 3.03 g | 2.33 g | 3.27 g | 57.25 |
| T <sub>6</sub> | 5.73 c | 5.63 c | 4.73 c | 4.13 c | 5.06 c | 33.86 |
| T <sub>7</sub> | 3.70 h | 3.20 h | 2.53 h | 1.73 h | 2.79 h | 63.53 |
| T <sub>8</sub> | 8.00 a | 9.17 a | 7.63 a | 5.80 a | 7.65 a |       |
| CV (%)         | 1.48   | 1.27   | 1.04   | 1.76   | 0.8    |       |
| LSD (0.05)     | 0.13   | 0.11   | 0.08   | 0.11   | 0.05   |       |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of aphid per five plants over control (63.53%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the number of aphid per five plants over control (49.15%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of aphid per five plants was  $T_7 > T_5 > T_4 > T_1 > T_2 > T_6 > T_3 > T_8$ .

### **4.2.2 Infestation of leaves by aphid per plant**

The significant variations were observed among different treatments used for the management practices in terms of number of infested leaves by aphid at different growing stage. The highest number of leaves per five plants was recorded in  $T_1$  (59.30 leaves/five plants), which was statistically different from other treatments and followed by  $T_5$  (55.20 leaves/five plants),  $T_8$  (51.70 leaves/five plants) and  $T_4$  (50.30 leaves /five plants). On the other hand, the lowest number of leaves per five plants was recorded in  $T_3$  (40.60 leaves /five plants), which was

statistically different from other treatments and followed by  $T_6$  (41.40 leaves /five plants),  $T_2$  (42.47 leaves /five plants) and  $T_7$  (47.10 leaves/five plants) (Table 5).

In case of number of infested leaves, the highest number of infested leaves per five plants was recorded in  $T_8$  (16.67 leaves/five plants), which was statistically different from other treatments and followed by  $T_1$  (10.67 leaves/five plants),  $T_3$  (10.33 leaves/five plants) and  $T_6$  (9.33 leaves /five plants). On the other hand, the lowest number of infested leaves per five plants was recorded in  $T_7$  (5.67 leaves /five plants), which was statistically different from other treatments and followed by  $T_5$  (8.00 leaves /five plants),  $T_4$  (8.33 leaves /five plants) and  $T_2$  (8.67 leaves/five plants) (Table 5).

In case of percent infestation of leaves per five plants, the highest percentage was recorded in  $T_8$  (32.24%) comprised of untreated control, which was statistically different from other treatments and followed by  $T_3$  (25.45%),  $T_6$  (22.54%) and  $T_2$  (20.41%). On the other hand, the lowest percentage was recorded in  $T_7$  (12.03%), which was statistically different with other treatments and followed by  $T_5$  (14.49%),  $T_4$  (16.57%) and  $T_1$  (17.99%) (Table 5). Considering the percent reduction of number of aphid per plant, the highest 62.67% reduction over control was achieved in  $T_7$  followed by  $T_5$  (55.05%) and  $T_4$  (48.60%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_3$  (21.05%) followed by  $T_6$  (30.06%).

**Table 5:** Effect of management practices on infestation of fully opened leaves by aphid per five

 plants

| Treatments     | Infestation of leave | Infestation of leaves by aphid per five plants         |         |                |  |  |  |
|----------------|----------------------|--|---------|----------------|--|--|--|
|                | No. of total         | No. of total No. of infested % infestation Infestation |         |                |  |  |  |
|                | leaves               | leaves leaves  |         | reduction over |  |  |  |
|                |                      |  |         | control (%)    |  |  |  |
| T <sub>1</sub> | 59.30 a              | 10.67 b  | 17.99 e | 44.20          |  |  |  |
| T <sub>2</sub> | 42.47 f              | 8.67 cd  | 20.41 d | 36.68          |  |  |  |

| T <sub>3</sub> | 40.60 h | 10.33 b | 25.45 b | 21.05 |
|----------------|---------|---------|---------|-------|
| T <sub>4</sub> | 50.30 d | 8.33 d  | 16.57 f | 48.60 |
| T <sub>5</sub> | 55.20 b | 8.00 d  | 14.49 g | 55.05 |
| T <sub>6</sub> | 41.40 g | 9.33 c  | 22.54 c | 30.06 |
| T <sub>7</sub> | 47.10 e | 5.67 e  | 12.03 h | 62.67 |
| T <sub>8</sub> | 51.70 c | 16.67 a | 32.24 a |       |
| CV (%)         | 0.22    | 4.21    | 4.15    |       |
| LSD (0.05)     | 0.19    | 0.69    | 1.42    |       |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of infested leaves by aphid per five plants over control (62.67%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the number of infested leaves by aphid per five plants over control (42.20%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of infested leaves by aphid per five plants was  $T_7 > T_5 > T_4 >$ 

 $T_1 > T_2 > T_6 > T_3 > T_8$ .

## 4.2.3. Infestation plants by aphid

The significant variations were not observed among different treatments used for the management practices in terms of number of total plants at different growing stage. In case of number of infested plants, the highest number of plants per plot was recorded in  $T_8$  (6.33 plants/plot), which was statistically similar with  $T_6$  (5.67 plants/plot),  $T_3$  (5.33 plants/plot),  $T_4$  (4.33 plants/plot) and  $T_2$  (4.33 plants/plot). On the other hand, the lowest number of infested

plants per plot was recorded in  $T_7$  (1.67 plants /plot), which was statistically similar with  $T_1$  (3.33 plants/plot) and  $T_5$  (3.33 plants/plot) (Table 6).

In case of percent infestation of plants per plot, the highest percentage was recorded in  $T_8$  (52.78%) untreated control, which was significantly similar with  $T_6$  (47.22%),  $T_3$  (44.45%)  $T_4$  (36.11%) and  $T_2$  (36.11%). On the other hand, the lowest number of infested plants per plot was recorded in  $T_7$  (13.89%), which was statistically different from other treatments and followed by  $T_1$  (27.78%) and  $T_5$  (27.78%) (Table 6). Considering the percent reduction of number of aphid per plant, the highest 73.68% reduction over control was achieved in  $T_7$  followed by  $T_1$  (47.37%) and  $T_5$  (47.37%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_6$  (10.53%) followed by  $T_6$  (15.78%).

| Treatments     | Infestation of plant | Infestation of plants by aphid per plot |               |                |  |  |
|----------------|----------------------|---|---------------|----------------|--|--|
|                | No. of total         | No. of infested                         | % infestation | Infestation    |  |  |
|                | plants               | plants                                  |               | reduction over |  |  |
|                |                      |   |               | control (%)    |  |  |
| T <sub>1</sub> | 12.00 a              | 3.33 d                                  | 27.78 d       | 47.37          |  |  |
| T <sub>2</sub> | 12.00 a              | 4.33 c                                  | 36.11 c       | 31.58          |  |  |
| T <sub>3</sub> | 12.00 a              | 5.33 b                                  | 44.45 b       | 15.78          |  |  |
| $T_4$          | 12.00 a              | 4.33 c                                  | 36.11 c       | 31.58          |  |  |
| T <sub>5</sub> | 12.00 a              | 3.33 cd                                 | 27.78 cd      | 47.37          |  |  |
| T <sub>6</sub> | 12.00 a              | 5.67 ab                                 | 47.22 ab      | 10.53          |  |  |
| T <sub>7</sub> | 12.00 a              | 1.67 e                                  | 13.89 e       | 73.68          |  |  |
| T <sub>8</sub> | 12.00 a              | 6.33 a                                  | 52.78 a       |                |  |  |
| CV (%)         | 0.0                  | 12.84                                   | 12.84         |                |  |  |
| LSD (0.05)     | 0.05                 | 0.93                                    | 7.76          |                |  |  |

Table 6: Effect of management practices of infestation of plants by aphid per plot

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of infested plants by aphid per plot over control (73.68%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the number of infested plants by aphid per plot over control (47.37%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of infested plant by aphid per plot was  $T_7 > T_1 > T_5 > T_2 > T_4 > T_3 > T_6 > T_8$ .

### 4.3 Infestation of epilachna beetle of brinjal

### 4.3.1 Number of epilachna beetle per plant

The significant variations were observed among different treatments used for the management practices in terms of number of epilachna beetle at different growing stage. At vegetative stage, the lowest number of epilachna beetle was recorded in  $T_7$  (1.53 epilachna beetle/plant), which was followed by  $T_5$  (2.17 epilachna beetle/plant),  $T_4$  (2.27 epilachna beetle/plant) and  $T_1$  (2.47 epilachna beetle/plant). On the other hand, the highest number of epilachna beetle was recorded in  $T_8$  (4.10 epilachna beetle/plant), which was followed by  $T_3$  (3.23 epilachna beetle/plant),  $T_6$  (3.17 epilachna beetle/plant) and  $T_2$  (2.63 epilachna beetle/plant). More or less similar trends of number of epilachna beetle were also recorded at early fruiting, mid fruiting stage and late fruiting stage (Table 7).

In case of mean number of epilachna beetle per plant, the highest number of epilachna beetle was recorded in  $T_8$  (4.53 epilachna beetle/plant) comprised of untreated control, which was significantly different from all other treatments and followed by  $T_3$  (2.94 epilachna beetle/plant),  $T_6$  (2.68 epilachna beetle/plant) and  $T_2$  (2.30 epilachna beetle/plant). On the other hand, the

lowest mean leaf infestation by number was recorded in  $T_7$  (1.14 epilachna beetle/plant), which was significantly different from all other treatments and followed by  $T_5$  (1.55 epilachna beetle/plant),  $T_4$  (1.77 epilachna beetle/plant) and  $T_1$  (2.03 epilachna beetle/plant) (Table 7). Considering the percent reduction of number of epilachna beetle per plant, the highest 74.83% reduction over control was achieved in  $T_7$  followed by  $T_5$  (65.78%) and  $T_4$  (60.93%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_3$  (35.10%) followed by  $T_6$  (40.84%).

**Table 7:** Effect of management practices of number of epilachna beetle on fully opened leaves

 per plant

| Treatments     | No. of epilachna beetle per plant |          |          |          |        |             |
|----------------|-----------------------------------|----------|----------|----------|--------|-------------|
|                | Vegetative                        | Early    | Mid      | Late     | Mean   | Incidence   |
|                | stage                             | fruiting | fruiting | fruiting |        | reduction   |
|                |                                   | stage    | Stage    | stage    |        | over        |
|                |                                   |          |          |          |        | control (%) |
| $T_1$          | 2.47 d                            | 2.13 e   | 1.83 e   | 1.67 e   | 2.03 e | 55.19       |
| T <sub>2</sub> | 2.63 c                            | 2.33 d   | 2.17 d   | 2.03 d   | 2.30 d | 49.23       |
| T <sub>3</sub> | 3.23 b                            | 3.13 b   | 2.73 b   | 2.63 b   | 2.94 b | 35.10       |
| $T_4$          | 2.27 e                            | 1.83 f   | 1.63 f   | 1.33 f   | 1.77 f | 60.93       |
| T <sub>5</sub> | 2.17 f                            | 1.73 f   | 1.23 g   | 1.07 g   | 1.55 g | 65.78       |
| T <sub>6</sub> | 3.17 b                            | 2.70 c   | 2.53 c   | 2.33 c   | 2.68 c | 40.84       |
| T <sub>7</sub> | 1.53 g                            | 1.30 g   | 1.07 h   | 0.67 h   | 1.14 h | 74.83       |
| T <sub>8</sub> | 4.10 a                            | 4.57 a   | 4.80 a   | 4.63 a   | 4.53 a |             |
| CV (%)         | 2.06                              | 2.80     | 2.20     | 2.26     | 1.14   |             |
| LSD (0.05)     | 0.09                              | 0.12     | 0.08     | 0.08     | 0.05   |             |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments, T<sub>7</sub> (spraying of

Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of epilachna beetle

per plants over control (74.83%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the number of epilachna beetle per plants over control (55.19%). As a result, the order of rank of efficacy of the treatments applied against epilachna beetle including untreated control in terms of reducing number of epilachna beetle per plants was  $T_7 > T_5 > T_4 > T_1 > T_2 > T_6 > T_3 > T_8$ .

### 4.2.2 Infestation of leaves by epilachna beetle per plant

The significant variations were observed among different treatments used for the management practices in terms of number of leaves by epilachna beetle at different growing stage. The highest number of leaves per five plants was recorded in  $T_1$  (59.30 leaves/five plants), which was statistically different from other treatments and followed by  $T_5$  (55.20 leaves/five plants),  $T_8$  (51.70 leaves/five plants) and  $T_4$  (50.30 leaves /five plants). On the other hand, the lowest number of leaves per five plants was recorded in  $T_3$  (40.60 leaves /five plants), which was statistically different from other treatments and followed by  $T_6$  (41.40 leaves /five plants),  $T_2$  (42.47 leaves /five plants) and  $T_7$  (47.10 leaves/five plants) (Table 8).

In case of number of infested leaves, the highest number of infested leaves per five plants was recorded in  $T_8$  (12.33 leaves/five plants), which was statistically different from other treatments and followed by  $T_1$  (9.00 leaves/five plants),  $T_3$  (9.00 leaves/five plants) and  $T_6$  (8.33 leaves /five plants). On the other hand, the lowest number of infested leaves per five plants was recorded in  $T_7$  (4.33 leaves /five plants), which was statistically different from other treatments and followed

by  $T_5$  (5.67 leaves /five plants),  $T_4$  (7.00 leaves /five plants) and  $T_2$  (7.00 leaves/five plants) (Table 8).

In case of percent infestation of leaves per five plants, the highest percentage was recorded in  $T_8$  (23.86%) comprised of untreated control, which was statistically similar with  $T_3$  (22.16%) and followed by  $T_6$  (20.13%). On the other hand, the lowest percentage was recorded in  $T_7$  (9.20%), which was statistically similar with  $T_5$  (10.27%) and followed by  $T_4$  (13.91%),  $T_1$  (15.18%) and  $T_2$  (16.48%) (Table 8). Considering the percent reduction of number of epilachna beetle per plant, the highest 61.44% reduction over control was achieved in  $T_7$  followed by  $T_5$  (56.96%) and  $T_4$  (41.70%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_3$  (7.12%) followed by  $T_6$  (15.63%).

**Table 8:** Effect of management practices on infestation of fully opened leaves by epilachna

 beetle per five plants

| Treatments            | Infestation of leaves by epilachna beetle per five plants |                 |               |                |  |
|-----------------------|---|-----------------|---------------|----------------|--|
|                       | No. of total  | No. of infested | % infestation | Infestation    |  |
|                       | leaves  | leaves          |               | reduction over |  |
|                       |   |                 |               | control (%)    |  |
| T <sub>1</sub>        | 59.30 a   | 9.00 b          | 15.18 c       | 36.38          |  |
| T <sub>2</sub>        | 42.47 f   | 7.00 c          | 16.48 c       | 30.93          |  |
| T <sub>3</sub>        | 40.60 h   | 9.00 b          | 22.16 ab      | 7.12           |  |
| $T_4$                 | 50.30 d   | 7.00 c          | 13.91 c       | 41.70          |  |
| T <sub>5</sub>        | 55.20 b   | 5.67 d          | 10.27 d       | 56.96          |  |
| T <sub>6</sub>        | 41.40 g   | 8.33 b          | 20.13 b       | 15.63          |  |
| <b>T</b> <sub>7</sub> | 47.10 e   | 4.33 e          | 9.20 d        | 61.44          |  |
| T <sub>8</sub>        | 51.70 c   | 12.33 a         | 23.86 a       |                |  |
| CV (%)                | 0.22  | 8.75            | 9.25          |                |  |
| LSD (0.05)            | 0.19  | 1.16            | 2.56          |                |  |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days at$ 

days interval;  $T_6$ : Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval;  $T_7$ : Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval;  $T_8$  Untreated control.]

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of infested leaves by epilachna beetle per five plants over control (61.44%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the number of infested leaves by epilachna beetle per five plants over control (36.38%). As a result, the order of rank of efficacy of the treatments applied against epilachna beetle including untreated control in terms of reducing number of infested leaves by epilachna beetle per five plants was  $T_7 > T_5 > T_4 > T_1 > T_2 > T_6 > T_3 > T_8$ .

### **4.3.3 Infestation plants by epilachna beetle**

The significant variations were not observed among different treatments used for the management practices in terms of number of total plants at different growing stage. In case of number of infested plants, the highest number of plants per plot was recorded in  $T_8$  (4.33 plants/plot), which was statistically different from other treatments and followed by  $T_3$  (3.67 plants/plot),  $T_6$  (2.67 plants/plot),  $T_4$  (2.33 plants/plot),  $T_6$  (2.67 plants/plot) and  $T_2$  (4.33 plants/plot). On the other hand, the lowest number of infested plants per plot was recorded in  $T_7$  (1.00 plants/plot), which was statistically similar with  $T_1$  (1.33 plants/plot) and  $T_5$  (1.00 plants/plot) (Table 9).

In case of percent infestation of plants per plot, the highest percentage was recorded in  $T_8$  (36.11%) comprised of untreated control, which was significantly similar with  $T_3$  (30.55%),  $T_6$  (22.22%),  $T_2$  (22.22%) and  $T_4$  (19.45%). On the other hand, the lowest percentage was recorded in  $T_7$  (8.33%), which was statistically different from other treatments and followed by  $T_5$ 

(8.33%) and T<sub>1</sub> (11.11%) (Table 9). Considering the percent reduction of infestation of plant by epilachna beetle per plot, the highest 76.93% reduction over control was achieved in T<sub>7</sub> followed by T<sub>5</sub> (76.93%) and T<sub>5</sub> (69.23%). On the other hand, the minimum reduction of plant infestation over control was found in T<sub>3</sub> (15.40%) followed by T<sub>6</sub> (38.47%) and T<sub>2</sub> (38.47%).

| Treatments            | Infestation of plants by epilachne beetle per plot |                 |               |                       |  |  |
|-----------------------|--|-----------------|---------------|-----------------------|--|--|
|                       | No. of total                                       | No. of infested | % infestation | Infestation reduction |  |  |
|                       | plants   | plants          |               | over control (%)      |  |  |
| <b>T</b> <sub>1</sub> | 12.00 a  | 1.33 d          | 11.11 d       | 69.23                 |  |  |
| T <sub>2</sub>        | 12.00 a  | 2.67 c          | 22.22 c       | 38.47                 |  |  |
| T <sub>3</sub>        | 12.00 a  | 3.67 b          | 30.55 b       | 15.40                 |  |  |
| $T_4$                 | 12.00 a  | 2.33 c          | 19.45 c       | 46.14                 |  |  |
| T <sub>5</sub>        | 12.00 a  | 1.00 d          | 8.33 d        | 76.93                 |  |  |
| T <sub>6</sub>        | 12.00 a  | 2.67 c          | 22.22 c       | 38.47                 |  |  |
| <b>T</b> <sub>7</sub> | 12.00 a  | 1.00 d          | 8.33 d        | 76.93                 |  |  |
| T <sub>8</sub>        | 12.00 a  | 4.33 a          | 36.11 a       |                       |  |  |
| CV (%)                | 00   | 14.89           | 14.89         |                       |  |  |
| LSD (0.05)            | 0.05   | 0.60            | 4.98          |                       |  |  |

**Table 9:** Effect of management practices on infestation of plants by epilachne beetle per plot

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the percentage of infested plant by epilachna beetle per plot over control (76.93%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the percentage of infested plants by epilachna beetle per plot over control (69.23%). As a result, the order of rank of efficacy of the treatments applied against epilachna beetle including untreated control in terms of reducing percentage of infested plants by epilachna beetle per plot was  $T_7 > T_5 > T_1 > T_4 > T_2 > T_6 > T_3 > T_8$ .

### 4.4 Infestation of leaf roller of brinjal

### 4.4.1 Number of leaf roller per plant

The significant variations were observed among different treatments used for the management practices in terms of number of leaf roller at different growing stage. At vegetative stage, the

lowest number of leaf roller was recorded in  $T_7$  (3.13 leaf roller/plant), which was followed by  $T_5$  (3.53 leaf roller/plant),  $T_4$  (3.83 leaf roller/plant) and  $T_1$  (4.13 leaf roller/plant). On the other hand, the highest number of leaf roller was recorded in  $T_8$  (6.13 leaf roller/plant), which was followed by  $T_3$  (5.20 leaf roller/plant),  $T_6$  (4.77 leaf roller/plant) and  $T_2$  (4.43 leaf roller/plant). More or less similar trends of number of leaf roller were also recorded at early fruiting, mid fruiting stage and late fruiting stage (Table 10).

In case of mean number of leaf roller per plant, the highest number of leaf roller was recorded in  $T_8$  (5.25 leaf roller/plant) comprised of untreated control, which was significantly different from all other treatments and followed by  $T_3$  (4.20 leaf roller/plant),  $T_6$  (3.78 leaf roller /plant) and  $T_2$  (3.63 leaf roller/plant). On the other hand, the lowest mean number of leaf roller was recorded in  $T_7$  (2.12 leaf roller/plant), which was significantly different from all other treatments and followed by  $T_5$  (2.45 leaf roller/plant),  $T_4$  (2.80 leaf roller/plant) and  $T_1$  (3.03 leaf roller/plant) (Table 10). Considering the percent reduction of number of leaf roller per plant, the highest 59.62% reduction over control was achieved in  $T_7$  followed by  $T_5$  (53.33%) and  $T_4$  (46.67%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_3$  (20.00%) followed by  $T_6$  (28.00%).

Table 10: Effect of management practices against number of leaf roller on fully opened leaves

per plant

| Treatments            | No. of Leaf roller per plant |          |          |          |        |                |  |
|-----------------------|------------------------------|----------|----------|----------|--------|----------------|--|
|                       | Vegetative                   | Early    | Mid      | Late     | Mean   | Incidence      |  |
|                       | stage                        | fruiting | fruiting | fruiting |        | reduction over |  |
|                       |                              | stage    | Stage    | stage    |        | control (%)    |  |
| $T_1$                 | 4.13 e                       | 3.37 e   | 2.83 e   | 1.77 e   | 3.03 e | 42.29          |  |
| T <sub>2</sub>        | 4.43 d                       | 3.70 d   | 3.13 d   | 2.17 d   | 3.63 d | 30.86          |  |
| T <sub>3</sub>        | 5.20 b                       | 4.53 b   | 3.87 b   | 3.20 b   | 4.20 b | 20.00          |  |
| $T_4$                 | 3.83 f                       | 3.17 f   | 2.63 f   | 1.53 f   | 2.80 f | 46.67          |  |
| T <sub>5</sub>        | 3.53 g                       | 2.83 g   | 2.17 g   | 1.23 g   | 2.45 f | 53.33          |  |
| T <sub>6</sub>        | 4.77 c                       | 4.17 c   | 3.53 c   | 2.63 c   | 3.78 c | 28.00          |  |
| <b>T</b> <sub>7</sub> | 3.13 h                       | 2.53 h   | 1.73 h   | 1.07 h   | 2.12 h | 59.62          |  |
| T <sub>8</sub>        | 6.13 a                       | 5.83 a   | 4.80 a   | 4.20     | 5.25 a |                |  |
| CV (%)                | 1.27                         | 1.79     | 2.06     | 2.88     | 0.89   |                |  |
| LSD (0.05)            | 0.09                         | 0.12     | 0.11     | 0.11     | 0.05   |                |  |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of leaf roller per plant over control (59.62%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the number of leaf roller per plant over control (42.29%). As a result, the order of rank of efficacy of the treatments applied against leaf roller including untreated control in terms of reducing number of leaf roller per plant was  $T_7 > T_5 > T_4 > T_1 > T_2 > T_6 > T_3 > T_8$ .

# 4.4.2 Infestation of leaves by leaf roller per plant

The significant variations were observed among different treatments used for the management practices in terms of number of infested leaves by leaf roller at different growing stage. The highest number of leaves per five plants was recorded in  $T_1$  (59.30 leaves/five plants), which was statistically different from other treatments and followed by  $T_5$  (55.20 leaves/five plants),  $T_8$ (51.70 leaves/five plants) and  $T_4$  (50.30 leaves /five plants). On the other hand, the lowest number of leaves per five plants was recorded in  $T_3$  (40.60 leaves /five plants), which was statistically different from other treatments and followed by  $T_6$  (41.40 leaves /five plants),  $T_2$ (42.47 leaves /five plants) and  $T_7$  (47.10 leaves/five plants) (Table 11).

In case of number of infested leaves, the highest number of infested leaves per five plants was recorded in  $T_8$  (10.67 leaves/five plants), which was statistically different from other treatments and followed by  $T_6$  (6.67 leaves/five plants),  $T_3$  (6.33 leaves/five plants),  $T_1$  (6.00 leaves/five plants) and  $T_4$  (5.67 leaves /five plants). On the other hand, the lowest number of infested leaves per five plants was recorded in  $T_7$  (2.67 leaves /five plants), which was statistically different from other treatments and followed by  $T_5$  (4.00 leaves /five plants) and  $T_2$  (5.33 leaves/five plants) (Table 11).

In case of percent infestation of leaves per five plants, the highest percentage was recorded in  $T_8$  (20.64%) comprised of untreated control, which was statistically different from other treatments and followed by  $T_6$  (16.10%),  $T_3$  (15.60%),  $T_2$  (12.55%),  $T_4$  (11.26%) and  $T_1$  (10.12%). On the other hand, the lowest percentage was recorded in  $T_7$  (5.66%), which was statistically similar with  $T_5$  (7.25%) (Table 11). Considering the percent reduction of number of leaf roller per plant, the highest 72.58% reduction over control was achieved in  $T_7$  followed by  $T_5$  (64.87%) and  $T_1$  (50.97%). On the other hand, the minimum reduction of leaf infestation over control was found in  $T_6$  (22.00%) followed by  $T_3$  (24.42%).

Table 11: Effect of management practices of infestation of fully opened leaves by leaf roller per

| five plants |  |
|-------------|--|
|             |  |

| Treatments     | Infestation of leaves by leaf roller per five plants |                 |               |                       |  |  |
|----------------|--|-----------------|---------------|-----------------------|--|--|
|                | No. of total   | No. of infested | % infestation | Infestation reduction |  |  |
|                | leaves   | leaves          |               | over control (%)      |  |  |
| T <sub>1</sub> | 59.30 a  | 6.00 bc         | 10.12 c       | 50.97                 |  |  |
| T <sub>2</sub> | 42.47 f  | 5.33 c          | 12.55 c       | 39.20                 |  |  |
| T <sub>3</sub> | 40.60 h  | 6.33 bc         | 15.60 b       | 24.42                 |  |  |
| T <sub>4</sub> | 50.30 d  | 5.67 bc         | 11.26 c       | 45.45                 |  |  |
| T <sub>5</sub> | 55.20 b  | 4.00 d          | 7.25 d        | 64.87                 |  |  |
| T <sub>6</sub> | 41.40 g  | 6.67 b          | 16.10 b       | 22.00                 |  |  |
| T <sub>7</sub> | 47.10 e  | 2.67 e          | 5.66 d        | 72.58                 |  |  |
| T <sub>8</sub> | 51.70 c  | 10.67 a         | 20.64 a       |                       |  |  |
| CV (%)         | 0.22   | 11.66           | 11.30         |                       |  |  |
| LSD (0.05)     | 0.19   | 1.165           | 2.37          |                       |  |  |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments, T<sub>7</sub> (spraying of

Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the number of infested leaves by leaf roller per five plants over control (72.58%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the number of infested leaves by leaf roller per five plants over control (50.97%). As a result, the order of rank of efficacy of the treatments applied against leaf roller including untreated control in terms of reducing number of infested leaves by leaf roller per five plants was  $T_7 > T_5 > T_1 > T_4 > T_2 > T_3 > T_6 > T_8$ .

### **4.4.3 Infestation plants by leaf roller**

The significant variations were not observed among different treatments used for the management practices in terms of number of total plants at different growing stage. In case of number of infested plants, the highest number of plants per plot was recorded in  $T_8$  (3.33 plants/plot), which was statistically similar with  $T_3$  (3.33 plants/plot) and followed by  $T_6$  (2.33 plants/plot),  $T_4$  (2.00 plants/plot) and  $T_2$  (2.00 plants/plot). On the other hand, the lowest number of infested plants per plot was recorded in  $T_7$  (1.00 plants/plot) which was statistically similar with  $T_1$  (1.00 plants/plot) and  $T_5$  (1.00 plants/plot) (Table 12).

In case of percent infestation of plants per plot, the highest percentage was recorded in  $T_8$  (27.78%) comprised of untreated control, which was significantly similar with  $T_3$  (27.78%) and followed by  $T_6$  (19.45%),  $T_4$  (16.67%) and  $T_2$  (16.67%). On the other hand, the lowest number of infested plants per plot was recorded in  $T_7$  (8.33%), which was statistically similar with  $T_1$  (8.33%) and  $T_5$  (8.33%) (Table 12). Considering the percent reduction of number of infested plant per plot, the highest 70.01% reduction over control was achieved in  $T_7$  and similar with  $T_1$  (70.01%) and  $T_5$  (70.01%). On the other hand, the minimum reduction of infestation of plant per plot over control was found in  $T_3$  (0.00%) followed by  $T_6$  (29.99%).

| Treatments            | Infestation of plants by leaf roller per plot |                 |               |                       |  |  |  |
|-----------------------|---|-----------------|---------------|-----------------------|--|--|--|
|                       | No. of total                                  | No. of infested | % infestation | Infestation reduction |  |  |  |
|                       | plants  | plants          |               | over control (%)      |  |  |  |
| T <sub>1</sub>        | 12.00 a                                       | 1.00 c          | 8.33 c        | 70.01                 |  |  |  |
| T <sub>2</sub>        | 12.00 a                                       | 2.00 b          | 16.67 b       | 39.99                 |  |  |  |
| T <sub>3</sub>        | 12.00 a                                       | 3.33 a          | 27.78 a       | 0.00                  |  |  |  |
| $T_4$                 | 12.00 a                                       | 2.00 b          | 16.67 b       | 39.99                 |  |  |  |
| T <sub>5</sub>        | 12.00 a                                       | 1.00 c          | 8.33 c        | 70.01                 |  |  |  |
| T <sub>6</sub>        | 12.00 a                                       | 2.33 b          | 19.45 b       | 29.99                 |  |  |  |
| <b>T</b> <sub>7</sub> | 12.00 a                                       | 1.00 c          | 8.33 c        | 70.01                 |  |  |  |
| T <sub>8</sub>        | 12.00 a                                       | 3.33 a          | 27.78 a       |                       |  |  |  |
| CV (%)                | 00  | 17.68           | 17.67         |                       |  |  |  |
| LSD (0.05)            | 0.05  | 0.60            | 4.98          |                       |  |  |  |

Table 12: Effect of management practices against infestation of plants by leaf roller per plot

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among the different treatments, T<sub>7</sub> (spraying of

Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced the percent of infested plant by leaf roller per plot over control (70.01%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the percent of infested plants by leaf roller per plot over control (70.01%). As a result, the order of rank of efficacy of the treatments applied against leaf roller including untreated control in terms of reducing percent of infested plants by leaf hopper per plot was  $T_7 > T_1 > T_5 >$ 

 $T_2 > T_4 > T_6 > T_3 > T_8$ .

### 4.5. Incidence of beneficial arthropods

### 4.5.1. Lady bird beetle

The significant variations were observed among the different treatments used for the management practices in terms of number of lady bird beetle. At vegetative stage, the highest number of lady bird beetle per plants was recorded in  $T_1$  (3.17 lady bird beetle/plant), which was statistically similar with T<sub>2</sub> (3.10 lady bird beetle/plant) and followed by T<sub>8</sub> (2.73 lady bird beetle/plant) and  $T_3$  (2.37 lady bird beetle/plant). On the other hand, the lowest number of lady bird beetle per plants was recorded in  $T_5$  (1.23 lady bird beetle/plant) which was statistically different with other treatments and followed by T<sub>7</sub> (1.60 lady bird beetle/plant), T<sub>4</sub> (1.73 lady bird beetle/plant) and T<sub>6</sub> (2.17 lady bird beetle/plant) (Table 13). In case of early fruiting stage, the highest number of lady bird beetle per plants was recorded in  $T_1$  (4.77 lady bird beetle/plant), which was statistically different from other treatments and followed by T<sub>2</sub> (4.37 lady bird beetle/plant), T<sub>8</sub> (4.17 lady bird beetle/plant) and T<sub>3</sub> (3.83 lady bird beetle/plant). On the other hand, the lowest number of lady bird beetle per plants was recorded in T<sub>5</sub> (2.30 lady bird beetle/plant) which was statistically different with other treatments and followed by  $T_7$  (2.73 lady bird beetle/plant), T<sub>4</sub> (3.13 lady bird beetle/plant) and T<sub>6</sub> (3.57 lady bird beetle/plant) (Table 13). More or less similar trends of number of lady bird beetle per plants were also recorded at mid fruiting stage and late fruiting stage.

In case of mean number of lady bird beetle, the highest number of lady bird beetle per plants was recorded in  $T_1$  (5.21 lady bird beetle/plant) comprised of untreated control, which was significantly different with other treatments and followed by  $T_2$  (4.87 lady bird beetle/plant),  $T_8$  (4.60 lady bird beetle/plant) and  $T_3$  (4.16 lady bird beetle/plant). On the other hand, the lowest mean number of lady bird beetle per plants was recorded in  $T_5$  (2.55 lady bird beetle/plant),

which was significantly different from other treatments and followed by  $T_7$  (3.00 lady bird beetle/plant),  $T_4$  (3.32 lady bird beetle/plant) and  $T_6$  (3.78 lady bird beetle/plant) (Table 13). Considering the percent increase or decrease of number of lady bird beetle per plants, the highest 13.26% increase over control was observed in  $T_1$  followed by  $T_2$  (5.87%). On the other hand, the minimum reduction of number of lady bird beetle over control was found in  $T_5$  (44.57%) followed by  $T_7$  (34.78%).

| Treatments            | Incidence of lady bird beetle per five plants |                |              |               |        |               |  |
|-----------------------|---|----------------|--------------|---------------|--------|---------------|--|
|                       | Vegetative                                    | Early fruiting | Mid fruiting | Late fruiting | Mean   | Incidence     |  |
|                       | stage   | stage          | stage        | stage         |        | increase over |  |
|                       |   |                |              |               |        | control (%)   |  |
| $T_1$                 | 3.17 a  | 4.77 a         | 5.27 a       | 7.63 a        | 5.21 a | 13.26         |  |
| <b>T</b> <sub>2</sub> | 3.10 a  | 4.37 b         | 4.83 b       | 7.17 b        | 4.87 b | 5.87          |  |
| T <sub>3</sub>        | 2.37 c  | 3.83 d         | 4.13 d       | 6.30 d        | 4.16 d | -9.57         |  |
| $T_4$                 | 1.73 e  | 3.13 f         | 3.23 f       | 5.17 f        | 3.32 f | -27.83        |  |
| T <sub>5</sub>        | 1.23 g  | 2.30 h         | 2.43 h       | 4.23 h        | 2.55 h | -44.57        |  |
| T <sub>6</sub>        | 2.17 d  | 3.57 e         | 3.67 e       | 5.70 e        | 3.78 e | -17.83        |  |
| <b>T</b> <sub>7</sub> | 1.60 f  | 2.73 g         | 2.80 g       | 4.83 g        | 3.00 g | -34.78        |  |
| T <sub>8</sub>        | 2.73 b  | 4.17 c         | 4.60 c       | 6.87 c        | 4.60 c |               |  |
| CV (%)                | 3.29  | 1.86           | 1.78         | 1.24          | 1.04   |               |  |
| LSD (0.05)            | 0.13  | 0.12           | 0.12         | 0.13          | 0.08   |               |  |

Table 13: Effect of management practices on incidence of lady bird beetle per plants

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among different treatments the Actara 25 WG

based treatment  $(T_5)$  reduced the highest incidence of lady bird beetle (44.57%) in the brinjal

field. Conversely, the neem oil based treatment (T1) performed as the least hazard. Management

practices, which increased (13.26%) lady bird beetle in the brinjal field rather than synthetic

treatments as well as other botanicals. As a result, the order of rank of efficacy of the treatments to increase number of lady bird beetle per plant was  $T_1 > T_2 > T_8 > T_3 > T_6 > T_4 > T_7 > T_5$ .

# 4.5.2. Spider

The significant variations were observed among the different treatments used for the management practices in terms of number of spider. At vegetative stage, the highest number of spider per plants was recorded in  $T_1$  (2.67 spider/plant), which was statistically similar with  $T_2$  (2.23 spider/plant) and followed by  $T_8$  (1.80 spider/plant) and  $T_3$  (1.67 spider/plant). On the other hand, the lowest number of spider per plants was recorded in  $T_5$  (0.83 spider/plant) which was statistically different from other treatments and followed by  $T_7$  (1.07 spider/plant),  $T_4$  (1.17 spider/plant) and  $T_6$  (1.27 spider/plant) (Table 14). More or less similar trends of number of spider per plants were also recorded at early fruiting stage, mid fruiting stage and late fruiting stage.

In case of mean number of spider, the highest number of spider per plants was recorded in  $T_1$  (3.53 spider/plant) comprised of untreated control, which was significantly different with other treatments and followed by  $T_2$  (3.20 spider/plant),  $T_8$  (2.89 spider/plant) and  $T_3$  (2.60 spider/plant). On the other hand, the lowest mean number of spider per plants was recorded in  $T_5$  (0.96 spider/plant), which was significantly different from other treatments and followed by  $T_7$  (1.74 spider/plant),  $T_4$  (2.06 spider/plant) and  $T_6$  (2.25 spider/plant) (Table 14). Considering the percent increase or decrease the number of spider per plants, the highest 22.15% increase over control was observed in  $T_1$  followed by  $T_2$  (10.73%). On the other hand, the minimum reduction of number of spider over control was found in  $T_5$  (66.78%) followed by  $T_7$  (39.79%).

| Treatments     | Incidence of spider per plot |                |          |          |        |               |  |
|----------------|------------------------------|----------------|----------|----------|--------|---------------|--|
|                | Vegetative                   | Early          | Mid      | Late     | Mean   | Incidence     |  |
|                | stage                        | fruiting stage | fruiting | fruiting |        | increase over |  |
|                |                              |                | stage    | stage    |        | control (%)   |  |
| $T_1$          | 2.67 a                       | 3.27 a         | 3.93 a   | 4.23 a   | 3.53 a | 22.15         |  |
| T <sub>2</sub> | 2.23 b                       | 3.07 b         | 3.43 b   | 4.07 b   | 3.20 b | 10.73         |  |
| T <sub>3</sub> | 1.67 d                       | 2.43 d         | 2.87 d   | 3.43 d   | 2.60 d | -10.03        |  |
| $T_4$          | 1.17 ef                      | 2.17 e         | 2.17 f   | 2.73 f   | 2.06 f | -28.72        |  |
| T <sub>5</sub> | 0.83 g                       | 0.87 g         | 1.07 h   | 1.07 h   | 0.96 h | -66.78        |  |
| T <sub>6</sub> | 1.27 e                       | 2.23 e         | 2.33 e   | 3.17 e   | 2.25 e | -22.15        |  |
| T <sub>7</sub> | 1.07 f                       | 1.73 f         | 1.87 g   | 2.30 g   | 1.74 g | -39.79        |  |
| T <sub>8</sub> | 1.80 c                       | 2.73 c         | 3.17 c   | 3.87 c   | 2.89 c |               |  |
| CV (%)         | 4.01                         | 2.67           | 2.24     | 2.12     | 1.24   |               |  |
| LSD (0.05)     | 0.11                         | 0.11           | 0.09     | 0.11     | 0.05   |               |  |

Table 14: Effect of management practices on incidence of spider per plot

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among different treatments the Actara 25 WG

based treatment  $(T_5)$  reduced the highest incidence of spider (66.78%) in the brinjal field.

Conversely, the neem oil based treatment  $(T_1)$  performed as the least hazard. Management

practices, which increased (22.15%) in the brinjal field rather than synthetic treatments as well as

other botanicals. As a result, the order of rank of efficacy of the treatments to increase number of

spider per plant was  $T_1 > T_2 > T_8 > T_3 > T_6 > T_4 > T_7 > T_5$ .

#### 4.5.3. Ants

The significant variations were observed among the different treatments used for the management practices in terms of number of ants. At vegetative stage, the highest number of ants per plants was recorded in  $T_1$  (3.83 ants/plant), which was statistically different from other treatments and followed by  $T_2$  (3.40 ants/plant),  $T_8$  (3.17 ants/plant) and  $T_3$  (2.83 ants/plant). On the other hand, the lowest number of ants per plants was recorded in  $T_5$  (1.67 ants/plant) which was statistically different from other treatments and followed by  $T_7$  (1.73 ants/plant),  $T_4$  (2.27 ants/plant) and  $T_6$  (2.50 ants/plant) (Table 15). More or less similar trends of number of ants per plants were also recorded at early fruiting stage, mid fruiting stage and late fruiting stage.

In case of mean number of ants, the highest number of ants per plants was recorded in  $T_1$  (4.05 ants/plant) comprised of untreated control, which was significantly different with other treatments and followed by  $T_2$  (3.73 ants/plant),  $T_8$  (3.45 ants/plant) and  $T_3$  (3.20 ants/plant). On the other hand, the lowest mean number of ants per plants was recorded in  $T_5$  (1.40 ants/plant), which was significantly different from other treatments and followed by  $T_7$  (1.84 ants/plant),  $T_4$  (2.23 ants/plant) and  $T_6$  (2.52 ants/plant) (Table 15). Considering the percent increase or decrease the number of ants per plants, the highest 17.39% increase over control was observed in  $T_1$  followed by  $T_2$  (8.12%). On the other hand, the minimum reduction of number of ants over control was found in  $T_5$  (59.42%) followed by  $T_7$  (46.67%).

Table 15: Effect of management practices on incidence of ants per plot

| Treatments            | Incidence of ants per plot |          |          |          |        |                    |
|-----------------------|----------------------------|----------|----------|----------|--------|--------------------|
|                       | Vegetative                 | Early    | Mid      | Late     | Mean   | Incidence increase |
|                       | stage                      | fruiting | fruiting | fruiting |        | over control (%)   |
|                       |                            | stage    | stage    | stage    |        |                    |
| <b>T</b> <sub>1</sub> | 3.83 a                     | 3.87 a   | 4.07 a   | 4.43 a   | 4.05 a | 17.39              |
| T <sub>2</sub>        | 3.40 b                     | 3.67 b   | 3.77 b   | 4.07 b   | 3.73 b | 8.12               |

| T <sub>3</sub> | 2.83 d | 3.10 d | 3.27 d | 3.60 c | 3.20 d | -7.25  |
|----------------|--------|--------|--------|--------|--------|--------|
| T <sub>4</sub> | 2.27 f | 2.53 f | 2.30 f | 1.80 e | 2.23 f | -35.36 |
| T <sub>5</sub> | 1.67 g | 1.57 h | 1.27 h | 1.10 f | 1.40 h | -59.42 |
| T <sub>6</sub> | 2.50 e | 2.73 e | 2.63 e | 2.20 d | 2.52 e | -26.96 |
| T <sub>7</sub> | 1.73 g | 2.23 g | 2.17 g | 1.23 f | 1.84 g | -46.67 |
| T <sub>8</sub> | 3.17 c | 3.30 c | 3.60 c | 3.73 c | 3.45 c |        |
| CV (%)         | 2.50   | 2.63   | 2.52   | 2.99   | 1.33   |        |
| LSD (0.05)     | 0.11   | 0.13   | 0.12   | 0.14   | 0.05   |        |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8 Untreated control.]$ 

From these above findings it was revealed that among different treatments the Actara 25 WG based treatment ( $T_5$ ) reduced the highest incidence of ants (59.42%) in the brinjal field. Conversely, the neem oil based treatment ( $T_1$ ) performed as the least hazard. Management practices, which increased (17.39%) in the brinjal field rather than synthetic treatments as well as other botanicals. As a result, the order of rank of efficacy of the treatments to increase number of ats per plant was  $T_1 > T_2 > T_8 > T_3 > T_6 > T_4 > T_7 > T_5$ 

#### 4.6. Yield attributes

#### **4.6.1.** Effect of management practices on fruit length of brinjal

The significant variations were observed among different treatments used for the management practices in terms of fruit length of brinjal at different growing stage. At early fruiting stage, the lowest fruit length of brinjal was recorded in  $T_8$  (7.40 cm), which was followed by  $T_3$  (8.82 cm),  $T_6$  (9.16 cm) and  $T_2$  (9.34 cm). On the other hand, the highest number of fruit length of brinjal was recorded in  $T_7$  (10.34 cm), which was followed by  $T_5$  (10.03 cm),  $T_4$  (10.02 cm) and  $T_1$  (9.84 cm). More or less similar trends of fruit length of brinjal were also recorded at mid fruiting stage and late fruiting stage (Table 16).

In case of mean fruit length of brinjal per five tagged plants, the highest fruit length of brinjal was recorded in  $T_7$  (11.01 cm) comprised of untreated control, which was significantly different from all other treatments and followed by  $T_5$  (10.29 cm),  $T_4$  (9.99 cm) and  $T_1$  (9.33 cm). On the other hand, the lowest mean fruit length of brinjal per five tagged plants was recorded in  $T_8$  (5.21 cm), which was significantly different from all other treatments and followed by  $T_3$  (6.99 cm),  $T_6$  (7.88 cm) and  $T_2$  (8.78 cm) (Table 16).

In case of percent infested fruit length of brinjal, the lowest percent was recorded in  $T_7$  (3.96%) and followed by  $T_1$  (4.47%),  $T_5$  (5.67%) and  $T_4$  (6.22%). On the other hand, the highest percent was recorded in  $T_8$  (22.39%), which was significantly different from other treatments and followed by  $T_3$  (8.76%),  $T_6$  (7.85%) and  $T_2$  (6.65%) (Table 16). Considering the percent reduction of fruit length of brinjal per five tagged plants, the highest 82.31% over control was achieved in  $T_7$  followed by  $T_1$  (80.04%) and  $T_5$  (74.68%). On the other hand, the minimum reduction of fruit length of brinjal over control was found in  $T_3$  (60.88%) followed by  $T_6$  (64.94%).

| Treatments            | Fruit length (cm) |          |          |         |          |             |
|-----------------------|-------------------|----------|----------|---------|----------|-------------|
|                       | Early             | Mid      | Late     | Mean    | % of     | Infestation |
|                       | fruiting          | fruiting | fruiting |         | infested | reduction   |
|                       | stage             | stage    | stage    |         | fruit    | over        |
|                       |                   |          |          |         | length   | control (%) |
| $T_1$                 | 9.84 c            | 9.83 d   | 8.33 d   | 9.33 d  | 4.47 g   | 80.04       |
| T <sub>2</sub>        | 9.34 d            | 9.33 e   | 7.67 e   | 8.78 e  | 6.65 d   | 70.30       |
| <b>T</b> <sub>3</sub> | 8.82 f            | 8.01 g   | 4.12 g   | 6.99 g  | 8.76 b   | 60.88       |
| $T_4$                 | 10.02 b           | 10.17 c  | 9.78 c   | 9.99 c  | 6.22 e   | 72.22       |
| T <sub>5</sub>        | 10.03 b           | 10.52 b  | 10.32 b  | 10.29 b | 5.67 f   | 74.68       |
| T <sub>6</sub>        | 9.16 e            | 9.17 f   | 5.33 f   | 7.88 f  | 7.85 c   | 64.94       |
| <b>T</b> <sub>7</sub> | 10.34 a           | 12.02 a  | 10.67 a  | 11.01 a | 3.96 h   | 82.31       |
| T <sub>8</sub>        | 7.40 g            | 6.13 h   | 2.10 h   | 5.21 h  | 22.39 a  |             |
| CV (%)                | 0.44              | 0.15     | 0.49     | 0.09    | 1.57     |             |

Table 16. Effect of management practices on fruit length of brinjal during growing season

| LSD (0.05) | 0.76 | 0.05 | 0.05 | 0.05 | 0.22 |      |
|------------|------|------|------|------|------|------|
|            |      |      |      |      |      | <br> |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8: Untreated control.]$ 

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) in reduced infested fruit length per five plants over control (82.31%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the infested fruit length over control (80.04%).

#### 4.6.2. Effect of management practices on fruit girth of brinjal

The significant variations were observed among different treatments used for the management practices in terms of fruit girth of brinjal per five tagged plants at different growing stage. At early fruiting stage, the lowest fruit girth of brinjal per five tagged plants was recorded in  $T_8$  (10.66 cm), which was followed by  $T_3$  (11.17 cm),  $T_6$  (12.16 cm) and  $T_2$  (12.33 cm). On the other hand, the highest fruit girth of brinjal per five tagged plants was recorded in  $T_7$  (13.17 cm), which was statistically similar with  $T_4$  (13.10 cm) and followed by  $T_5$  (13.03 cm) and  $T_1$  (12.67 cm). More or less similar trends of fruit girth of brinjal per five tagged plants were also recorded at mid fruiting stage and late fruiting stage (Table 17).

In case of mean fruit girth of brinjal per five tagged plants, the highest fruit girth of brinjal was recorded in  $T_7$  (13.31 cm) comprised of untreated control, which was significantly different from all other treatments and followed by  $T_5$  (12.57 cm),  $T_4$  (12.36 cm) and  $T_1$  (12.02 cm). On the other hand, the lowest mean fruit girth of brinjal per five tagged plants was recorded in  $T_8$  (9.28

cm), which was significantly different from all other treatments and followed by  $T_3$  (10.72 cm),  $T_6$  (11.44 cm) and  $T_2$  (11.67 cm) (Table 17).

In case of percent infested fruit girth of brinjal, the lowest percent was recorded in  $T_7$  (3.36%) and followed by  $T_5$  (4.58%),  $T_1$  (4.62%) and  $T_4$  (4.73%). On the other hand, the highest percent was recorded in  $T_8$  (12.38%), which was significantly different from other treatments and followed by  $T_6$  (9.45%),  $T_3$  (7.21%) and  $T_2$  (6.58%) (Table 17). Considering the percent reduction of fruit girth of brinjal per five tagged plants, the highest 72.86% over control was achieved in  $T_7$  followed by  $T_5$  (63.00%) and  $T_1$  (62.68%). On the other hand, the minimum reduction of fruit girth of brinjal over control was found in  $T_6$  (23.67%) followed by  $T_3$  (41.76%) and  $T_2$  (46.85%).

| Treatments            | Fruit girth (c | Fruit girth (cm) |          |         |          |              |
|-----------------------|----------------|------------------|----------|---------|----------|--------------|
|                       | Early          | Mid              | Late     | Mean    | % of     | Infestation  |
|                       | fruiting       | fruiting         | fruiting |         | infested | reduction    |
|                       | stage          | stage            | stage    |         | fruit    | over control |
|                       |                |                  |          |         | girth    | (%)          |
| T <sub>1</sub>        | 12.67 c        | 12.07 d          | 11.33 d  | 12.02 d | 4.62 e   | 62.68        |
| T <sub>2</sub>        | 12.33 d        | 11.50 e          | 11.17 e  | 11.67 e | 6.58 d   | 46.85        |
| <b>T</b> <sub>3</sub> | 11.17 f        | 10.67 g          | 10.33 g  | 10.72 g | 7.21 c   | 41.76        |
| $T_4$                 | 13.10 ab       | 12.33 c          | 11.66 c  | 12.36 c | 4.73 e   | 61.79        |
| T <sub>5</sub>        | 13.03 b        | 12.51 b          | 12.18 b  | 12.57 b | 4.58 e   | 63.00        |
| T <sub>6</sub>        | 12.16 e        | 11.13 f          | 11.03 f  | 11.44 f | 9.45 b   | 23.67        |
| T <sub>7</sub>        | 13.17 a        | 12.67 a          | 14.10 a  | 13.31 a | 3.36 f   | 72.86        |
| T <sub>8</sub>        | 10.66 g        | 9.67 h           | 7.50 h   | 9.28 h  | 12.38 a  |              |
| CV (%)                | 0.32           | 0.36             | 0.36     | 0.19    | 3.62     |              |
| LSD (0.05)            | 0.08           | 0.08             | 0.08     | 0.05    | 0.40     |              |

**Table 17**. Effect of management practices on fruit girth of brinjal during growing season

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8: Untreated control.]$ 

From these above findings it was revealed that among the different treatments,  $T_7$  (spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) reduced infested fruit girth per five tagged plants over control (72.86%). Considering the botanical treatments,  $T_1$  (spraying of Neem oil @ 3.0 ml/L of water at 7 days interval) performed as the best treatment in terms of reducing the infested fruit girth per five tagged plants over control (62.68%).

#### 4.6.3. Effect of management practices on single fruit weight of brinjal

The significant variations were observed among different treatments used for the management practices in terms of single fruit weight of brinjal at different growing stage. At early fruiting stage, the lowest single fruit weight of brinjal was recorded in  $T_8$  (17.31 gm), which was followed by  $T_3$  (22.33 gm),  $T_6$  (23.36 gm) and  $T_2$  (26.17 gm). On the other hand, the highest single fruit weight of brinjal was recorded in  $T_7$  (37.53 gm), which was followed by  $T_1$  (33.82 gm),  $T_5$  (32.33 gm) and  $T_4$  (27.29 gm). More or less similar trends of single fruit weight of brinjal were also recorded at mid fruiting stage and late fruiting stage (Table 18).

In case of mean single fruit weight, the highest single fruit weight of brinjal was recorded in  $T_7$  (39.00 gm) comprised of untreated control, which was significantly different from all other treatments and followed by  $T_5$  (36.78 gm),  $T_1$  (34.66 gm) and  $T_4$  (31.78 gm). On the other hand, the lowest mean single fruit weight of brinjal was recorded in  $T_8$  (19.11 gm), which was significantly different from all other treatments and followed by  $T_3$  (23.22 gm),  $T_6$  (24.66 gm) and  $T_2$  (29.22 gm) (Table 18). Considering the percent increase of single fruit weight of brinjal, the maximum 104.08% over control was achieved in  $T_7$  followed by  $T_5$  (92.46%) and  $T_1$  (81.37%). On the other hand, the minimum percent increase of single fruit weight of brinjal over control was found in  $T_3$  (21.51%) followed by  $T_6$  (29.04%).

| Treatments            | Single fruit weight per plant (gm) |              |               |         |                       |
|-----------------------|------------------------------------|--------------|---------------|---------|-----------------------|
|                       | Early fruiting                     | Mid fruiting | Late fruiting | Mean    | Fruit weight increase |
|                       | stage                              | stage        | stage         |         | over control (%)      |
| $T_1$                 | 33.82 b                            | 38.33 b      | 32.33 b       | 34.66 c | 81.37                 |
| $T_2$                 | 26.17 e                            | 33.33 d      | 28.00 c       | 29.22 e | 52.90                 |
| T <sub>3</sub>        | 22.33 g                            | 25.67 f      | 21.67 e       | 23.22 g | 21.51                 |
| $T_4$                 | 27.29 d                            | 36.33 c      | 31.67 b       | 31.78 d | 66.30                 |
| T <sub>5</sub>        | 32.33 c                            | 41.67 a      | 36.33 a       | 36.78 b | 92.46                 |
| T <sub>6</sub>        | 23.36 f                            | 27.33 e      | 23.33 d       | 24.66 f | 29.04                 |
| <b>T</b> <sub>7</sub> | 37.53 a                            | 42.33 a      | 37.33 a       | 39.00 a | 104.08                |
| T <sub>8</sub>        | 17.31 h                            | 21.67 g      | 18.33 f       | 19.11 h |                       |
| CV (%)                | 1.88                               | 1.84         | 2.27          | 1.46    |                       |
| LSD (0.05)            | 0.87                               | 1.03         | 1.01          | 0.73    |                       |

**Table 18.** Effect of management practices on fruit girth of brinjal during growing season

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8: Untreated control.]$ 

From these above findings it was revealed that among the different treatments, T<sub>7</sub> (spraying of

Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval) increased single fruit weight of brinjal

over control (104.08%). Considering the botanical treatments, T<sub>1</sub> (spraying of Neem oil @ 3.0

ml/L of water at 7 days interval) performed as the best treatment in terms of increasing single

fruit weight of brinjal over control (81.37%).

# 4.6.4. Effects of management practices on number of branch per plant, plant height per plot and yield

There were no significant variance among number of branch per plant and plant height per plot at brinjal field at different treatments throughout the growing season of brinjal cultivation.

In case of yield per plot, the highest yield was recorded in  $T_7$  (1534.00 kg/plot) which was followed by  $T_1$  (1475.00 kg/plot),  $T_5$  (1312.00 kg/plot) and  $T_4$  (1199.00 kg/plot). On the other hand, the lowest yield was recorded in  $T_8$  (549.70 kg/plot) which was followed by  $T_3$  (827.10 kg/plot),  $T_6$  (988.30 kg/plot) and  $T_2$  (1132.00 kg/plot).

**Table 19.** Effects of management practices on number of branch per plant, plant height per plot

 and yield

| Treatments            | Number of     | Plant height (cm)/ | Yield (Kg/plot) | Yield (t/ha) |
|-----------------------|---------------|--------------------|-----------------|--------------|
|                       | branch/ plant | plot               |                 |              |
| $T_1$                 | 8.33 a        | 46.20 a            | 1475.00 b       | 24.59 b      |
| T <sub>2</sub>        | 7.33 a        | 40.73 a            | 1132.00 e       | 18.87 e      |
| <b>T</b> <sub>3</sub> | 8.27 a        | 46.33 a            | 827.10 g        | 13.79 g      |
| $T_4$                 | 8.93 a        | 46.73 a            | 1199.00 d       | 19.98 d      |
| T <sub>5</sub>        | 8.33 a        | 48.07 a            | 1312.00 c       | 21.86 с      |
| T <sub>6</sub>        | 8.53 a        | 47.00 a            | 988.30 f        | 16.47 f      |
| <b>T</b> <sub>7</sub> | 7.40 a        | 44.07 a            | 1534.00 a       | 25.57 a      |
| T <sub>8</sub>        | 8.47 a        | 45.27 a            | 549.70 h        | 9.16 h       |
| CV (%)                | 10.25         | 8.86               | 0.09            | 0.09         |
| LSD (0.05)            | 1.42          | 6.82               | 1.70            | 0.05         |

[DAT = Day After Transplanting, In a column, numeric value represents the mean of 3 replications; each replication is derived from 5 plants per treatment; in a column means having similar letter(s) are statistically identical at 0.05 level of probability]

 $[T_1: Spraying of Neem oil @ 3.0 ml/L of water at 7 days interval; T_2: Spraying of Neem seed kernel extract @ 3.0 ml/L of water at 7 days interval; T_3: Spraying of Bioneem plus @ 3.0 ml/L of water at 7 days; T_4: Spraying of Marshal 25 EC @ 3.0 ml/L of water at 7 days interval; T_5: Spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval; T_6: Spraying of Emitaf 20 SL @ 0.1 ml/L of water at 7 days interval; T_7: Spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval; T_8: Untreated control.]$ 

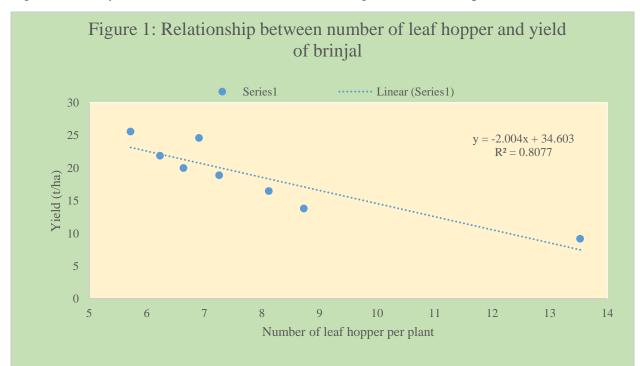
In terms of yield per hectares, the highest yield was recorded in  $T_7$  (25.57 t/ha) which was followed by  $T_1$  (24.59 t/ha),  $T_5$  (21.86 t/ha) and  $T_4$  (19.98 t/ha). On the other hand, the lowest

yield was recorded in T<sub>8</sub> (9.16 t/ha) which was followed by T<sub>3</sub> (13.79 t/ha), T<sub>6</sub> (16.47 t/ha) and T<sub>2</sub> (18.87 t/ha). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of aphid per plant was T<sub>7</sub>> T<sub>1</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

#### 4.7. Relationship between number of insect and yield of brinjal

#### 4.7.1. Leaf hopper

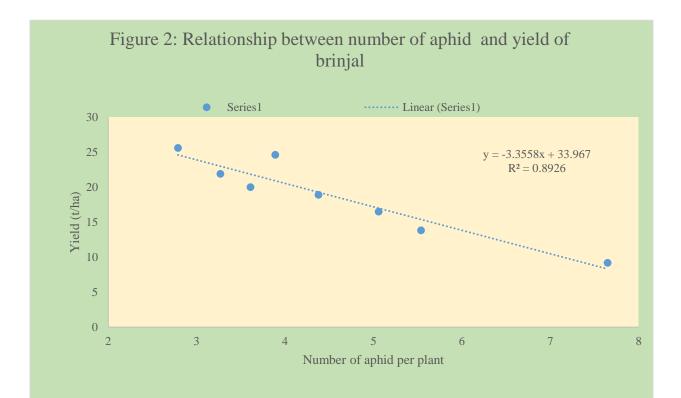
Correlation study was done to establish the relationship between number of leaf hopper per plant and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between the number of leaf hopper per plant and yield of brinjal (Figure 1). It was evident from the Figure 1 that the regression equation y = -2.004x + 34.603 gave a good fit to the data, and the co-efficient of determination (R<sup>2</sup> = 0.8077) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between the number of



leaf hopper per plant and yield of brinjal, i.e., the yield decreased with the increase of the number of leaf hopper per plant during the growing season of brinjal.

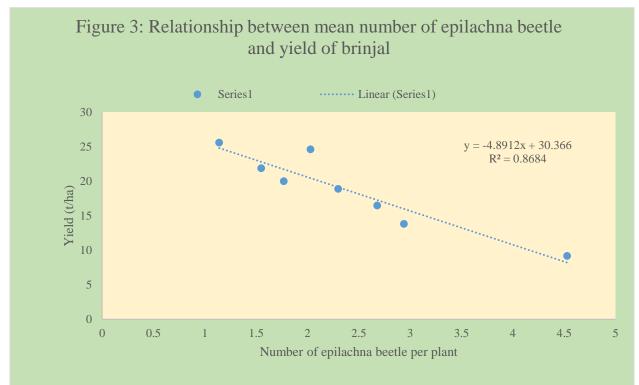
#### 4.7.2. Aphid

Correlation study was done to establish the relationship between number of aphid per plant and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between the number of aphid per plant and yield of brinjal (Figure 2). It was evident from the Figure 2 that the regression equation y = -3.3558x + 33.967 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.8926$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between the number of aphid per plant during the growing season of brinjal.



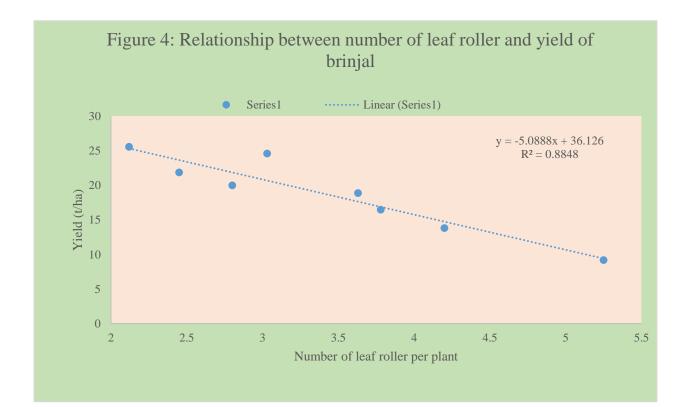
#### 4.7.3. Epilachna beetle

Correlation study was done to establish the relationship between number of epilachna beetle per plant and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between the number of epilachna beetle per plant and yield of brinjal (Figure 3). It was evident from the Figure 3 that the regression equation y = -4.8912x + 30.366 gave a good fit to the data, and the co-efficient of determination ( $\mathbb{R}^2 = 0.8684$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between the number of epilachna beetle per plant and yield of brinjal, i.e., the yield decreased with the increase of the number of epilachna beetle per plant during the growing season of brinjal.



#### 4.7.4. Leaf roller

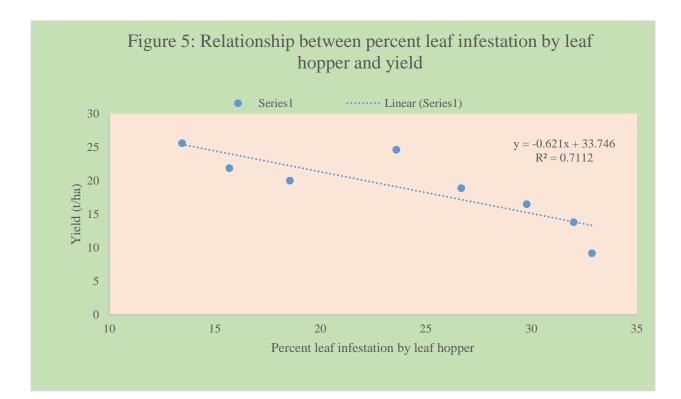
Correlation study was done to establish the relationship between number of leaf roller per plant and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between the number of leaf roller per plant and yield of brinjal (Figure 4). It was evident from the Figure 4 that the regression equation y = -5.0888x + 36.126 gave a good fit to the data, and the co-efficient of determination ( $R^2 =$ 0.8848) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between the number of leaf roller per plant and yield of brinjal, i.e., the yield decreased with the increase of the number of leaf roller per plant during the growing season of brinjal.



#### 4.8. Relationship between percent leaf infestation and yield

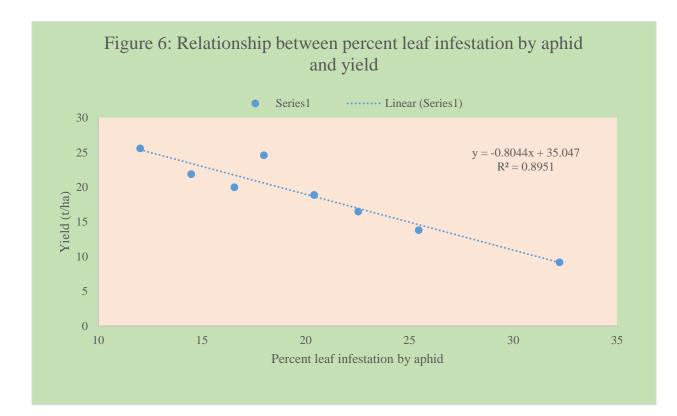
#### 4.8.1. Leaf hopper

Correlation study was done to establish the relationship between percent leaf infestation by leaf hopper and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent leaf infestation by leaf hopper and yield of brinjal (Figure 5). It was evident from the Figure 5 that the regression equation y = -0.621x + 33.746 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.7112$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent leaf infestation by leaf hopper and yield of brinjal, i.e., the yield decreased with the increase of percent leaf infestation by leaf hopper during the growing season of brinjal.



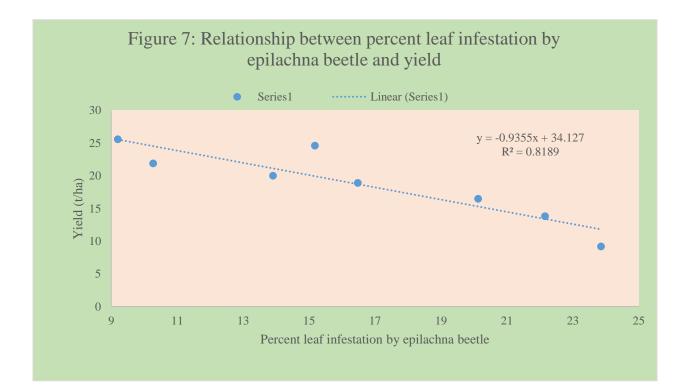
#### 4.8.2. Aphid

Correlation study was done to establish the relationship between percent leaf infestation by aphid and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent leaf infestation by aphid and yield of brinjal (Figure 6). It was evident from the Figure 6 that the regression equation y = -0.8044x + 35.047 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.8951$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent leaf infestation by aphid and yield of brinjal, i.e., the yield decreased with the increase of percent leaf infestation by aphid during the growing season of brinjal.



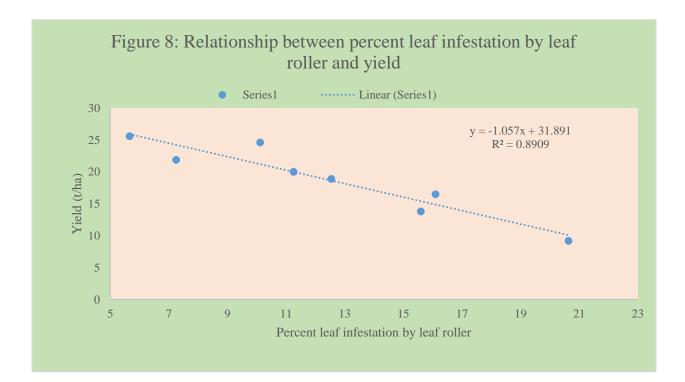
#### 4.8.3. Epilachna beetle

Correlation study was done to establish the relationship between percent leaf infestation by epilachna beetle and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent leaf infestation by epilachna beetle and yield of brinjal (Figure 7). It was evident from the Figure 7 that the regression equation y = -0.9355x + 34.127 gave a good fit to the data, and the coefficient of determination ( $R^2 = 0.8189$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent leaf infestation by epilachna beetle and yield of brinjal, i.e., the yield decreased with the increase of percent leaf infestation by epilachna beetle during the growing season of brinjal.



#### 4.8.4. Leaf roller

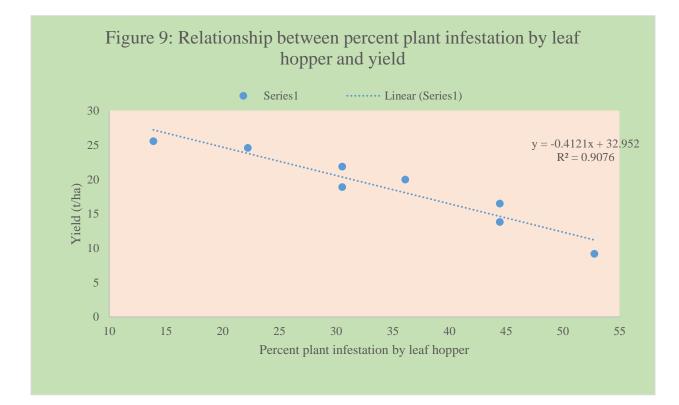
Correlation study was done to establish the relationship between percent leaf infestation by leaf roller and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent leaf infestation by leaf roller and yield of brinjal (Figure 8). It was evident from the Figure 8 that the regression equation y = -1.057x + 31.891 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.8909$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent leaf infestation by leaf roller and yield of brinjal, i.e., the yield decreased with the increase of percent leaf infestation by leaf roller during the growing season of brinjal.



#### 4.9. Relationship between percent plant infestation and yield

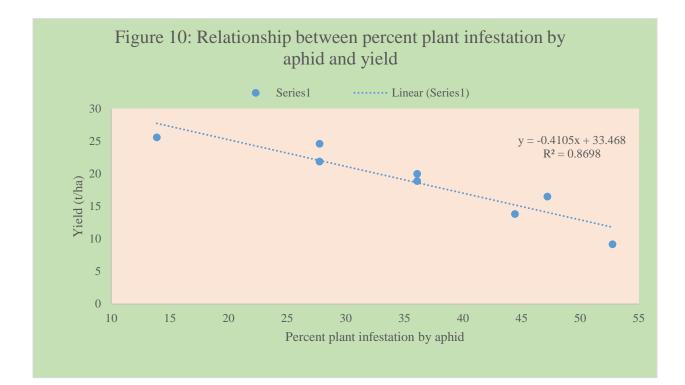
#### 4.9.1. Leaf hopper

Correlation study was done to establish the relationship between percent plant infestation by leaf hopper and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent plant infestation by leaf hopper and yield of brinjal (Figure 9). It was evident from the Figure 9 that the regression equation y = -0.4121x + 32.952 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.9076$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent plant infestation by leaf hopper and yield of brinjal, i.e., the yield decreased with the increase of percent plant infestation by leaf hopper during the growing season of brinjal.



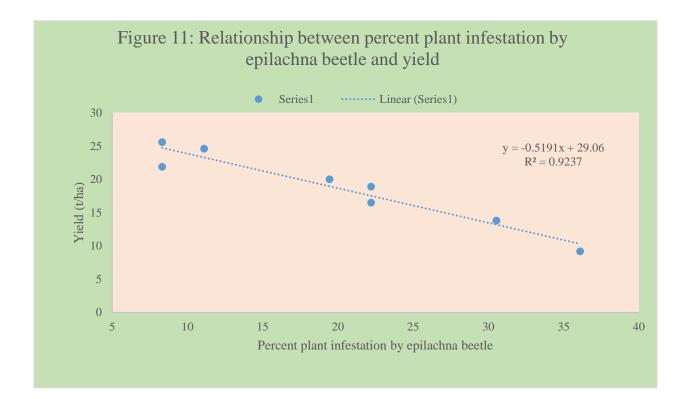
#### 4.9.2. Aphid

Correlation study was done to establish the relationship between percent plant infestation by aphid and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent plant infestation by aphid and yield of brinjal (Figure 10). It was evident from the Figure 10 that the regression equation y = -0.4105x + 33.468 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.8698$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent plant infestation by aphid and yield of brinjal, i.e., the yield decreased with the increase of percent plant infestation by aphid during the growing season of brinjal.



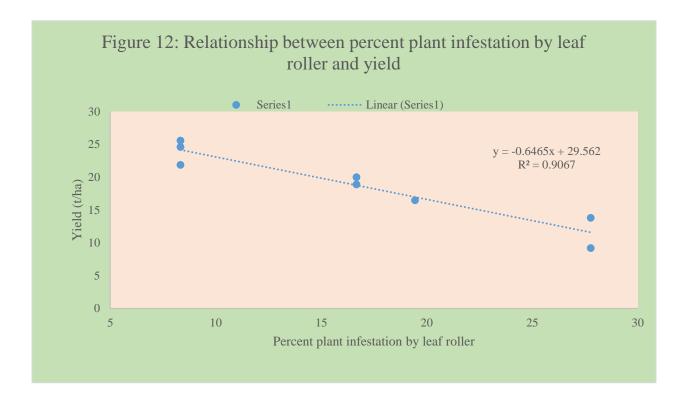
#### 4.9.3. Epilachna beetle

Correlation study was done to establish the relationship between percent plant infestation by epilachna beetle and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent plant infestation by epilachna beetle and yield of brinjal (Figure 11). It was evident from the Figure 11 that the regression equation y = -0.5191x + 29.06 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.9237$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent plant infestation by epilachna beetle and yield of brinjal, i.e., the yield decreased with the increase of percent plant infestation by epilachna beetle during the growing season of brinjal.



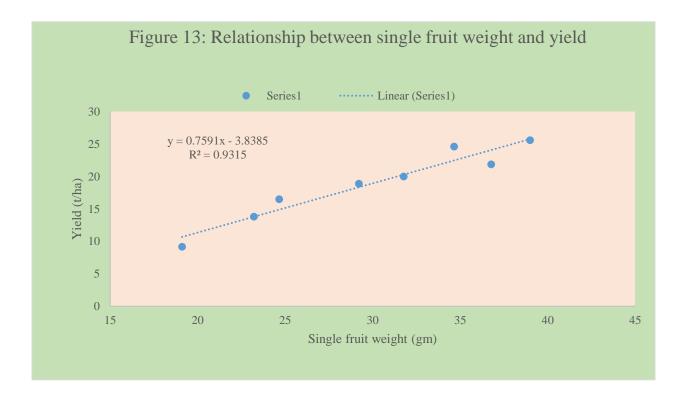
#### 4.9.4. Leaf roller

Correlation study was done to establish the relationship between percent plant infestation by leaf roller and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between percent plant infestation by leaf roller and yield of brinjal (Figure 12). It was evident from the Figure 12 that the regression equation y = -0.6465x + 29.562 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.9067$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a negative relationship between percent plant infestation by leaf roller and yield of brinjal, i.e., the yield decreased with the increase of percent plant infestation by leaf roller during the growing season of brinjal.



#### 4.10. Relationship between single fruit weight and yield of brinjal

Correlation study was done to establish the relationship between single fruit weight and yield (t/ha) of brinjal during the management of sucking and foliage insects. From the study it was revealed that significant correlation was observed between single fruit weight and yield of brinjal (Figure 13). It was evident from the Figure 13 that the regression equation y = 0.7591x - 3.8385 gave a good fit to the data, and the co-efficient of determination ( $R^2 = 0.9315$ ) showed that, fitted regression line had a significant regression co-efficient. From this regression analysis, it was evident that there was a positive relationship between single fruit weight and yield of brinjal, i.e., the yield increased with the increase of single fruit weight during the growing season of brinjal.



#### **CHAPTER V**

## SUMMARY AND CONCLUSION

The experiment was conducted in the experimental field of Sher-e-Bangla Agricultural University, Dhaka, Bangladesh during the period from November, 2017 to March, 2018 to evaluate some management practices applied against sucking and foliage insect pests of brinjal. The experiment consisted of control measures with chemical and botanical.

#### SUMMARY

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing number of leaf hopper per plant over control (57.72%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the leaf infestation over control (48.93%). As a result, the order of rank of efficacy of the treatments applied against brinjal leaf hopper including untreated control in terms of reducing number of brinjal leaf hopper per plant was T<sub>7</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>1</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested leaves by leaf hopper per five plants over control (59.09%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested leaves by leaf hopper per five plants over control (28.19%). As a result, the order of rank of efficacy of the treatments applied against brinjal leaf hopper including untreated control in terms of reducing number of infested leaves by leaf hopper per five plants was T<sub>7</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>1</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested plants by leaf hopper per plot over control (73.68%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested plants by leaf hopper per plot over control (57.90%). As a result, the order of rank of efficacy of the treatments applied against brinjal leaf hopper including untreated control in terms of reducing number of infested plants by leaf hopper per plot was T<sub>7</sub>> T<sub>1</sub>> T<sub>5</sub>> T<sub>2</sub>> T<sub>4</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing number of aphid per plant over control (63.53%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the leaf infestation over control (49.15%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of aphid per plant was T<sub>7</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>1</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested leaves by aphid per five plants over control (62.67%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested leaves by aphid per five plants over control (42.20%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of infested leaves by aphid per five plants was T<sub>7</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>1</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested plants by aphid per five plants over control (73.68%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested plants by aphid per five plants over control (47.37%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of infested plants by aphid per five plants was T<sub>7</sub>> T<sub>1</sub>> T<sub>5</sub>> T<sub>2</sub>> T<sub>4</sub>> T<sub>3</sub>> T<sub>6</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing number of epilachna beetle per plant over control (74.83%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of epilachna beetle over control (55.19%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of aphid per plant was T<sub>7</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>1</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment  $T_7$  comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested leaves by epilachna beetle per five plants over control (61.44%). Considering the botanical treatments,  $T_1$  comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested leaves by epilachna beetle per five plants over control (36.38%). As a result, the order of rank of efficacy of the treatments applied against epilachna beetle including untreated control in terms of reducing number of infested leaves by epilachna beetle per five plants was  $T_7 > T_5 >$  $T_4 > T_1 > T_2 > T_6 > T_3 > T_8$ . The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested plants by epilachna beetle per five plants over control (76.93%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested plants by epilachna beetle per five plants over control (69.23%). As a result, the order of rank of efficacy of the treatments applied against epilachna beetle including untreated control in terms of reducing number of infested plants by epilachna beetle per five plants was T<sub>7</sub>> T<sub>5</sub>> T<sub>1</sub>> T<sub>4</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing number of leaf roller per plant over control (59.62%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the leaf infestation over control (42.29%). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of aphid per plant was T<sub>7</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>1</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested leaves by leaf roller per five plants over control (72.58%). Considering the botanical treatments, T<sub>1</sub> comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested leaves by leaf roller per five plants over control (50.97%). As a result, the order of rank of efficacy of the treatments applied against leaf roller including untreated control in terms of reducing number of infested leaves by leaf roller per five plants was T<sub>7</sub>> T<sub>5</sub>> T<sub>1</sub>> T<sub>4</sub>> T<sub>2</sub>> T<sub>3</sub>> T<sub>6</sub>> T<sub>8</sub>.

The treatment T<sub>7</sub> comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing the number of infested plants by leaf roller per plot over control (70.01%). Considering the botanical treatments,  $T_1$  comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the number of infested plants by leaf roller per plot over control (70.01%). As a result, the order of rank of efficacy of the treatments applied against leaf roller including untreated control in terms of reducing number of infested plants by leaf roller per plot was  $T_7 > T_1 > T_5 > T_2 > T_4 > T_6 > T_3 > T_8$ . Actara 25 WG based treatment ( $T_5$ ) reduced the highest incidence of lady bird beetle (44.57%) in the brinjal field. Conversely, the neem oil based treatment  $(T_1)$  performed as the least hazard. Management practices, which increased (13.26%) lady bird beetle in the brinjal field rather than synthetic treatments as well as other botanicals. As a result, the order of rank of efficacy of the treatments to increase number of lady bird beetle per plant was  $T_1 > T_2 > T_8 > T_3 > T_6 > T_4 > T_7 > T_5$ . Actara 25 WG based treatment ( $T_5$ ) reduced the highest incidence of field spider (66.78%) in the brinjal field. Conversely, the neem oil based treatment  $(T_1)$  performed as the least hazard. Management practices, which increased (22.15%) in the brinjal field rather than synthetic treatments as well as other botanicals. As a result, the order of rank of efficacy of the treatments to increase number of field spider per plant was  $T_1 > T_2 > T_3 > T_3 > T_4 > T_7 > T_5$ .

Actara 25 WG based treatment ( $T_5$ ) reduced the highest incidence of ants (59.42%) in the brinjal field. Conversely, the neem oil based treatment ( $T_1$ ) performed as the least hazard. Management practices, which increased (17.39%) in the brinjal field rather than synthetic treatments as well as other botanicals. As a result, the order of rank of efficacy of the treatments to increase number of ants per plant was  $T_1 > T_2 > T_8 > T_3 > T_6 > T_4 > T_7 > T_5$ .

The treatment  $T_7$  comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing infested fruit length per five plants over control (82.31%). Considering the botanical treatments,  $T_1$  comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the infested fruit length over control (80.04%).

The treatment  $T_7$  comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval in reducing infested fruit girth per five tagged plants over control (72.86%). Considering the botanical treatments,  $T_1$  comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days interval performed as the best treatment in terms of reducing the infested fruit girth per five tagged plants over control (62.68%).

The treatment  $T_7$  comprised with spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days in increasing single fruit weight of brinjal over control (104.08%). Considering the botanical treatments,  $T_1$  comprised with spraying of Neem oil @ 3.0 ml/L of water at 7 days performed as the best treatment in terms of increasing single fruit weight of brinjal over control (81.37%).

There were no significant variance among number of branch per plant and plant height per plot at brinjal field at different treatments throughout the growing season of brinjal cultivation.

The treatment T<sub>7</sub> (25.57 t/ha) which was followed by T<sub>1</sub> (24.59 t/ha), T<sub>5</sub> (21.86 t/ha) and T<sub>4</sub> (19.98 t/ha). On the other hand, the lowest yield was recorded in T<sub>8</sub> (9.16 t/ha) which was followed by T<sub>3</sub> (13.79 t/ha), T<sub>6</sub> (16.47 t/ha) and T<sub>2</sub> (18.87 t/ha). As a result, the order of rank of efficacy of the treatments applied against aphid including untreated control in terms of reducing number of aphid per plant was T<sub>7</sub>> T<sub>1</sub>> T<sub>5</sub>> T<sub>4</sub>> T<sub>2</sub>> T<sub>6</sub>> T<sub>3</sub>> T<sub>8</sub>.

## CONCLUSION

From the present study, it may be concluded that incidence of sucking and foliar insects (leaf hopper, aphid, epilachna beetle, leaf roller etc.) of brinjal was significantly varied among the treatments. The overall study revealed that the highest performance was achieved from  $T_7$  spraying of Ripcord 20 EC @ 1.0 ml/L of water at 7 days interval.  $T_5$  spraying of Actara 25 WG @ 0.2 gm/L of water at 7 days interval and  $T_1$  spraying of Neem oil @ 3.0 ml/L of water at 7 days interval also showed better performance against sucking and foliar insects of brinjal.  $T_1$  might increase the number of beneficial arthropods (lady bird beetle, field spider, ants etc.), weight of single fruit, length of fruit, girth of fruit and yield. Considering the results of the present study and environmental issues it can be concluded that  $T_1$  can be used for the management of sucking and other foliar insect pests of brinjal.

Considering the findings of the study the following recommendations can be drawn:

- 1. Chemical insecticides should be less used for management practices against sucking and foliar insects of brinjal.
- Botanical insecticides should be more used to increase the number of beneficial arthropods in the brinjal field.
- 3. Further study should be needed in different locations of Bangladesh for accuracy of the results obtained from the present experiment.

## **CHAPTER VI**

### REFERENCES

- Afzal, M. and Ghani, M. A. (1953). Cotton jassid in Panjab. The Pakistan Association for the Advancement of Science, University institute chemistry, Lahore. p:101.
- Alam, M. Z. (1969). Insect pest of vegetable and their control in East Pakistan published by the Agriculture Information Services, Department of Agriculture, 3, R. K. Mission Road, Dhaka-3, East Pakistan.
- Alam, S. N., Rashid M. A., Rouf F. M. A., Jhala R. C., Patel J. R. S., Satpathy T. M., Shivalingaswamy S. R., Wahundeniya I., Cork A., Ammaranan C. and Talekar S. N. (2003). Development of an integrated pest management strategy for eggplant fruit and shoot boreer in South Asia. Technical Bulletin 28. AVRDC The World Vegetable Center, Shanhua, Taiwan. p: 66.
- Alam, S. N., Hossion, M. I., Rouf, F. M. A., Jhala, R. C., Patel, M. G., Nath, L. K., Sengupta, A., Baral, K., Shylesha, A. N., Satpathy, S., Shivalingaswamy, T. M., Cork, A. and Talekar, N. S. (2006). Control of eggplant and shoot and fruit borer in South Asia. Technical Bulletin 36, AVRDC- The World Vegetable Center, Shanua, Taiwan. p: 88.
- Alam, S. N., Sarker, D and Rahman, A. K. M. Z. (2005). Annual Report. Division of Entomology, Bangladesh Agricultural Research Institute, Joydebpur, Gazipur. p: 82.
- Ali, M. I. (1987). Report on the entomological experiments on cotton. Division of entomology, Bangladesh Agricultural Research Institute, Joydebpur, Gazipur. p: 34.
- Ali, M. I. (1990). The development of systems due to attack of the cotton jassid, *Amrasca biguttula* (Homoptera: Jassidae) on cotton. *Bangladesh J. Zool.* **18**(2): 211-214.

- Ali, M. I. and Karim, M. A. (1994). Spraying threshold level of cotton jassid, *Amrasca devastans* (Dist.) on cotton in Bangladesh. *Bangladesh J. Zool.* 2l(1): 47-54.
- Anand, G. K. S., Sharma, R. K. and Shankarganesh, K. (2013). Efficacy of Newer Insecticides against Leaf Hopper and Whitefly Infesting Brinjal and its Effect on Coccinellids. *Pesticide Research Journal.* 25(1): 6-11.
- Anonymous, (1996). Integrated control of the brinjal shoot and fruit borer, *Leucinodes orbonalis* Guenee at Jessore. **In**: Annual Research Report of (1993-94). BARI, Joydebpur, Gazipur, Bangladesh. pp: 44-46.
- Anonymous. (2005). Development of management approach against jassid infestation on lady's finger (*Abelmoschus esculentus*). Annual Research Report, Entomology Division.
   Bangladesh Agricultural Research Institute. Gazipur, Bangladesh. pp: 46-48.

AphlD. (2012). Aphis fabae. AphID. 2012. Retrieved 2013-01-02.

- Atwal, A. S. (1986). Agricultural pests of India and Southeast Asia. Kalyani Publish Ludhiana p: 502.
- Barroga, G. F. and Bernardo, E. N. (1993). Biology, feeding behavior and damage of the cotton leafhopper Amrasca biguttula biguttula Ishida on susceptible and resistant varieties of okra Abelmoschus esculentus L. Moench. Philippine Entomologist. 9(2): 186-200.
- BBS. (2017). Year book of Agricultural Statistic of Bangladesh Ministry of planning, Dhaka.
- Berim, M. N. (2009). "Aphis fabae Scopoli Black Bean Aphid". Interactive Agricultural Ecological Atlas of Russia and Neighboring Countries. AgroAtlas. Retrieved 2013-01-03.
- Berry, R. E. and Taylor, L. R. (1968). "High-Altitude Migration of Aphids in Maritime and Continental Climates". *J Ani. Eco.* **37**(3): 713–722. doi:10.2307/3084. JSTOR 3084.

- Bhaduri, N., Ram, S. and Patil, B. D. (1989). Evaluation of some plant extracts as protectants against the pulse beetle, *Callosobruchus maculates* (Fab.) infestaing cowpea seeds. J. *Entomol. Res.* 9(2): 183-187.
- Bhargava, K. K., Sharma, H. C. and Kaul, C. L. (2001). Bioefficacy of insecticides against okra jassid and fruit borer. *Pest Manage. Econ. Zool.* **9**(2): 193 -195.
- Bhat, M. G., Joshi, A. B. and Singh, M. (1984). Relative losses of seed cotton yield by jassid and bollworm in some cotton genotypes (*Gossypium hirsutum* L.). *Indian J. Entomol.* 46: 169-173.
- Bohlen, E. (1984). Cotton pests in Bangladesh. A field guide for identification and control. Bangladesh Cotton development board, Dhaka, Bangladesh. pp: 1-6.
- Borah, R. K. (1995). Incidence of jassid leaf roller in relation to the time of brinjal planting in the hill zone of Assam. *Ann. Agril. Res.* **16**(2): 220-221.
- Chinery and Michael. (1993). Collins Field Guide to the Insects of Britain and Northern Europe. Harper Collins. ISBN 0002199181.
- Chongtham, N. S., Ibohal, K, H. and John, W. S. (2009). Laboratory Evaluation of Certain Cow Urine Extract of Indigenous Plants Against Mustard Aphid, *Lipaphis erysimi* (Kaltenbach) Infesting Cabbage. *Hexapoda*. pp: 11–13.
- Cloyd, R. A. and Bethke, J. A. (2011). Impact of neonicotinoid insecticides on natural enemies in greenhouse and interiorscape environments. *Pes. Management Sci.* **67**: 3-9.
- CSRIO. (2005). Balancing Act: a Triple Bottom Line Analysis of the Australian Economy. University of Sydney and CSRIO Sustainable Ecosystems. Canberra, ACT. pp: 4 vols.
- Cueva, F. M. D., Pascual, C. B., Bajet, C. M. and Dalisay, T. U. (2015). Pests and diseases of economically important crops in the Philippines. Pest management council of the

Philippines, Inc c/o crop protection cluster, University of the Philippines Los Barios, College, Laguna.

Davidson, R. H. (1987). Insect pest of farm garden and orchard. Eighth edition. Canada. 278p.

- Dharpure, S. R. (2003). Changing scenario of insect pests of polato in *Satpura plateau* of Madhya Pradesh. *J. Indian Potato Assoc.* **29**(3/4): 135-138.
- Drees, C., Hufner, S., Matern, A., Neve, G. and Assmann, T. (2009). Repeated sampling detects gene flow in a flightless ground beetle in a fragmented landscape. Wiley online library. 148(1).
- El-Shafie, H. A. F. (2001). The use of neem products for sustainable management of homopterous key pests on potato and eggplant in the Sudan. Ph. D. Thesis, University of Giessen, Germany.
- El-Tom, H. A. (1987). Integrated pest management for cotton in Bangladesh FAO/UNDP cotton improvement programme. Cotton Research Station, Rangpur, Bangladesh. p.43.
- FAO (Food and Agricultural Organization). (1995). Celebrates its 50th Anniversary Agricultural magagine for the Middle East and Arab world Sept-Oc. 1995. Issue No. 7.
- FAO (Food and Agriultural Organization). (2003). Inter country programme for integrated pest management in vegetables in south and South-East Asia. Eggplant integrated pest management: An ecological guide. p: 177.
- Fiswick, R. B. (1988). Pesticide residues in gain arising from postharvest treatments. *Aspects Appl. Biol.* **17**(2): 37-46.
- Frisbie, R. E. (1984). Guidelines for integrated control of cotton pests. FAO plant production and protection paper 48. FAO, Rome, Italy. pp: 187.

- Gahukar, R. T. (2000). Use of neem products pesticides in cotton pest management. *International J. Pest Manage.* **46**(1): 149-160.
- Godfrey, L. D.; Trumble, J. T. (2009). "UC IPM Pest Management Guidelines: Celery". UC IPM Online. Retrieved 2013-01-03.
- Goodland, R., Watson, C. and Ladac, G. (1985). Biocides bring, poisoning and pollution to third world. The Bangladesh observer, 16<sup>th</sup> and 17<sup>th</sup> January. 1985.
- Hagen, K. S. and Franz, J. M. (1973). A history of biological control. In: Smith, R. F., Mittler, T. E. and Smith, C. N. (eds.). History of Entomology. Ann. Rev. Inc. Palocetto, California. p: 872.
- Hill, L. (2012). "The currant lettuce aphid, Nasonovia ribisnigri arrives in Tasmania: Part 1". Victorian Entomologist. 42(2): 29–31.
- Hillocks, R. J. (1995). Integrated management of insect pests, diseases and weeds of cotton in Africa. Springer Science+Business Media B.V., Formaly Kluwer Academic Publishers B.V. 1(1): 31-47.
- Hossain, S. M. A., Rahman, H. and Rahman, F. (2003). Effect of soap water against sucking pests of cotton. Annual. Research. Report. Cotton Development Board. Khamarbari, Farmgate, Dhaka. <u>http://onlinelibrary.wiley.com/doi/10.1111/j.1095-8312.1979.tb00038.x/abstract. Retrieved 2011-12-21</u>.

Husain, M. (1984). Controlling rice borers under Bangladesh conditions. *Pestology*. 8(8): 28-30.

- Husain, M. (1993). Anistakar Kit-patango Daman (control of harmful insects). Bangla Academy, Dhaka. pp: 220.
- HYPP. (2013). Beet leaf aphid, Bean aphid, Black bean aphid. HYPP Zoology. Retrieved 2013-01-02.

- Inee, G., Dut, B. C. and Gogoi, I. (2000). Seasonal abundance of cotton jassid, *Amrasca biguttula biguttula* on okra. J. Agril. Sci. Soc. **13**(1): 22-26.
- Isard, S. A., Irwin, M. E. and Hollinger, S. E. (1990). "Vertical Distribution of Aphids (Homoptera: Aphididae) in the Planetary Boundary Layer". *Enviro. Entom.* **19**(5): 1473–1484.
- doi:10.1093/ee/19.5.1473.
- Jacob, P. S., Ramasubbara, V. and Punnaiah, K. C. (2000). Leafhopper fauna associated with oil seeds crops in andhra Pradesh, India. *Pest Manage. Econo. Zool.* **8**(1): 11-27.
- John, Y. (2001). Hike Pennsylvania: An Atlas of Pennsylvania's Greatest Hiking Adventures. Guilford, Connecticut: The Globe Pequot Press. p.: 127. ISBN 0-7627-0924-3.
- John M. T., Kasprowicz, L., Malloch, G. L. and Fenton, B. (2009). Tracking the global dispersal of a cosmopolitan insect pest, the peach potato aphid. *BMC Ecol.* **9**: 13. doi:10.1186/1472-6785-9-13. PMC 2687420. PMID 19432979.
- John, A. and Immaraju. (1997). The commercial use of *Azadirachtin* and its integration into viable pest control programs. *Pesticide Sci.* **54**(3): 285-289.
- Johnson, C. G. (1963). The aerial migration of insects, pp. 188-194. In T. Eisner & E.O. Wilson [eds.], The insects. W.H. Freeman, San Francisco.
- Johnson, M. S. (2001). Acropyga and Azteca Ants (Hymenoptera: Formicidae) with Scale Insects (Sternorrhyncha: Coccoidea): 20 Million Years of Intimate Symbiosis. American Museum Novitates. 35: 1–18.
- Kavadia, V. S., Pareek, B. L. and Sharma. (1984). Residues of malathion and carbaryl in stored sorghum. *Bull. Grain. Tech.* 22(3): 247-250.

- Kennedy, J. S.; Booth, C. O. (1951). Host alternation in *Aphis fabae* Scop. I. feeding preferences and fecundity in relation to the age and kind of leaves. *Ann. App. Biol.* 38(1): 25–64. doi:10.1111/j.1744-7348.1951.tb07788.x.
- Kumawat, R. L., Pareek, B. L. and Meena, B. L. (2000). Seasonal incidence of jassid and whitefly on okra and their correlation with abiotic factor. *Ann. Bio. Hiss.* **16**(2): 167-169.
- Latif, M. A. (2007). Ph. D. Dissertation on, "Effectiveness of some insecticides in managing brinjal shoot and fruit borer, *Leucinodes orbonalis* Guenee and their impact on arthropod biodiversity and soil microbial respiration", Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Salna, Gazipur, Bangladesh.
- Lawrence, A. D., Williams and Mansingh, A. (1996). The insecticidal and acaricidal actions of compounds from *Azadirachta indica* (A. Juss.) and their use in tropical pest management. *Integrated Pest Manage. Rev.* 1(3): 133-145.
- Luckmann, W. H. and Metcalf, R. L. (1975). The pest management concept. **In:** Metcalf, R. L. and Luckmann, W. H. (eds.). Introduction to Insect Pest Management. John wiley and Sons, New York. pp: 3-35.
- Mall, N. P., Pandy, R. S., Singh, S. V. and Singh. S. K. (1992). Seasonal incidence of insect pest and estimation of the losses caused by shoot and fruit borer on brinjal. *Indian. J. Ent.* 53(3): 241-246.
- Mamun, M. R. (2006). Host preference and integrated management of jassid (Amrasca devastans) (Distant). Homoptera cicadellidae. MS thesis. Sher-e-bangla Agricultural University. Dhaka. p: 27.
- Martin, N. A. (2018). Hadda beetle- *Epilachna vigintioctopunctata*. Interesting Insects and other Invertebrates. New Zealand Arthropod Factsheet Series Number 38.

http://nzacfactsheets.landcareresearch.co.nz/Index.html. Date Accessed. ISSN 1179-643X.

- Misra, H. P. and Senapati, B. (2003). Evaluation of new insecticides against okra jassid (*Amrasca biguttula biguttula*). *Indian J. Agric. Sci.* **73**(10): 576-578.
- Mote, U. N. and Bhavikatti, S. (2003). Efficacy of chemical and non- chemical insecticides against major pests of brinjal in kharif season. *J. Appl. Zool. Res.* **14**(1): 54-56.
- Munakata, K. (1997). Insect feeding deterents in plants. p: 93-102. In: Chemical control of insect behavior. Shorey, H. H. and Mekelvy, J. (eds.). Jhon Wiley and Sons, New york. p: 522.
- Muthukumar, M. and Kalyanasundaram, M. (2003b). Influence of abiotic factors on the incidence of major insect pests in brinjal (*Solanum melongena* L.). *South Indian Hort*. 51(1/6): 214-218.
- Nair, M. R. G. K. (1986). Insects and Mites of Crop in India. Printed in India at Allied Publishers (Pvt.) Ltd. New Delhi, India. P: 106.
- Nandagopal, V. and Soni, V. C. (1992). Residual effect of some insecticides and neem-oil against jassid on groundnut. *J. Maharashtra Agril. Univ.* **17**(3): 420-422.
- Narayanasamy, P. (2002). Botanical pesticides for effective plant protection. The Hindu National daily Newspaper, India.
- Navasero, M. V. (2003). Population dynamics of arthropods associated with eggplant. Terminal Report. UPLB-Crop Life Philippines.
- Nayer, K. K., Ananthakrisnan, T. N. and David, B. V. (1984). General and applied Entomology. Tata Macgraw Hill Publishing Company Ltd. New Delhi, India. p: 589.

Nonnecke, L. (1989). Vegetable production. 1989<sup>th</sup> ed. Springer.

- Ofori, O.D. and Sackey, J. (2003). Field evaluation of non-synthetic insecticides for the management of insects pests of okra, *Abelmoschus esculentus* (L.) in Ghana. Sinet. *Ethiopian J. Sci.* 26(2): 145-150.
- Padwal, G. K., Singh, K. S. and Sharma, S. K. (2016). Major insect pests of brinjal and their management. *Popular Kheti*. 4(2): 48-53.
- Panchabhavi, K. S., Kulkarni, K. A., Veeresh, G. K., Hiremath, P. C. and Hegde, R. K. (1990). Comparative efficiency of techniques for assessing loss due to inset pests in upland cotton. *Indian J. Agril. Sci.* **60** (4): 252-254.
- Patel, Z. P. and Patel, J. R. (1998). Resurgence of jassid, *Amrasca biguttula biguttula* in brinjal and development strategy to overcome the resurgence in brinjal. *Indian J. Entomol.* 60(2): 152-164.
- Patel, Z. P. and Patel, Z. R. (1998). Effect of botanicals on behavioural response and growth of jassids, *Amrasca biguttula biguttula*. *Indian J. Plant Protec.* 24: 28-32.
- Pimentel, D. (1981). An overview of integrated pest management (Mimeograph). Dept. of Entomology, Section of Ecology and Systematic, Cornell University, Ithaca, New York. p: 52.
- Pip Courtney (2005). Scientist battles lettuce aphid. Landline. Retrieved January 1, 2007.
- Poonia, F. S. (2005). Population dynamics of *Empoasca kerri pruthi*, a jassid pest on cowpea crop in arid region of Rajasthan. Arid legumes for sustainable agriculture and trade. 1: 179-180.
- Rao, N. S. and Rajendran, R. (2002). Joint action potential of neem with other plant extracts against the leaf hopper, *Amrasca devastans* (Distant) on okra. *Pest Manage. Econ. Zool.* 10(2): 131-136.

- Rashid, M. M. (1999). Begun Paribarer Shabji. **In:** Shabji Biggan (in Bangla). First edn. Bangla Academy, Dhaka, Bangladesh.
- RIR. (2013). Black bean aphid. Rothamsted Insect Research. Rothamsted Research. Retrieved 2013-01-03.
- Rote, N. B., Patel, B. K., Mehta, N. P., Shah, A. H. and Raza, K. R. (1985). Threshold level of *Amrasca biguttula* causing economic injury to cotton. *Indian. J. Agril.* **35**(7): 491-492.
- Sathyaseelan, V. and Bhaskaran, V. (2010). Efficacy of some native botanical extracts on the repellency property against the pink mealy bug, *Maconellicoccus hirsutus* (green) in mulberry crop. Recent Research in Science and Technology 2010. ISSN: 2076-5061. 2(10): 35-38.
- Schneider, A. and Madel, G. (1992). Fecundity and vitality of adult parasitoids following exposure to neem (*Azadirachta indica*) treated surfaces. *Mitteilungen der Deutschen Gesellschaft fur Allgemeine und Angewandate Entomologie*. **8**(1/3): 273-278.
- Sexena, R. C., Dixit, O. P. and Harshan, V. (1992). Insecticidal action of Lantana camara against Callosobruchus chinensis (Coleoptera: Bruchidae). J. Stored Prod. Res. 28(4): 279-281.
- Shrestha, P., Koirala, P. and Tamrakar, A. S. (2010). Knowledge, practice and use of pesticides among commercial vegetable growers of Dhading district, Nepal. J. Agric. Environ. 11: 95-100.
- Singh, A. K. and Kumar, M. (2003). Efficacy and economics of neem based products against cotton jassid, Amrasca biguttula biguttula Ishida in okra. Crop Res. Hisar. 26(2): 271-274.

- Singh, J., Dhaliwal, Z. S., Sandhu, S. S. and Sidhu, A. S. (1990). Temporal changes in the dispersion of populations of three homopterous insect pests in upland cotton. *Insect Sci. app.* **11**(1): 73-77.
- Singh, S. P. and Choudhary, S. K. (2001). Chemical control of jassid (*Amrasca biguttula biguttula Ishida*) on Okra. J. Res. Binsa Agric. Univ. **13**(2): 215-216.
- Singh, W., Kotwal, O. R., Singh, R. W. and Singh, R. (1991). Evaluation of some contact insecticides for the control of jassid (*Amrasca biguttula biguttula Ishida*) on okra. *Indian J. Plant Prot.* 19(2): 182-184.
- Srinivasan, R. (2009). Insect and mite pests on eggplant: a field guide for identification and arrangement. AVRDC–The World Vegetable Center, Shanhua, Taiwan. AVRDC publication No. 09-729. p: 64.
- Shukla, R. P. (1989). Population fluctuation of *Leucinodes orbonalis* and *Amrasca biguttula* biguttula in brinjal (Solanum melongena) in relation to abiotic factors Meghalaya.Indian J. Agric. Sci. 59(4): 260-264.
- Talerico, L. R., Newton, M. C. and Valentine T. H. (1978). Pest-control decisions by decision-Tree analysis. J. Forestry. 76(1): 16-19.
- Toba, H. H., Kishaba, A. N., Bohn, G. W. and Hield, H. (1977). Protecting muskmelon against aphid-borne viruses. *Phytopathology* . **67**: 1418-1423.

USDAFS. (1998). United States Department of Agriculture Forest Service. p.: 28.

USDAFS. (2011). United States Department of Agriculture Forest Service. p.: 193.

Walker, F. (1863). List of the Specimens of Lepidopterous Insects in the Collection of the British Museum. Part XXVII. Crambites & Tortricites. British Museum (Natural History), London. pp.: 1-286.

- Way, M. J. and Banks, C. J. (1964). Natural mortality of eggs of the black bean aphid, *Aphis fabae* Scop., on the spindle tree, *Euonymus europaeus* L. Ann. App. Bio. 54(2): 255–267. doi:10.1111/j.1744-7348.1964.tb01189.x.
- Yadgirwar, P. V., Radke, S. G. and Parlawar, N. D. (1994). Efficacy of combinations of synthetic pyrethroids and organophosphatic compounds against cotton jassids. J. Soils and Crops. 4(2): 158-159.
- Żyła, D., Homan, A. and Wegierek, P. (2017). "Polyphyly of the extinct family Oviparosiphidae and its implications for inferring aphid evolution (Hemiptera, Sternorrhyncha)". *PLOS One*. **12**(4): e0174791.
- doi:10.1371/journal.pone.0174791. PMC 5405925. PMID 28445493.

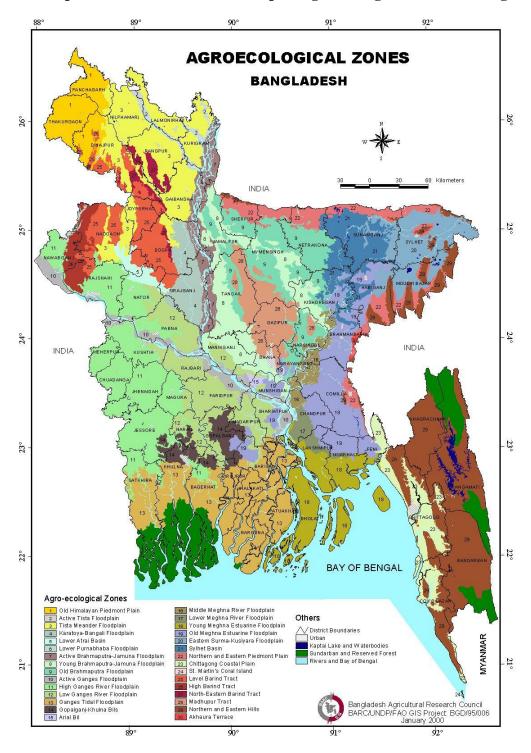
## **CHAPTER VII**

## **APPENDICES**

Appendix I: Monthly record of air temperature, rainfall and relative humidity of the experimental site during the period from November 2017 to March 2018

| Month    | Temperature ( |         | Relative humidity | Rainfall (mm) |
|----------|---------------|---------|-------------------|---------------|
|          | Maximum       | Minimum | (%)               | (Total)       |
| November | 30.2          | 20.6    | 67                | 6.0           |
| December | 26.8          | 17.1    | 76                | 33.0          |
| January  | 23.6          | 12.6    | 68                | 0.0           |
| February | 29.2          | 18.1    | 61                | 20.0          |
| March    | 33.3          | 22.3    | 59                | 3.0           |

Source: Bangladesh Meteorological Department (Climate and Weather Division), Agargoan, Dhaka- 1207.



Appendix II. Experimental location on the map of Agro-ecological Zones of Bangladesh.

Source: Bangladesh Agricultural Research Council, Khamarbari, Dhaka.

## Appendix III. The physical and chemical characteristics of soil of the experimental site as observed prior to experimentation (0-15 cm depth)

| Constituents   | Percent    |
|----------------|------------|
| Sand           | 26         |
| Silt           | 45         |
| Clay           | 29         |
| Textural class | Silty clay |

## **Chemical composition:**

| Soil characters    | Value               |
|--------------------|---------------------|
| Organic carbon (%) | 0.45                |
| Organic matter (%) | 0.54                |
| Total nitrogen (%) | 0.027               |
| Phosphorus         | 6.3 μg/g soil       |
| Sulphur            | 8.42 μg/g soil      |
| Magnesium          | 1.17 meq/100 g soil |
| Boron              | 0.88 µg/g soil      |
| Copper             | 1.64 µg/g soil      |
| Zinc               | 1.54 µg/g soil      |
| Potassium          | 0.10 meg/100g soil  |

## Source: Soil Resources Development Institute (SRDI), Khamarbari, Dhaka

**APPENDIX IV: Some plates of the study** 



Plate 1: Seedlings on seedbed



Plate 3: Main field of brinjal



Plate 2: Transplanted seedlings of brinjal on poly bag





Plate 5: Data collection

Plate 4: Experimental working at main field of brinjal



Plate 6: Leaf hopper on brinjal leaf